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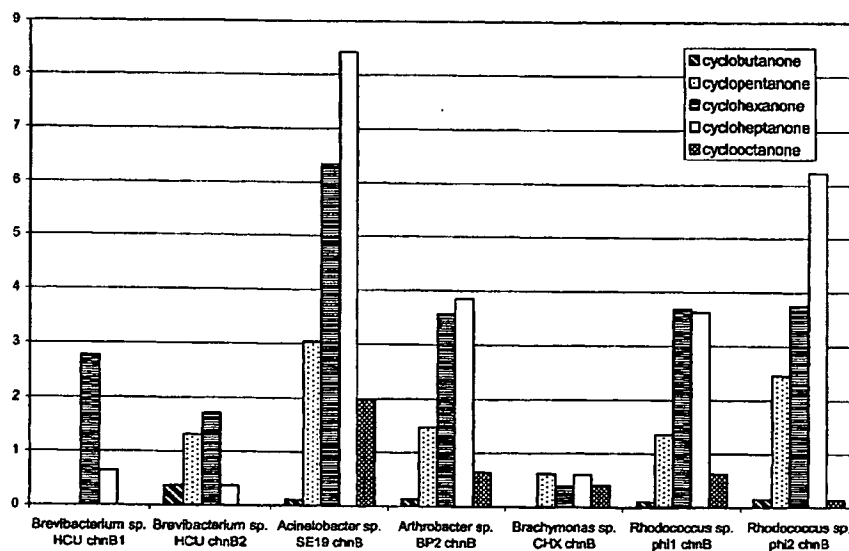
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(54) Title: GENES ENCODING BAEYER-VILLIGER MONOOXYGENASES



(57) Abstract: Genes have been isolated from a variety of bacteria encoding Baeyer-Villiger monooxygenase activity. The genes and their products are useful for the conversion of ketones to the corresponding esters. A series of motifs, common to all genes, has been identified as diagnostic for genes encoding proteins of this activity.



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## TITLE

### GENES ENCODING BAEYER-VILLIGER MONOOXYGENASES

#### FIELD OF THE INVENTION

The invention relates to the field of molecular biology and  
5 microbiology. More specifically, genes have been isolated from a variety  
of bacteria encoding Baeyer-Villiger monooxygenase activity.

#### BACKGROUND OF THE INVENTION

In 1899, Baeyer and Villiger reported on a reaction of cyclic ketones  
with peroxymonosulfuric acid to produce lactones (*Chem Ber*  
10 32:3625-3633 (1899)). Since then, the Baeyer-Villiger (BV) reaction has  
been broadly used in organic synthesis. BV reactions are one of only a  
few methods available for cleaving specific carbon-carbon bonds under  
mild conditions, thereby converting ketones into esters (Walsh and Chen,  
*Angew.Chem.Int.Ed. Engl* 27:333-343 (1988)).

15 In the last several decades, the importance of minimizing  
environmental impact in industrial processes has catalyzed a trend  
whereby alternative methods are replacing established chemical  
techniques. In the arena of Baeyer-Villiger (BV) oxidations, considerable  
interest has focused on discovery of enantioselective versions of the  
20 Baeyer-Villiger oxidation that are not based on peracids. Enzymes, which  
are often enantioselective, are valued alternatives as renewable,  
biodegradable resources.

Many microbial Baeyer-Villiger monooxygenases enzymes (BVMOs  
) , which convert ketones to esters or the corresponding lactones (cyclic  
25 esters) (Stewart, *Curr. Org. Chem.* 2:195-216 (1998), have been identified  
from both bacterial and fungal sources. In general, microbial BV  
reactions are carried out by monooxygenases (EC 1.14.13.x) which use  
O<sub>2</sub> and either NADH or NADPH as a co-reductant. One of the oxygen  
atoms is incorporated into the lactone product between the carbonyl  
30 carbon and the flanking carbon while the other is used to oxidize the  
reduced NADPH producing H<sub>2</sub>O (Banerjee, A. In *Stereosel, Biocatal.*;  
Patel, R.N., Ed.; Marcel Dekker: New York, 2000; Chapter 29,  
pp 867-876). All known BVMOs have a flavin coenzyme which acts in the  
oxidation reaction; the predominant coenzyme form is flavin adenine  
35 dinucleotide cofactor (FAD).

The natural physiological role of most characterized BVMOs is  
degradation of compounds to permit utilization of smaller hydrocarbons  
and/or alcohols as sources of carbon and energy. As a result of this,

BVMOs display remarkably broad substrate acceptance, high enantioselectivities, and great stereoselectivity and regioselectivity (Mihovilovic et al. *J. Org. Chem.* 66:733-738 (2001)). Suitable substrates for the enzymes can be broadly classified as cyclic ketones, ketoterpenes, and steroids. However, few enzymes have been subjected to extensive biochemical characterization. Key studies in relation to each broad ketone substrate class are summarized below.

1. Cyclic ketones: Activity of cyclohexanone monooxygenase upon cyclic ketone substrates in *Acinetobacter* sp. NCIB 9871 has been studied extensively (reviewed in Stewart, *Curr. Org. Chem.* 2:195-216 (1998), Table 2; Walsh and Chen, *Angew.Chem.Int.Ed. Engl* 27:333-343 (1988), Tables 4-5). Specificity has also been biochemically analyzed in *Brevibacterium* sp. HCU (Brzostowicz et al., *J. Bact.* 182(15):4241-4248 (2000)).

2. Ketoterpenes: A monocyclic monoterpene ketone monooxygenase has been characterized from *Rhodococcus erythropolis* DCL14 (Van der Werf, *J. Biochem.* 347:693-701 (2000)). In addition to broad substrate specificity against ketoterpenes, the enzyme also has activity against substituted cyclohexanones.

3. Steroids: The steroid monooxygenase of *Rhodococcus rhodochrous* (Morii et al. *J. Biochem* 126:624-631 (1999)) is well characterized, both biochemically and by sequence data.

The genes and gene products listed above are useful for specific Baeyer-Villiger reactions targeted toward cyclic ketone, ketoterpene, or steroid compounds, however the enzymes are limited in their ability to predict other newly discovered proteins which would have similar activity.

The problem to be solved, therefore is to provide a suite of bacterial flavoprotein Baeyer-Villiger monooxygenase enzymes that can efficiently perform oxygenation reactions on cyclic ketones and ketoterpenes compounds. Identity of a suite of enzymes with this broad substrate acceptance would facilitate commercial applications of these enzymes and reduce efforts with respect to optimization of multiple enzymes for multiple reactions. Maximum efficiency is especially relevant today, when many enzymes are genetically engineered such that the enzyme is recombinantly expressed in a desirable host organism. Additionally, a collection of BVMO's with diverse amino acid sequences could be used to create a general predictive model based on amino acid sequence

conservation of other BVMO enzymes. Finally, a broad class of BVMO's could also be used as basis for the *in vitro* evolution of novel enzymes.

Applicants have solved the stated problem by isolating several novel organisms with BVMO activity, identifying and characterizing BMVO genes, expressing these genes in microbial hosts, and demonstrating activity of the genes against a wide range of ketone substrates, including cyclic ketones and ketoterpenes. Several signature sequences have been identified, based on amino acid sequence alignments, which are characteristic of specific BVMO families and have diagnostic utility.

#### 10 SUMMARY OF THE INVENTION

The invention provides an isolated nucleic acid fragment isolated from *Rhodococcus* selected from the group consisting of:

(a) an isolated nucleic acid fragment encoding a Baeyer-Villiger monooxygenase polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NOs:8, 10, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, and 46.

(b) an isolated nucleic acid molecule encoding a Baeyer-Villiger monooxygenase polypeptide that hybridizes with (a) under the following hybridization conditions: 0.1X SSC, 0.1% SDS, 65°C and washed with 2X SSC, 0.1% SDS followed by 0.1X SSC, 0.1% SDS; or  
an isolated nucleic acid fragment that is complementary to (a) or (b).

Similarly the invention provides an isolated nucleic acid fragment isolated from *Arthrobacter* selected from the group consisting of:

(a) an isolated nucleic acid fragment encoding a Baeyer-Villiger monooxygenase polypeptide having an amino acid sequence as set forth in SEQ ID NO:12;

(b) an isolated nucleic acid molecule encoding a Baeyer-Villiger monooxygenase polypeptide that hybridizes with (a) under the following hybridization conditions: 0.1X SSC, 0.1% SDS, 65°C and washed with 2X SSC, 0.1% SDS followed by 0.1X SSC, 0.1% SDS; or  
an isolated nucleic acid fragment that is complementary to (a), or (b).

Additionally the invention provides an isolated nucleic acid fragment isolated from *Acidovorax* selected from the group consisting of:

(a) an isolated nucleic acid fragment encoding a Baeyer-Villiger monooxygenase polypeptide having an amino acid sequence as set forth in SEQ ID NO:18

(b) an isolated nucleic acid molecule encoding a Baeyer-Villiger monooxygenase polypeptide that hybridizes with (a) under the following hybridization conditions: 0.1X SSC, 0.1% SDS, 65°C and washed with 2X SSC, 0.1% SDS followed by 0.1X SSC, 0.1% SDS; or

5 an isolated nucleic acid fragment that is complementary to (a), or (b).

In additional embodiments the invention provides polypeptides encoded by the present sequences as well as genetic chimera of the present sequences and transformed hosts expressing the same.

10 In a preferred embodiment the invention provides a method for the identification of a polypeptide having monooxygenase activity comprising:

(a) obtaining the amino acid sequence of a polypeptide suspected of having monooxygenase activity; and

(b) aligning the amino acid sequence of step (a) with the amino acid  
15 sequence of a Baeyer-Villiger monooxygenase consensus sequence selected from the group consisting of SEQ ID NO:47, SEQ ID NO:48 and SEQ ID NO:49,

wherein where at least 80% of the amino acid residues at positions p1- p74 of SEQ ID NO:47, or at least 80% of the amino acid residues at  
20 p1-p76 of SEQ ID NO:48 or at least 80% of the amino acid residues of p1-p41 of SEQ ID NO:49 are completely conserved, the polypeptide of (a) is identified as having monooxygenase activity.

In an alternate embodiment the invention provides a method for identifying a gene encoding a Baeyer-Villiger monooxygenase polypeptide  
25 comprising:

(a) probing a genomic library with a nucleic acid fragment encoding a polypeptide wherein where at least 80% of the amino acid residues at positions p1- p74 of SEQ ID NO:47, or at least 80% of the amino acid  
30 residues at p1-p76 of SEQ ID NO:48 or at least 80% of the amino acid residues of p1-p41 of SEQ ID NO:49 are completely conserved;

(b) identifying a DNA clone that hybridizes with a nucleic acid fragment of step (a);

(c) sequencing the genomic fragment that comprises the clone identified in step (b),

35 wherein the sequenced genomic fragment encodes a Baeyer-Villiger monooxygenase polypeptide.

In a preferred embodiment the invention provides a method for the biotransformation of a ketone substrate to the corresponding ester,

comprising: contacting a transformed host cell under suitable growth conditions with an effective amount of ketone substrate whereby the corresponding ester is produced, said transformed host cell comprising a nucleic acid fragment encoding an isolated nucleic acid fragment of any of the present nucleic acid sequences; under the control of suitable regulatory sequences.

In an alternate embodiment the invention provides a method for the *in vitro* transformation of a ketone substrate to the corresponding ester, comprising: contacting a ketone substrate under suitable reaction conditions with an effective amount of a Baeyer-Villiger monooxygenase enzyme, the enzyme having an amino acid sequence selected from the group consisting of SEQ ID NOs:8, 10, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, and 46.

Additionally the invention provides a mutated microbial gene encoding a protein having an altered biological activity produced by a method comprising the steps of:

(i) digesting a mixture of nucleotide sequences with restriction endonucleases wherein said mixture comprises:

a) a native microbial gene selected from the group consisting of SEQ ID NOs:7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, and 45;

b) a first population of nucleotide fragments which will hybridize to said native microbial sequence;

c) a second population of nucleotide fragments which will not hybridize to said native microbial sequence;

wherein a mixture of restriction fragments are produced;

(ii) denaturing said mixture of restriction fragments;

(iii) incubating the denatured said mixture of restriction fragments of step (ii) with a polymerase;

(iv) repeating steps (ii) and (iii) wherein a mutated microbial gene is produced encoding a protein having an altered biological activity.

Additionally the invention provides unique strains of *Acidovorax* sp. comprising the 16s rDNA sequence as set forth in SEQ ID NO:5, *Arthrobacter* sp. comprising the 16s rDNA sequence as set forth in SEQ ID NO:1, and *Rhodococcus* sp. comprising the 16s rDNA sequence as set forth in SEQ ID NO:6.

In another embodiment the invention provides an *Acidovorax* sp. comprising the 16s rDNA sequence as set forth in SEQ ID NO:5.

Additionally the invention provides an *Arthrobacter* sp. comprising the 16s rDNA sequence as set forth in SEQ ID NO:1. Similarly the invention provides a *Rhodococcus* sp. comprising the 16s rDNA sequence as set forth in SEQ ID NO:6.

- 5        Additionally the invention provides an isolated nucleic acid useful for the identification of a BV monooxygenase selected from the group consisting of SEQ ID 70-113.

#### BRIEF DESCRIPTION OF THE DRAWINGS,

#### AND SEQUENCE DESCRIPTIONS

- 10        Figures 1, 2, 3, 4, and 5 show *chnB* monooxygenase activity of *Brevibacterium* sp. HCU, *Acinetobacter* SE19, *Rhodococcus* sp. phi1, *Rhodococcus* sp. phi2, *Arthrobacter* sp. BP2 and *Acidovorax* sp. CHX genes over-expressed in *E. coli* assayed against various ketone substrates.

- 15        Figure 6 illustrates the signature sequences of the three BVMO groups based on the consensus sequences derived from the alignments of Figure 7, Figure 8 and Figure 9.

Figure 7 shows a Clustal W alignment of a family of Baeyer-Villiger monooxygenases (Family 1) and the associated signature sequence.

- 20        Figure 8 shows a Clustal W alignment of a family of Baeyer-Villiger monooxygenases (Family 2) and the associated signature sequence.

Figure 9 shows a Clustal W alignment of a family of BC monooxygenases (Family 3) and the associated signature sequence.

- 25        The invention can be more fully understood from the following detailed description and the accompanying sequence descriptions which form a part of this application.

- 30        The following sequences conform with 37 C.F.R. 1.821-1.825 ("Requirements for Patent Applications Containing Nucleotide Sequences and/or Amino Acid Sequence Disclosures - the Sequence Rules") and consistent with World Intellectual Property Organization (WIPO) Standard ST.25 (1998) and the sequence listing requirements of the EPO and PCT (Rules 5.2 and 49.5(a-bis), and Section 208 and Annex C of the Administrative Instructions). The symbols and format used for nucleotide and amino acid sequence data comply with the rules set forth in
- 35        37 C.F.R. §1.822.

SEQ ID NOs:1-49 are full length genes or proteins as identified in Table 1.

#### Table 1



## Summary of Gene and Protein SEQ ID Numbers

Gene Name	Organism	Gene SEQ ID No	Protein SEQ ID No
16s rDNA sequence	<i>Arthrobacter sp. BP2</i>	1	--
16s rDNA sequence	<i>Rhodococcus sp. phi1</i>	2	--
16s rDNA sequence	<i>Rhodococcus sp. phi2</i>	3	--
16s rDNA sequence	<i>Brevibacterium sp. HCU</i>	4	--
16s rDNA sequence	<i>Acidovorax sp. CHX</i>	5	--
16s rDNA sequence	<i>Rhodococcus erythropolis AN12</i>	6	--
<i>chnB</i> Monooxygenase phi1	<i>Rhodococcus sp. phi1</i>	7	8
<i>chnB</i> Monooxygenase phi2	<i>Rhodococcus sp. phi2</i>	9	10
<i>chnB</i> Monooxygenase BP2	<i>Arthrobacter sp. BP2</i>	11	12
<i>chnB1</i> Monooxygenase HCU #1	<i>Brevibacterium sp. HCU</i>	13	14
<i>chnB2</i> Monooxygenase HCU #2	<i>Brevibacterium sp. HCU</i>	15	16
<i>chnB</i> Monooxygenase CHX	<i>Acidovorax sp. CHX</i>	17	18
<i>chnB</i> Monooxygenase SE19	<i>Acinetobacter sp. SE19</i>	19	20
ORF 8 <i>chnB</i> Monooxygenase (1413)	<i>Rhodococcus erythropolis AN12</i>	21	22
ORF 9 <i>chnB</i> Monooxygenase (1985)	<i>Rhodococcus erythropolis AN12</i>	23	24
ORF 10 <i>chnB</i> Monooxygenase (1273)	<i>Rhodococcus erythropolis AN12</i>	25	26
ORF 11 <i>chnB</i> Monooxygenase (2034)	<i>Rhodococcus erythropolis AN12</i>	27	28
ORF 12 <i>chnB</i> Monooxygenase (1870)	<i>Rhodococcus erythropolis AN12</i>	29	30
ORF 13 <i>chnB</i> Monooxygenase (1861)	<i>Rhodococcus erythropolis AN12</i>	31	32
ORF 14 <i>chnB</i>	<i>Rhodococcus</i>	33	34

Gene Name	Organism	Gene SEQ ID No	Protein SEQ ID No
Monooxygenase (2005)	<i>erythropolis AN12</i>		
ORF 15 <i>chnB</i> Monooxygenase (2035)	<i>Rhodococcus</i> <i>erythropolis AN12</i>	35	36
ORF 16 <i>chnB</i> Monooxygenase (2022)	<i>Rhodococcus</i> <i>erythropolis AN12</i>	37	38
ORF 17 <i>chnB</i> Monooxygenase (1976)	<i>Rhodococcus</i> <i>erythropolis AN12</i>	39	40
ORF 18 <i>chnB</i> Monooxygenase (1294)	<i>Rhodococcus</i> <i>erythropolis AN12</i>	41	42
ORF 19 <i>chnB</i> Monooxygenase (2082)	<i>Rhodococcus</i> <i>erythropolis AN12</i>	43	44
ORF 20 <i>chnB</i> Monooxygenase (2093)	<i>Rhodococcus</i> <i>erythropolis AN12</i>	45	46
Signature Sequence #1	Consensus Sequence	—	47
Signature Sequence #2	Consensus Sequence	—	48
Signature Sequence #3	Consensus Sequence	—	49

SEQ ID NOs:50-62 are primers used for 16s rDNA sequencing.

SEQ ID NO:63 describes a primer used for RT-PCR and out-PCR.

SEQ ID NOs:64 and 65 are primers used for sequencing of inserts

5 within pCR2.1

SEQ ID NOs:66 and 67 are primers used to amplify monooxygenase genes from *Acinetobacter* sp. SE19.

10 SEQ ID NOs:68-107 are primers used for amplification of full length Baeyer-Villiger monooxygenases.

SEQ ID NOs:108-113 are primers used to screen cosmid libraries.

#### DETAILED DESCRIPTION OF THE INVENTION

15 The invention provides nucleic acid and amino acid sequences defining a group of Baeyer-Villiger monooxygenase enzymes. These enzymes have been found to have the ability to use a wide variety of ketone substrates that include two general classes of compounds, cyclic ketones and ketoterpenes. These enzymes are characterized by function as well as a series of diagnostic signature sequences. The enzymes may

be expressed recombinantly for the conversion of ketone substrates to the corresponding lactones or esters.

In this disclosure, a number of terms and abbreviations are used.

The following definitions are provided.

5 "Open reading frame" is abbreviated ORF.

"Polymerase chain reaction" is abbreviated PCR.

"Gas Chromatography Mass spectrometry" is abbreviated GC-MS.

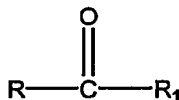
"Baeyer-Villiger" is abbreviated BV.

"Baeyer-Villiger monooxygenase" is abbreviated BVMO.

10 The term "Baeyer-Villiger monooxygenase", refers to a bacterial enzyme that has the ability to oxidize a ketone substrate to the corresponding lactone or ester.

The term "ketone substrate" includes a substrate for a Baeyer-Villiger monooxygenase that comprises a class of compounds which

15 include cyclic ketones and ketoterpenes. Ketone substrates of the invention are defined by the general formula:



20 wherein R and R<sub>1</sub> are independently selected from substituted or unsubstituted phenyl, substituted or unsubstituted alkyl, substituted or unsubstituted alkenyl, or substituted or unsubstituted alkylidene.

The term "alkyl" will mean a univalent group derived from alkanes by removal of a hydrogen atom from any carbon atom: C<sub>n</sub>H<sub>2n+1</sub>-. The groups derived by removal of a hydrogen atom from a terminal carbon atom of unbranched alkanes form a subclass of normal alkyl (*n*-alkyl) groups: H[CH<sub>2</sub>]<sub>n</sub>-. The groups RCH<sub>2</sub>-, R<sub>2</sub>CH- (R not equal to H), and R<sub>3</sub>C- (R not equal to H) are primary, secondary and tertiary alkyl groups respectively.

30 The term "alkenyl" will mean an acyclic branched or unbranched hydrocarbon having one carbon-carbon double bond and the general formula C<sub>n</sub>H<sub>2n</sub>-. Acyclic branched or unbranched hydrocarbons having more than one double bond are alkadienes, alkatrienes, etc.

35 The term "alkylidene" will mean the divalent groups formed from alkanes by removal of two hydrogen atoms from the same carbon atom, the free valiances of which are part of a double bond (e.g. (CH<sub>3</sub>)<sub>2</sub>C, also known as propan-2-ylidene).

As used herein, an "isolated nucleic acid molecule" is a polymer of RNA or DNA that is single- or double-stranded, optionally containing synthetic, non-natural or altered nucleotide bases. An isolated nucleic acid fragment in the form of a polymer of DNA may be comprised of one or more segments of cDNA, genomic DNA or synthetic DNA.

A nucleic acid molecule is "hybridizable" to another nucleic acid molecule, such as a cDNA, genomic DNA, or RNA, when a single stranded form of the nucleic acid molecule can anneal to the other nucleic acid molecule under the appropriate conditions of temperature and solution ionic strength. Hybridization and washing conditions are well known and exemplified in Sambrook, J., Fritsch, E. F. and Maniatis, T. Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor (1989), particularly Chapter 11 and Table 11.1 therein (entirely incorporated herein by reference). The conditions of temperature and ionic strength determine the "stringency" of the hybridization. Stringency conditions can be adjusted to screen for moderately similar fragments, such as homologous sequences from distantly related organisms, to highly similar fragments, such as genes that duplicate functional enzymes from closely related organisms. Typical stringent hybridization conditions are for example, hybridization at 0.1X SSC, 0.1% SDS, 65°C with a wash with 2X SSC, 0.1% SDS followed by 0.1X SSC, 0.1% SDS. Generally post-hybridization washes determine stringency conditions. One set of preferred conditions uses a series of washes starting with 6X SSC, 0.5% SDS at room temperature for 15 min, then repeated with 2X SSC, 0.5% SDS at 45°C for 30 min, and then repeated twice with 0.2X SSC, 0.5% SDS at 50°C for 30 min. A more preferred set of stringent conditions uses higher temperatures in which the washes are identical to those above except for the temperature of the final two 30 min washes in 0.2X SSC, 0.5% SDS was increased to 60°C. Another preferred set of highly stringent conditions uses two final washes in 0.1X SSC, 0.1% SDS at 65°C. Hybridization requires that the two nucleic acids contain complementary sequences, although depending on the stringency of the hybridization, mismatches between bases are possible. The appropriate stringency for hybridizing nucleic acids depends on the length of the nucleic acids and the degree of complementation, variables well known in the art. The greater the degree of similarity or homology between two nucleotide sequences, the greater the value of  $T_m$  for hybrids of nucleic acids having

those sequences. The relative stability (corresponding to higher  $T_m$ ) of nucleic acid hybridizations decreases in the following order: RNA:RNA, DNA:RNA, DNA:DNA. For hybrids of greater than 100 nucleotides in length, equations for calculating  $T_m$  have been derived (see Sambrook *et al.*, *supra*, 9.50-9.51). For hybridizations with shorter nucleic acids, i.e., oligonucleotides, the position of mismatches becomes more important, and the length of the oligonucleotide determines its specificity (see Sambrook *et al.*, *supra*, 11.7-11.8). In one embodiment the length for a hybridizable nucleic acid is at least about 10 nucleotides. Preferable a minimum length for a hybridizable nucleic acid is at least about 15 nucleotides; more preferably at least about 20 nucleotides; and most preferably the length is at least 30 nucleotides. Furthermore, the skilled artisan will recognize that the temperature and wash solution salt concentration may be adjusted as necessary according to factors such as length of the probe.

The term "complementary" is used to describe the relationship between nucleotide bases that are capable to hybridizing to one another. For example, with respect to DNA, adenosine is complementary to thymine and cytosine is complementary to guanine. Accordingly, the instant invention also includes isolated nucleic acid fragments that are complementary to the complete sequences as reported in the accompanying Sequence Listing as well as those substantially similar nucleic acid sequences.

The term "percent identity", as known in the art, is a relationship between two or more polypeptide sequences or two or more polynucleotide sequences, as determined by comparing the sequences. In the art, "identity" also means the degree of sequence relatedness between polypeptide or polynucleotide sequences, as the case may be, as determined by the match between strings of such sequences. "Identity" and "similarity" can be readily calculated by known methods, including but not limited to those described in: Computational Molecular Biology (Lesk, A. M., ed.) Oxford University Press, New York (1988); Biocomputing: Informatics and Genome Projects (Smith, D. W., ed.) Academic Press, New York (1993); Computer Analysis of Sequence Data, Part I (Griffin, A. M., and Griffin, H. G., eds.) Humana Press, New Jersey (1994); Sequence Analysis in Molecular Biology (von Heinje, G., ed.) Academic Press (1987); and Sequence Analysis Primer (Gribskov, M. and Devereux, J., eds.) Stockton Press, New York (1991). Preferred methods to determine

identity are designed to give the best match between the sequences tested. Methods to determine identity and similarity are codified in publicly available computer programs. Sequence alignments and percent identity calculations may be performed using the Megalign program of the  
5 LASERGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI). Multiple alignment of the sequences was performed using the Clustal method of alignment (Higgins and Sharp (1989) *CABIOS*. 5:151-153) with the default parameters (GAP PENALTY=10, GAP LENGTH  
10 PENALTY=10). Default parameters for pairwise alignments using the Clustal method were KTUPLE 1, GAP PENALTY=3, WINDOW=5 and DIAGONALS SAVED=5.

Suitable nucleic acid fragments (isolated polynucleotides of the present invention) encode polypeptides that are at least about 70% identical, preferably at least about 80% identical to the amino acid  
15 sequences reported herein. Preferred nucleic acid fragments encode amino acid sequences that are about 85% identical to the amino acid sequences reported herein. More preferred nucleic acid fragments encode amino acid sequences that are at least about 90% identical to the amino acid sequences reported herein. Most preferred are nucleic acid  
20 fragments that encode amino acid sequences that are at least about 95% identical to the amino acid sequences reported herein. Suitable nucleic acid fragments not only have the above homologies but typically encode a polypeptide having at least 50 amino acids, preferably at least 100 amino acids, more preferably at least 150 amino acids, still more preferably at  
25 least 200 amino acids, and most preferably at least 250 amino acids.

"Codon degeneracy" refers to the nature in the genetic code permitting variation of the nucleotide sequence without effecting the amino acid sequence of an encoded polypeptide. Accordingly, the instant invention relates to any nucleic acid fragment that encodes all or a  
30 substantial portion of the amino acid sequence encoding the instant microbial polypeptides as set forth in SEQ ID NOs:8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, and 46. The skilled artisan is well aware of the "codon-bias" exhibited by a specific host cell in usage of nucleotide codons to specify a given amino acid. Therefore,  
35 when synthesizing a gene for improved expression in a host cell, it is desirable to design the gene such that its frequency of codon usage approaches the frequency of preferred codon usage of the host cell.

"Synthetic genes" can be assembled from oligonucleotide building blocks that are chemically synthesized using procedures known to those skilled in the art. These building blocks are ligated and annealed to form gene segments which are then enzymatically assembled to construct the entire gene. "Chemically synthesized", as related to a sequence of DNA, means that the component nucleotides were assembled *in vitro*. Manual chemical synthesis of DNA may be accomplished using well established procedures, or automated chemical synthesis can be performed using one of a number of commercially available machines. Accordingly, the genes can be tailored for optimal gene expression based on optimization of nucleotide sequence to reflect the codon bias of the host cell. The skilled artisan appreciates the likelihood of successful gene expression if codon usage is biased towards those codons favored by the host. Determination of preferred codons can be based on a survey of genes derived from the host cell where sequence information is available.

"Gene" refers to a nucleic acid fragment that expresses a specific protein, including regulatory sequences preceding (5' non-coding sequences) and following (3' non-coding sequences) the coding sequence. "Native gene" refers to a gene as found in nature with its own regulatory sequences. "Chimeric gene" refers to any gene that is not a native gene, comprising regulatory and coding sequences that are not found together in nature. Accordingly, a chimeric gene may comprise regulatory sequences and coding sequences that are derived from different sources, or regulatory sequences and coding sequences derived from the same source, but arranged in a manner different than that found in nature. "Endogenous gene" refers to a native gene in its natural location in the genome of an organism. A "foreign" gene refers to a gene not normally found in the host organism, but that is introduced into the host organism by gene transfer. Foreign genes can comprise native genes inserted into a non-native organism, or chimeric genes. A "transgene" is a gene that has been introduced into the genome by a transformation procedure.

"Coding sequence" refers to a DNA sequence that codes for a specific amino acid sequence. "Suitable regulatory sequences" refer to nucleotide sequences located upstream (5' non-coding sequences), within, or downstream (3' non-coding sequences) of a coding sequence, and which influence the transcription, RNA processing or stability, or translation of the associated coding sequence. Regulatory sequences

may include promoters, translation leader sequences, introns, polyadenylation recognition sequences, RNA processing site, effector binding site and stem-loop structures.

“Promoter” refers to a DNA sequence capable of controlling the expression of a coding sequence or functional RNA. In general, a coding sequence is located 3' to a promoter sequence. Promoters may be derived in their entirety from a native gene, or be composed of different elements derived from different promoters found in nature, or even comprise synthetic DNA segments. It is understood by those skilled in the art that different promoters may direct the expression of a gene in different tissues or cell types, or at different stages of development, or in response to different environmental or physiological conditions. Promoters which cause a gene to be expressed in most cell types at most times are commonly referred to as “constitutive promoters”. It is further recognized that since in most cases the exact boundaries of regulatory sequences have not been completely defined, DNA fragments of different lengths may have identical promoter activity.

The “3' non-coding sequences” refer to DNA sequences located downstream of a coding sequence and include polyadenylation recognition sequences and other sequences encoding regulatory signals capable of affecting mRNA processing or gene expression. The polyadenylation signal is usually characterized by affecting the addition of polyadenylic acid tracts to the 3' end of the mRNA precursor.

“RNA transcript” refers to the product resulting from RNA polymerase-catalyzed transcription of a DNA sequence. When the RNA transcript is a perfect complementary copy of the DNA sequence, it is referred to as the primary transcript or it may be a RNA sequence derived from post-transcriptional processing of the primary transcript and is referred to as the mature RNA. “Messenger RNA (mRNA)” refers to the RNA that is without introns and that can be translated into protein by the cell. “cDNA” refers to a double-stranded DNA that is complementary to and derived from mRNA. “Sense” RNA refers to RNA transcript that includes the mRNA and so can be translated into protein by the cell. “Antisense RNA” refers to a RNA transcript that is complementary to all or part of a target primary transcript or mRNA and that blocks the expression of a target gene (U.S. Patent No. 5,107,065; WO 9928508). The complementarity of an antisense RNA may be with any part of the specific gene transcript, i.e., at the 5' non-coding sequence, 3' non-coding



sequence, or the coding sequence. "Functional RNA" refers to antisense RNA, ribozyme RNA, or other RNA that is not translated yet has an effect on cellular processes.

5 The term "operably linked" refers to the association of nucleic acid sequences on a single nucleic acid fragment so that the function of one is affected by the other. For example, a promoter is operably linked with a coding sequence when it is capable of affecting the expression of that coding sequence (i.e., that the coding sequence is under the transcriptional control of the promoter). Coding sequences can be  
10 operably linked to regulatory sequences in sense or antisense orientation.

The term "expression", as used herein, refers to the transcription and stable accumulation of sense (mRNA) or antisense RNA derived from the nucleic acid fragment of the invention. Expression may also refer to translation of mRNA into a polypeptide.

15 "Transformation" refers to the transfer of a nucleic acid fragment into the genome of a host organism, resulting in genetically stable inheritance. Host organisms containing the transformed nucleic acid fragments are referred to as "transgenic" or "recombinant" or "transformed" organisms.

20 The terms "plasmid", "vector" and "cassette" refer to an extra chromosomal element often carrying genes which are not part of the central metabolism of the cell, and usually in the form of circular double-stranded DNA molecules. Such elements may be autonomously replicating sequences, genome integrating sequences, phage or  
25 nucleotide sequences, linear or circular, of a single- or double-stranded DNA or RNA, derived from any source, in which a number of nucleotide sequences have been joined or recombined into a unique construction which is capable of introducing a promoter fragment and DNA sequence for a selected gene product along with appropriate 3' untranslated  
30 sequence into a cell. "Transformation cassette" refers to a specific vector containing a foreign gene and having elements in addition to the foreign gene that facilitate transformation of a particular host cell. "Expression cassette" refers to a specific vector containing a foreign gene and having elements in addition to the foreign gene that allow for enhanced  
35 expression of that gene in a foreign host.

The term "sequence analysis software" refers to any computer algorithm or software program that is useful for the analysis of nucleotide or amino acid sequences. "Sequence analysis software" may be

commercially available or independently developed. Typical sequence analysis software will include but is not limited to the GCG suite of programs (Wisconsin Package Version 9.0, Genetics Computer Group (GCG), Madison, WI), BLASTP, BLASTN, BLASTX (Altschul *et al.*, *J. Mol. Biol.* 215:403-410 (1990), and DNASTAR (DNASTAR, Inc. 1228 S. Park St. Madison, WI 53715 USA), and the FASTA program incorporating the Smith-Waterman algorithm (W. R. Pearson, *Comput. Methods Genome Res.*, [Proc. Int. Symp.] (1994), Meeting Date 1992, 111-20. Editor(s): Suhai, Sandor. Publisher: Plenum, New York, NY). Within the context of this application it will be understood that where sequence analysis software is used for analysis, that the results of the analysis will be based on the "default values" of the program referenced, unless otherwise specified. As used herein "default values" will mean any set of values or parameters which originally load with the software when first initialized.

The term "signature sequence" means a set of amino acids conserved at specific positions along an aligned sequence of evolutionarily related proteins. While amino acids at other positions can vary between homologous proteins, amino acids which are highly conserved at specific positions indicate amino acids which are essential in the structure, the stability, or the activity of a protein. Because they are identified by their high degree of conservation in aligned sequences of a family of protein homologues, they can be used as identifiers, or "signatures", to determine if a protein with a newly determined sequence belongs to a previously identified protein family. Signature sequences of the present invention are specifically described Figure 6 showing the signature sequence comprised of p1-p74 of SEQ ID NO:47, p1-p76 of SEQ ID NO:48 and p1-p41 of SEQ ID NO:49.

Standard recombinant DNA and molecular cloning techniques used here are well known in the art and are described by Sambrook, J., Fritsch, E. F. and Maniatis, T., Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (1989) (hereinafter "Maniatis"); and by Silhavy, T. J., Bennis, M. L. and Enquist, L. W., Experiments with Gene Fusions, Cold Spring Harbor Laboratory Cold Press Spring Harbor, NY (1984); and by Ausubel, F. M. *et al.*, Current Protocols in Molecular Biology, published by Greene Publishing Assoc. and Wiley-Interscience (1987).

### Isolation Of Microorganisms Having Baeyer-Villiger Monooxygenase Activity

Microorganisms having Baeyer-Villiger monooxygenase activity may be isolated from a variety of sources. Suitable sources include industrial waste streams, soil from contaminated industrial sites and waste stream treatment facilities. The Baeyer-Villiger monooxygenase containing microorganisms of the instant invention were isolated from activated sludge from waste water treatment plants.

Samples suspected of containing a microorganism having Baeyer-Villiger monooxygenase activity may be enriched by incubation in a suitable growth medium in combination with at least one ketone substrate. Suitable ketone substrates for use in the instant invention include cyclic ketones and ketoterpenes having the general formula:



wherein R and R<sub>1</sub> are independently selected from substituted or unsubstituted phenyl, substituted or unsubstituted alkyl, or substituted or unsubstituted alkenyl or substituted or unsubstituted alkylidene. These compounds may be synthetic or natural secondary metabolites

Particularly useful ketone substrates include, but are not limited to Norcamphor, Cyclobutanone, Cyclopentanone, 2-methyl-cyclopentanone, Cyclohexanone, 2-methyl-cyclohexanone, Cyclohex-2-ene-1-one, 1,2-cyclohexanedione, 1,3-cyclohexanedione, 1,4-cyclohexanedione, Cycloheptanone, Cyclooctanone, Cyclodecanone, Cycloundecanone, Cyclododecanone, Cyclotridecanone, Cyclopenta-decanone, 2-tridecanone, dihexyl ketone, 2-phenyl-cyclohexanone, Oxindole, Levoglucosenone, dimethyl sulfoxide, dimethy-2-piperidone, Phenylboronic acid, and beta-ionone. Growth medium and techniques needed in the enrichment and screening of microorganisms are well known in the art and examples may be found in Manual of Methods for General Bacteriology (Phillipp Gerhardt, R. G. E. Murray, Ralph N. Costilow, Eugene W. Nester, Willis A. Wood, Noel R. Krieg and G. Briggs Phillips, eds), American Society for Microbiology, Washington, DC. (1994)); or by Thomas D. Brock in Biotechnology: A Textbook of Industrial Microbiology, Second Edition, Sinauer Associates, Inc., Sunderland, MA (1989).

Characterization of the Baeyer-Villiger Monooxygenase Containing Microorganisms:

The sequence of the small subunit ribosomal RNA or DNA (16S rDNA) is frequently used for taxonomic identification of novel bacterial. Currently, more than 7,000 bacterial 16S rDNA sequences are now available. Highly conserved regions of the 16S rDNA provide priming sites for broad-range polymerase chain reaction (PCR) (or RT-PCR) and obviate the need for specific information about a targeted microorganism before this procedure. This permits identification of a previously uncharacterized bacterium by broad range bacterial 16S rDNA amplification, sequencing, and phylogenetic analysis.

This invention describes the isolation and identification of 7 different bacteria based on their taxonomic identification following amplification of the 16S rDNA using primers corresponding to conserved regions of the 16S rDNA molecule (Amann, R.I. et al. *Microbiol. Rev.* 59(1):143-69 (1995); Kane, M.D. et al. *Appl. Environ. Microbiol.* 59:682-686 (1993)), followed by sequencing and BLAST analysis (Basic Local Alignment Search Tool; Altschul, S. F., et al., *J. Mol. Biol.* 215:403-410 (1993); see also [www.ncbi.nlm.nih.gov/BLAST/](http://www.ncbi.nlm.nih.gov/BLAST/)). Bacterial strains were identified as highly homologous to bacteria of the genera *Brevibacterium*, *Arthrobacter*, *Acinetobacter*, *Acidovorax*, and *Rhodococcus*.

Comparison of the 16S rRNA nucleotide base sequence from strain AN12 to public databases reveals that the most similar known sequences (98% homologous) are the 16S rRNA gene sequences of bacteria belonging to the genus *Rhodococcus*.

Comparison of the 16S rRNA nucleotide base sequence from strain CHX to public databases reveals that the most similar known sequences (97% homologous) are the 16S rRNA gene sequences of bacteria of the genus *Acidovorax*.

Comparison of the 16S rRNA nucleotide base sequence from strain BP2 to public databases reveals that the most similar known sequences (99% homologous) are the 16S rRNA gene sequences of bacteria of the genus *Arthrobacter*. Comparison of the 16S rRNA nucleotide base sequence from strain SE19 to public databases reveals that the most similar known sequences (99% homologous) are the 16S rRNA gene sequences of bacteria of the genus *Acinetobacter*.

Comparison of the 16S rRNA nucleotide base sequence from strains phi1 and phi2 to public databases reveals that the most similar

known sequences (99% homologous) are the 16S rRNA gene sequences of bacteria belonging to the genus *Rhodococcus*.

Identification of Baeyer-Villiger Monooxygenase Homologs

The present invention provides examples of Baeyer-Villiger  
5 monooxygenase genes and gene products having the ability to convert  
suitable ketone substrates comprising cyclic ketones and ketoterpenes to  
the corresponding lactone or ester. For example, genes encoding  
BVMO's have been isolated from *Arthrobacter* (SEQ ID NO:11),  
*Brevibacterium* (SEQ ID NOs:13 and 15), *Acidovorax* (SEQ ID NO:17),  
10 *Acinetobacter* (SEQ ID NO:19), and *Rhodococcus* (SEQ ID NOs:7, 9, 21,  
23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, and 45).

Comparison of the *Arthrobacter* sp. BP2 *chnB* nucleotide base and  
deduced amino acid sequences to public databases reveals that the most  
similar known sequences range from a distant as about 57% identical to  
15 the amino acid sequence of reported herein over length of 532 amino  
acids using a Smith-Waterman alignment algorithm (W. R. Pearson,  
*supra*). Preferred amino acid fragments are at least about 70% - 80% and  
more preferred amino acid fragments are at least about 80%-90%  
identical to the sequences herein. Most preferred are nucleic acid  
20 fragments that are at least 95% identical to the amino acid fragments  
reported herein. Similarly, preferred *chnB* encoding nucleic acid  
sequences corresponding to the instant ORF's are those encoding active  
proteins and which are at least 80% identical to the nucleic acid  
sequences reported herein. More preferred *chnB* nucleic acid fragments  
25 are at least 90% identical to the sequences herein. Most preferred are  
*chnB* nucleic acid fragments that are at least 95% identical to the nucleic  
acid fragments reported herein.

Comparison of the *Acidovorax* sp. CHX *chnB* nucleotide base and  
deduced amino acid sequences to public databases reveals that the most  
similar known sequences range from a distant as about 57% identical to  
30 the amino acid sequence of reported herein over length of 538 amino  
acids using a Smith-Waterman alignment algorithm (W. R. Pearson,  
*supra*). Preferred amino acid fragments are at least about 70% - 80% and  
more preferred amino acid fragments are at least about 80%-90%  
identical to the sequences herein. Most preferred are nucleic acid  
35 fragments that are at least 95% identical to the amino acid fragments  
reported herein. Similarly, preferred *chnB* encoding nucleic acid  
sequences corresponding to the instant ORF's are those encoding active

proteins and which are at least 80% identical to the nucleic acid sequences reported herein. More preferred *chnB* nucleic acid fragments are at least 90% identical to the sequences herein. Most preferred are *chnB* nucleic acid fragments that are at least 95% identical to the nucleic acid fragments reported herein.

Comparison of the *Rhodococcus sp. phi1 chnB* nucleotide base and deduced amino acid sequences to public databases reveals that the most similar known sequences range from a distant as about 55% identical to the amino acid sequence of reported herein over length of 542 amino acids using a Smith-Waterman alignment algorithm (W. R. Pearson, *supra*). Preferred amino acid fragments are at least about 70% - 80% and more preferred amino acid fragments are at least about 80%-90% identical to the sequences herein. Most preferred are nucleic acid fragments that are at least 95% identical to the amino acid fragments reported herein. Similarly, preferred *chnB* encoding nucleic acid sequences corresponding to the instant ORF's are those encoding active proteins and which are at least 80% identical to the nucleic acid sequences reported herein. More preferred *chnB* nucleic acid fragments are at least 90% identical to the sequences herein. Most preferred are *chnB* nucleic acid fragments that are at least 95% identical to the nucleic acid fragments reported herein.

Comparison of the *Rhodococcus sp. phi2 chnB* nucleotide base and deduced amino acid sequences to public databases reveals that the most similar known sequences range from a distant as about 53% identical to the amino acid sequence of reported herein over length of 541 amino acids using a Smith-Waterman alignment algorithm (W. R. Pearson, *supra*). Preferred amino acid fragments are at least about 70% - 80% and more preferred amino acid fragments are at least about 80%-90% identical to the sequences herein. Most preferred are nucleic acid fragments that are at least 95% identical to the amino acid fragments reported herein. Similarly, preferred *chnB* encoding nucleic acid sequences corresponding to the instant ORF's are those encoding active proteins and which are at least 80% identical to the nucleic acid sequences reported herein. More preferred *chnB* nucleic acid fragments are at least 90% identical to the sequences herein. Most preferred are *chnB* nucleic acid fragments that are at least 95% identical to the nucleic acid fragments reported herein.

Comparison of the *Rhodococcus erythropolis* AN12 ORF8 *chnB* nucleotide base and deduced amino acid sequences to public databases reveals that the most similar known sequences range from a distant as about 37% identical to the amino acid sequence of reported herein over  
5 length of 439 amino acids using a Smith-Waterman alignment algorithm (W. R. Pearson, *supra*). Preferred amino acid fragments are at least about 70% - 80% and more preferred amino acid fragments are at least about 80%-90% identical to the sequences herein. Most preferred are nucleic acid fragments that are at least 95% identical to the amino acid fragments  
10 reported herein. Similarly, preferred *chnB* encoding nucleic acid sequences corresponding to the instant ORF's are those encoding active proteins and which are at least 80% identical to the nucleic acid sequences reported herein. More preferred *chnB* nucleic acid fragments are at least 90% identical to the sequences herein. Most preferred are  
15 *chnB* nucleic acid fragments that are at least 95% identical to the nucleic acid fragments reported herein.

Comparison of the *Rhodococcus erythropolis* AN1 ORF9 *chnB* nucleotide base and deduced amino acid sequences to public databases reveals that the most similar known sequences range from a distant as  
20 about 44% identical to the amino acid sequence of reported herein over length of 518 amino acids using a Smith-Waterman alignment algorithm (W. R. Pearson, *supra*). Preferred amino acid fragments are at least about 70% - 80% and more preferred amino acid fragments are at least about 80%-90% identical to the sequences herein. Most preferred are nucleic  
25 acid fragments that are at least 95% identical to the amino acid fragments reported herein. Similarly, preferred *chnB* encoding nucleic acid sequences corresponding to the instant ORF's are those encoding active proteins and which are at least 80% identical to the nucleic acid sequences reported herein. More preferred *chnB* nucleic acid fragments  
30 are at least 90% identical to the sequences herein. Most preferred are *chnB* nucleic acid fragments that are at least 95% identical to the nucleic acid fragments reported herein.

Comparison of the *Rhodococcus erythropolis* AN1 ORF10 *chnB* nucleotide base and deduced amino acid sequences to public databases  
35 reveals that the most similar known sequences range from a distant as about 64% identical to the amino acid sequence of reported herein over length of 541 amino acids using a Smith-Waterman alignment algorithm. (W. R. Pearson, *supra*). Preferred amino acid fragments are at least about

70% - 80% and more preferred amino acid fragments are at least about 80%-90% identical to the sequences herein. Most preferred are nucleic acid fragments that are at least 95% identical to the amino acid fragments reported herein. Similarly, preferred *chnB* encoding nucleic acid sequences corresponding to the instant ORF's are those encoding active proteins and which are at least 80% identical to the nucleic acid sequences reported herein. More preferred *chnB* nucleic acid fragments are at least 90% identical to the sequences herein. Most preferred are *chnB* nucleic acid fragments that are at least 95% identical to the nucleic acid fragments reported herein.

Comparison of the *Rhodococcus erythropolis* AN1 ORF11 *chnB* nucleotide base and deduced amino acid sequences to public databases reveals that the most similar known sequences range from a distant as about 65% identical to the amino acid sequence of reported herein over length of 462 amino acids using a Smith-Waterman alignment algorithm (W. R. Pearson, *supra*). Preferred amino acid fragments are at least about 70% - 80% and more preferred amino acid fragments are at least about 80%-90% identical to the sequences herein. Most preferred are nucleic acid fragments that are at least 95% identical to the amino acid fragments reported herein. Similarly, preferred *chnB* encoding nucleic acid sequences corresponding to the instant ORF's are those encoding active proteins and which are at least 80% identical to the nucleic acid sequences reported herein. More preferred *chnB* nucleic acid fragments are at least 90% identical to the sequences herein. Most preferred are *chnB* nucleic acid fragments that are at least 95% identical to the nucleic acid fragments reported herein.

Comparison of the *Rhodococcus erythropolis* AN1 ORF12 *chnB* nucleotide base and deduced amino acid sequences to public databases reveals that the most similar known sequences range from a distant as about 45% identical to the amino acid sequence of reported herein over length of 523 amino acids using a Smith-Waterman alignment algorithm (W. R. Pearson, *supra*). Preferred amino acid fragments are at least about 70% - 80% and more preferred amino acid fragments are at least about 80%-90% identical to the sequences herein. Most preferred are nucleic acid fragments that are at least 95% identical to the amino acid fragments reported herein. Similarly, preferred *chnB* encoding nucleic acid sequences corresponding to the instant ORF's are those encoding active proteins and which are at least 80% identical to the nucleic acid



sequences reported herein. More preferred *chnB* nucleic acid fragments are at least 90% identical to the sequences herein. Most preferred are *chnB* nucleic acid fragments that are at least 95% identical to the nucleic acid fragments reported herein.

5           Comparison of the *Rhodococcus erythropolis* AN1 ORF13 *chnB* nucleotide base and deduced amino acid sequences to public databases reveals that the most similar known sequences range from a distant as about 55% identical to the amino acid sequence of reported herein over length of 493 amino acids using a Smith-Waterman alignment algorithm  
10 (W. R. Pearson, *supra*). Preferred amino acid fragments are at least about 70% - 80% and more preferred amino acid fragments are at least about 80%-90% identical to the sequences herein. Most preferred are nucleic acid fragments that are at least 95% identical to the amino acid fragments reported herein. Similarly, preferred *chnB* encoding nucleic acid  
15 sequences corresponding to the instant ORF's are those encoding active proteins and which are at least 80% identical to the nucleic acid sequences reported herein. More preferred *chnB* nucleic acid fragments are at least 90% identical to the sequences herein. Most preferred are *chnB* nucleic acid fragments that are at least 95% identical to the nucleic  
20 acid fragments reported herein.

          Comparison of the *Rhodococcus erythropolis* AN1 ORF14 *chnB* nucleotide base and deduced amino acid sequences to public databases reveals that the most similar known sequences range from a distant as about 51% identical to the amino acid sequence of reported herein over  
25 length of 539 amino acids using a Smith-Waterman alignment algorithm (W. R. Pearson, *supra*). Preferred amino acid fragments are at least about 70% - 80% and more preferred amino acid fragments are at least about 80%-90% identical to the sequences herein. Most preferred are nucleic acid fragments that are at least 95% identical to the amino acid fragments  
30 reported herein. Similarly, preferred *chnB* encoding nucleic acid sequences corresponding to the instant ORF's are those encoding active proteins and which are at least 80% identical to the nucleic acid sequences reported herein. More preferred *chnB* nucleic acid fragments are at least 90% identical to the sequences herein. Most preferred are  
35 *chnB* nucleic acid fragments that are at least 95% identical to the nucleic acid fragments reported herein.

          Comparison of the *Rhodococcus erythropolis* AN1 ORF15 *chnB* nucleotide base and deduced amino acid sequences to public databases

reveals that the most similar known sequences range from a distant as about 39% identical to the amino acid sequence of reported herein over length of 649 amino acids using a Smith-Waterman alignment algorithm (W. R. Pearson, *supra*). Preferred amino acid fragments are at least about 5 70% - 80% and more preferred amino acid fragments are at least about 80%-90% identical to the sequences herein. Most preferred are nucleic acid fragments that are at least 95% identical to the amino acid fragments reported herein. Similarly, preferred *chnB* encoding nucleic acid sequences corresponding to the instant ORF's are those encoding active 10 proteins and which are at least 80% identical to the nucleic acid sequences reported herein. More preferred *chnB* nucleic acid fragments are at least 90% identical to the sequences herein. Most preferred are *chnB* nucleic acid fragments that are at least 95% identical to the nucleic acid fragments reported herein.

15        Comparison of the *Rhodococcus erythropolis* AN1 ORF16 *chnB* nucleotide base and deduced amino acid sequences to public databases reveals that the most similar known sequences range from a distant as about 43% identical to the amino acid sequence of reported herein over length of 494 amino acids using a Smith-Waterman alignment algorithm 20 (W. R. Pearson, *supra*). Preferred amino acid fragments are at least about 70% - 80% and more preferred amino acid fragments are at least about 80%-90% identical to the sequences herein. Most preferred are nucleic acid fragments that are at least 95% identical to the amino acid fragments reported herein. Similarly, preferred *chnB* encoding nucleic acid 25 sequences corresponding to the instant ORF's are those encoding active proteins and which are at least 80% identical to the nucleic acid sequences reported herein. More preferred *chnB* nucleic acid fragments are at least 90% identical to the sequences herein. Most preferred are *chnB* nucleic acid fragments that are at least 95% identical to the nucleic acid fragments reported herein. 30

Comparison of the *Rhodococcus erythropolis* AN1 ORF17 *chnB* nucleotide base and deduced amino acid sequences to public databases reveals that the most similar known sequences range from a distant as about 53% identical to the amino acid sequence of reported herein over 35 length of 499 amino acids using a Smith-Waterman alignment algorithm (W. R. Pearson, *supra*). Preferred amino acid fragments are at least about 70% - 80% and more preferred amino acid fragments are at least about 80%-90% identical to the sequences herein. Most preferred are nucleic

acid fragments that are at least 95% identical to the amino acid fragments reported herein. Similarly, preferred *chnB* encoding nucleic acid sequences corresponding to the instant ORF's are those encoding active proteins and which are at least 80% identical to the nucleic acid sequences reported herein. More preferred *chnB* nucleic acid fragments are at least 90% identical to the sequences herein. Most preferred are *chnB* nucleic acid fragments that are at least 95% identical to the nucleic acid fragments reported herein.

Comparison of the *Rhodococcus erythropolis* AN1 ORF18 *chnB* nucleotide base and deduced amino acid sequences to public databases reveals that the most similar known sequences range from a distant as about 44% identical to the amino acid sequence of reported herein over length of 493 amino acids using a Smith-Waterman alignment algorithm (W. R. Pearson, *supra*). Preferred amino acid fragments are at least about 70% - 80% and more preferred amino acid fragments are at least about 80%-90% identical to the sequences herein. Most preferred are nucleic acid fragments that are at least 95% identical to the amino acid fragments reported herein. Similarly, preferred *chnB* encoding nucleic acid sequences corresponding to the instant ORF's are those encoding active proteins and which are at least 80% identical to the nucleic acid sequences reported herein. More preferred *chnB* nucleic acid fragments are at least 90% identical to the sequences herein. Most preferred are *chnB* nucleic acid fragments that are at least 95% identical to the nucleic acid fragments reported herein.

Comparison of the *Rhodococcus erythropolis* AN1 ORF19 *chnB* nucleotide base and deduced amino acid sequences to public databases reveals that the most similar known sequences range from a distant as about 54% identical to the amino acid sequence of reported herein over length of 541 amino acids using a Smith-Waterman alignment algorithm (W. R. Pearson, *supra*). Preferred amino acid fragments are at least about 70% - 80% and more preferred amino acid fragments are at least about 80%-90% identical to the sequences herein. Most preferred are nucleic acid fragments that are at least 95% identical to the amino acid fragments reported herein. Similarly, preferred *chnB* encoding nucleic acid sequences corresponding to the instant ORF's are those encoding active proteins and which are at least 80% identical to the nucleic acid sequences reported herein. More preferred *chnB* nucleic acid fragments are at least 90% identical to the sequences herein. Most preferred are

*chnB* nucleic acid fragments that are at least 95% identical to the nucleic acid fragments reported herein.

Comparison of the *Rhodococcus erythropolis* AN1 ORF20 *chnB* nucleotide base and deduced amino acid sequences to public databases  
5 reveals that the most similar known sequences range from a distant as about 42% identical to the amino acid sequence of reported herein over length of 545 amino acids using a Smith-Waterman alignment algorithm (W. R. Pearson, *supra*). Preferred amino acid fragments are at least about 70% - 80% and more preferred amino acid fragments are at least about  
10 80%-90% identical to the sequences herein. Most preferred are nucleic acid fragments that are at least 95% identical to the amino acid fragments reported herein. Similarly, preferred *chnB* encoding nucleic acid sequences corresponding to the instant ORF's are those encoding active proteins and which are at least 80% identical to the nucleic acid  
15 sequences reported herein. More preferred *chnB* nucleic acid fragments are at least 90% identical to the sequences herein. Most preferred are *chnB* nucleic acid fragments that are at least 95% identical to the nucleic acid fragments reported herein.

In addition to the identification of the above mentioned sequences  
20 and the biochemical characterization of the activity of the gene product, Applicants have made the discovery that many of these monooxygenase proteins share diagnostic signature sequences which may be used for the identification of other proteins having similar activity. For example, the present monooxygenases may be grouped into three general families  
25 based on sequence alignment. One group, referred to herein BV Family 1, is comprised of the monooxygenase sequences shown in Figure 7 and generating the consensus sequence as set forth in SEQ ID NO:47. As will be seen in Figure 7, there are a group of completely conserved amino acids in 74 positions across all of the sequences of Figure 7. These  
30 positions are further delineated in Figure 6, and indicated as p1 – p74.

Similarly, BV Family 2 is comprised of the monooxygenase sequences shown on Figure 8, and generating the consensus sequence as set forth in SEQ ID NO:48. The signature sequence of BV Family 2 monooxygenases is shown in Figure 6 having the positions p1-p76.  
35 BV Family 3 monooxygenases are shown in Figure 9, generating the consensus sequence as set for the in SEQ ID NO:49, having the signature sequence as shown in Figure 6 of positions p1-p41.

Although there is variation among the sequences of the various families, all of the individual members of these families have been shown to possess monooxygenase activity. Thus, it is contemplated that where a polypeptide possesses the signature sequences as defined in Figures 6-9 that it will have monooxygenase activity. It is thus within the scope of the present invention to provide a method for identifying a gene encoding a Baeyer-Villiger monooxygenase polypeptide comprising:

- (a) probing a genomic library with a nucleic acid fragment encoding a polypeptide wherein where at least 80% of the amino acid residues at positions p1- p74 of SEQ ID NO:47, or at least 80% of the amino acid residues at p1-p76 of SEQ ID NO:48 or at least 80% of the amino acid residues of p1-p41 of SEQ ID NO:49 are completely conserved;
- (b) identifying a DNA clone that hybridizes with a nucleic acid fragment of step (a);
- (c) sequencing the genomic fragment that comprises the clone identified in step (b),

wherein the sequenced genomic fragment encodes a Baeyer-Villiger monooxygenase polypeptide.

In a preferred embodiment the invention provides the above method wherein where at least 100% of the amino acid residues at positions p1- p74 of SEQ ID NO:47, or at least 100% of the amino acid residues at p1-p76 of SEQ ID NO:48 or at least 100% of the amino acid residues of p1-p41 of SEQ ID NO:49 are completely conserved.

It will be appreciated that other Baeyer-Villiger monooxygenase genes having similar substrate specificity may be identified and isolated on the basis of sequence dependent protocols or according to alignment against the signature sequences disclosed herein.

Isolation of homologous genes using sequence-dependent protocols is well known in the art. Examples of sequence-dependent protocols include, but are not limited to, methods of nucleic acid hybridization, and methods of DNA and RNA amplification as exemplified by various uses of nucleic acid amplification technologies (e.g polymerase chain reaction (PCR), Mullis *et al.*, U.S. Patent 4,683,202), ligase chain reaction (LCR), Tabor, S. *et al.*, *Proc. Acad. Sci. USA* 82: 1074, (1985)) or strand displacement amplification (SDA, Walker, *et al.*, *Proc. Natl. Acad. Sci. U.S.A.*, 89: 392, (1992)).

For example, genes encoding similar proteins or polypeptides to the present Baeyer-Villiger monooxygenases could be isolated directly by using all or a portion of the nucleic acid fragments set forth in SEQ ID NOs:7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, and 45 or as DNA hybridization probes to screen libraries from any desired bacteria using methodology well known to those skilled in the art. Specific oligonucleotide probes based upon the instant nucleic acid sequences can be designed and synthesized by methods known in the art (Maniatis, *supra*). Moreover, the entire sequences can be used directly to synthesize DNA probes by methods known to the skilled artisan such as random primers DNA labeling, nick translation, or end-labeling techniques, or RNA probes using available *in vitro* transcription systems. In addition, specific primers can be designed and used to amplify a part of or full-length of the instant sequences. The resulting amplification products can be labeled directly during amplification reactions or labeled after amplification reactions, and used as probes to isolate full length DNA fragments under conditions of appropriate stringency.

Typically, in PCR-type primer directed amplification techniques, the primers have different sequences and are not complementary to each other. Depending on the desired test conditions, the sequences of the primers should be designed to provide for both efficient and faithful replication of the target nucleic acid. Methods of PCR primer design are common and well known in the art. (Thein and Wallace, "The use of oligonucleotide as specific hybridization probes in the Diagnosis of Genetic Disorders", in *Human Genetic Diseases: A Practical Approach*, K. E. Davis Ed., (1986) pp. 33-50 IRL Press, Herndon, Virginia; Rychlik, W. (1993) In White, B. A. (ed.), *Methods in Molecular Biology*, Vol. 15, pages 31-39, PCR Protocols: Current Methods and Applications. Humana Press, Inc., Totowa, NJ.)

Generally PCR primers may be used to amplify longer nucleic acid fragments encoding homologous genes from DNA or RNA. However, the polymerase chain reaction may also be performed on a library of cloned nucleic acid fragments wherein the sequence of one primer is derived from the instant nucleic acid fragments, and the sequence of the other primer takes advantage of the presence of the polyadenylic acid tracts to the 3' end of the mRNA precursor encoding microbial genes. Alternatively, the second primer sequence may be based upon sequences derived from the cloning vector. For example, the skilled artisan can

follow the RACE protocol (Frohman *et al.*, *PNAS USA* 85:8998 (1988)) to generate cDNAs by using PCR to amplify copies of the region between a single point in the transcript and the 3' or 5' end. Primers oriented in the 3' and 5' directions can be designed from the instant sequences. Using  
5 commercially available 3' RACE or 5' RACE systems (BRL), specific 3' or 5' cDNA fragments can be isolated (Ohara *et al.*, *PNAS USA* 86:5673 (1989); Loh *et al.*, *Science* 243:217 (1989)).

Accordingly the invention provides a method for identifying a nucleic acid molecule encoding a Baeyer-Villiger monooxygenase  
10 comprising: (a) synthesizing at least one oligonucleotide primer corresponding to a portion of the sequence selected from the group consisting of SEQ ID NOs:7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, and 45 and (b) amplifying an insert present in a cloning vector using the oligonucleotide primer of step (a); wherein the  
15 amplified insert encodes a Baeyer-Villiger monooxygenase

Alternatively the instant sequences may be employed as hybridization reagents for the identification of homologs. The basic components of a nucleic acid hybridization test include a probe, a sample suspected of containing the gene or gene fragment of interest, and a  
20 specific hybridization method. Probes of the present invention are typically single stranded nucleic acid sequences which are complementary to the nucleic acid sequences to be detected. Probes are "hybridizable" to the nucleic acid sequence to be detected. The probe length can vary from 5 bases to tens of thousands of bases, and will depend upon the specific  
25 test to be done. Typically a probe length of about 15 bases to about 30 bases is suitable. Only part of the probe molecule need be complementary to the nucleic acid sequence to be detected. In addition, the complementarity between the probe and the target sequence need not be perfect. Hybridization does occur between imperfectly complementary  
30 molecules with the result that a certain fraction of the bases in the hybridized region are not paired with the proper complementary base.

Hybridization methods are well defined. Typically the probe and sample must be mixed under conditions which will permit nucleic acid hybridization. This involves contacting the probe and sample in the  
35 presence of an inorganic or organic salt under the proper concentration and temperature conditions. The probe and sample nucleic acids must be in contact for a long enough time that any possible hybridization between the probe and sample nucleic acid may occur. The concentration of probe

or target in the mixture will determine the time necessary for hybridization to occur. The higher the probe or target concentration the shorter the hybridization incubation time needed. Optionally a chaotropic agent may be added. The chaotropic agent stabilizes nucleic acids by inhibiting  
5 nuclease activity. Furthermore, the chaotropic agent allows sensitive and stringent hybridization of short oligonucleotide probes at room temperature [Van Ness and Chen (1991) *Nucl. Acids Res.* 19:5143-5151]. Suitable chaotropic agents include guanidinium chloride, guanidinium thiocyanate, sodium thiocyanate, lithium tetrachloroacetate, sodium perchlorate,  
10 rubidium tetrachloroacetate, potassium iodide, and cesium trifluoroacetate, among others. Typically, the chaotropic agent will be present at a final concentration of about 3M. If desired, one can add formamide to the hybridization mixture, typically 30-50% (v/v).

Various hybridization solutions can be employed. Typically, these  
15 comprise from about 20 to 60% volume, preferably 30%, of a polar organic solvent. A common hybridization solution employs about 30-50% v/v formamide, about 0.15 to 1M sodium chloride, about 0.05 to 0.1M buffers, such as sodium citrate, Tris-HCl, PIPES or HEPES (pH range about 6-9), about 0.05 to 0.2% detergent, such as sodium dodecylsulfate,  
20 or between 0.5-20 mM EDTA, FICOLL (Pharmacia Inc.) (about 300-500 kilodaltons), polyvinylpyrrolidone (about 250-500 kdal), and serum albumin. Also included in the typical hybridization solution will be unlabeled carrier nucleic acids from about 0.1 to 5 mg/mL, fragmented nucleic DNA, e.g., calf thymus or salmon sperm DNA, or yeast RNA, and  
25 optionally from about 0.5 to 2% wt/vol glycine. Other additives may also be included, such as volume exclusion agents which include a variety of polar water-soluble or swellable agents, such as polyethylene glycol, anionic polymers such as polyacrylate or polymethylacrylate, and anionic saccharidic polymers, such as dextran sulfate.

30 Thus, the invention provides a method for identifying a nucleic acid molecule encoding a Baeyer-Villiger monooxygenase comprising:(a) probing a genomic library with a portion of a nucleic acid molecule selected from the group consisting of SEQ ID NOs:7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, and 45 ;(b) identifying a DNA  
35 clone that hybridizes under conditions of 0.1X SSC, 0.1% SDS, 65°C and washed with 2X SSC, 0.1% SDS followed by 0.1X SSC, 0.1% SDS with the nucleic acid molecule of (a); and (c) sequencing the genomic fragment



that comprises the clone identified in step (b), wherein the sequenced genomic fragment encodes Baeyer-Villiger monooxygenase.

#### Recombinant Expression—Microbial

The genes and gene products of the present BVMO sequences may be introduced into microbial host cells. Preferred host cells for expression of the instant genes and nucleic acid molecules are microbial hosts that can be found broadly within the fungal or bacterial families and which grow over a wide range of temperature, pH values, and solvent tolerances. Because of transcription, translation and the protein biosynthetic apparatus is the same irrespective of the cellular feedstock, functional genes are expressed irrespective of carbon feedstock used to generate cellular biomass. Large scale microbial growth and functional gene expression may utilize a wide range of simple or complex carbohydrates, organic acids and alcohols, saturated hydrocarbons such as methane or carbon dioxide in the case of photosynthetic or chemoautotrophic hosts. However, the functional genes may be regulated, repressed or depressed by specific growth conditions, which may include the form and amount of nitrogen, phosphorous, sulfur, oxygen, carbon or any trace micronutrient including small inorganic ions. In addition, the regulation of functional genes may be achieved by the presence or absence of specific regulatory molecules that are added to the culture and are not typically considered nutrient or energy sources. Growth rate may also be an important regulatory factor in gene expression. Examples of suitable host strains include but are not limited to fungal or yeast species such as *Aspergillus*, *Trichoderma*, *Saccharomyces*, *Pichia*, *Candida*, *Hansenula*, or bacterial species such as member of the proteobacteria and actinomycetes as well as the specific genera *Rhodococcus*, *Acinetobacter*, *Arthrobacter*, *Mycobacteria*, *Nocardia*, *Brevibacterium*, *Acidovorax*, *Bacillus*, *Streptomyces*, *Escherichia*, *Salmonella*, *Pseudomonas*, *Aspergillus*, *Saccharomyces*, *Pichia*, *Candida*, *Corynebacterium*, and *Hansenula*.

Particularly suitable in the present invention as hosts for monooxygenase are the members of the Proteobacteria and Actinomycetes. The Proteobacteria form a physiologically diverse group of microorganisms and represent five subdivisions ( $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\epsilon$ ,  $\delta$ ) (Madigan et al., Brock Biology of Microorganisms, 8th edition, Prentice Hall, UpperSaddle River, NJ (1997)). All five subdivisions of the Proteobacteria contain microorganisms that use organic compounds as sources of

carbon and energy. Members of the Proteobacteria suitable in the present invention include, but are not limited to *Burkholderia*, *Alcaligenes*, *Pseudomonas*, *Sphingomonas*, *Pandoraea*, *Delftia* and *Comamonas*.

Microbial expression systems and expression vectors containing  
5 regulatory sequences that direct high level expression of foreign proteins are well known to those skilled in the art. Any of these could be used to construct chimeric genes for production of the any of the gene products of the instant sequences. These chimeric genes could then be introduced into appropriate microorganisms via transformation to provide high level  
10 expression of the enzymes.

Vectors or cassettes useful for the transformation of suitable host cells are well known in the art. Typically the vector or cassette contains sequences directing transcription and translation of the relevant gene, a selectable marker, and sequences allowing autonomous replication or  
15 chromosomal integration. Suitable vectors comprise a region 5' of the gene which harbors transcriptional initiation controls and a region 3' of the DNA fragment which controls transcriptional termination. It is most preferred when both control regions are derived from genes homologous to the transformed host cell, although it is to be understood that such  
20 control regions need not be derived from the genes native to the specific species chosen as a production host.

Initiation control regions or promoters, which are useful to drive expression of the instant ORF's in the desired host cell are numerous and familiar to those skilled in the art. Virtually any promoter capable of driving  
25 these genes is suitable for the present invention including but not limited to *CYC1*, *HIS3*, *GAL1*, *GAL10*, *ADH1*, *PGK*, *PHO5*, *GAPDH*, *ADC1*, *TRP1*, *URA3*, *LEU2*, *ENO*, *TPI* (useful for expression in *Saccharomyces*); *AOX1* (useful for expression in *Pichia*); and *lac*, *ara*, *tet*, *trp*, *IP<sub>L</sub>*, *IP<sub>R</sub>*, *T7*, *tac*, and *trc* (useful for expression in *Escherichia coli*) as well as the *amy*,  
30 *apr*, *npr* promoters and various phage promoters useful for expression in *Bacillus*.

Termination control regions may also be derived from various genes native to the preferred hosts. Optionally, a termination site may be unnecessary, however, it is most preferred if included.

#### 35 Recombinant Expression—Plants

The sequences encoding the BVMO's of the present invention may be used to create transgenic plants having the ability to express the

microbial proteins. Preferred plant hosts will be any variety that will support a high production level of the instant proteins.

Suitable green plants will included but are not limited to of soybean, rapeseed (*Brassica napus*, *B. campestris*), sunflower (*Helianthus annuus*),  
5 cotton (*Gossypium hirsutum*), corn, tobacco (*Nicotiana tabacum*), alfalfa (*Medicago sativa*), wheat (*Triticum sp*), barley (*Hordeum vulgare*), oats (*Avena sativa*, L), sorghum (*Sorghum bicolor*), rice (*Oryza sativa*),  
*Arabidopsis*, cruciferous vegetables (broccoli, cauliflower, cabbage, parsnips, etc.), melons, carrots, celery, parsley, tomatoes, potatoes,  
10 strawberries, peanuts, grapes, grass seed crops, sugar beets, sugar cane, beans, peas, rye, flax, hardwood trees, softwood trees, and forage grasses. Algal species include but not limited to commercially significant hosts such as *Spirulina* and *Dunaliella*. Overexpression of the proteins of the instant invention may be accomplished by first constructing chimeric  
15 genes in which the coding region are operably linked to promoters capable of directing expression of a gene in the desired tissues at the desired stage of development. For reasons of convenience, the chimeric genes may comprise promoter sequences and translation leader sequences derived from the same genes. 3' Non-coding sequences encoding  
20 transcription termination signals must also be provided. The instant chimeric genes may also comprise one or more introns in order to facilitate gene expression.

Any combination of any promoter and any terminator capable of inducing expression of a coding region may be used in the chimeric  
25 genetic sequence. Some suitable examples of promoters and terminators include those from nopaline synthase (*nos*), octopine synthase (*ocs*) and cauliflower mosaic virus (*CaMV*) genes. One type of efficient plant promoter that may be used is a high level plant promoter. Such promoters, in operable linkage with the genetic sequences or the present  
30 invention should be capable of promoting expression of the present gene product. High level plant promoters that may be used in this invention include the promoter of the small subunit (ss) of the ribulose-1,5-bisphosphate carboxylase from example from soybean (Berry-Lowe *et al.*, *J. Molecular and App. Gen.*, 1:483-498 1982)), and the promoter of the  
35 chlorophyll a/b binding protein. These two promoters are known to be light-induced in plant cells (See, for example, Genetic Engineering of Plants, an Agricultural Perspective, A. Cashmore, Plenum, New York (1983), pages 29-38; Coruzzi, G. *et al.*, *The Journal of Biological*

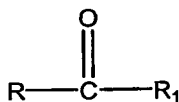
*Chemistry*, 258:1399 (1983), and Dunsmuir, P. *et al.*, *Journal of Molecular and Applied Genetics*, 2:285 (1983)).

Plasmid vectors comprising the instant chimeric genes can then be constructed. The choice of plasmid vector depends upon the method that will be used to transform host plants. The skilled artisan is well aware of the genetic elements that must be present on the plasmid vector in order to successfully transform, select and propagate host cells containing the chimeric gene. The skilled artisan will also recognize that different independent transformation events will result in different levels and patterns of expression (Jones *et al.*, *EMBO J.* 4:2411-2418 (1985); De Almeida *et al.*, *Mol. Gen. Genetics* 218:78-86 (1989)), and thus that multiple events must be screened in order to obtain lines displaying the desired expression level and pattern. Such screening may be accomplished by Southern analysis of DNA blots (Southern, *J. Mol. Biol.* 98:503, (1975)). Northern analysis of mRNA expression (Kroczeck, *J. Chromatogr. Biomed. Appl.*, 618 (1-2):133-145 (1993)), Western analysis of protein expression, or phenotypic analysis.

For some applications it will be useful to direct the instant proteins to different cellular compartments. It is thus envisioned that the chimeric genes described above may be further supplemented by altering the coding sequences to encode enzymes with appropriate intracellular targeting sequences such as transit sequences (Keegstra, K., *Cell* 56:247-253 (1989)), signal sequences or sequences encoding endoplasmic reticulum localization (Chrispeels, J.J., *Ann. Rev. Plant Phys. Plant Mol. Biol.* 42:21-53 (1991)), or nuclear localization signals (Raikhel, N. *Plant Phys.* 100:1627-1632 (1992)) added and/or with targeting sequences that are already present removed. While the references cited give examples of each of these, the list is not exhaustive and more targeting signals of utility may be discovered in the future that are useful in the invention.

#### Process for the Production of Lactones and Esters from Ketone Substrates

Once the appropriate nucleic acid sequence has been expressed in a recombinant organism, the organism may be contacted with a suitable ketone substrate for the production of the corresponding ester. The Baeyer-Villiger monooxygenases of the instant invention will act on a variety of ketone substrates comprising cyclic ketones and ketoterpenes to produce the corresponding lactone or ester. Suitable ketone substrates for the conversion to esters are defined by the general formula:



wherein R and R<sub>1</sub> are independently selected from substituted or  
5 unsubstituted phenyl, substituted or unsubstituted alkyl, or substituted or  
unsubstituted alkenyl or substituted or unsubstituted alkylidene.  
Particularly useful ketone substrates include, but are not limited to  
Norcamphor, Cyclobutanone, Cyclopentanone, 2-methyl-cyclopentanone,  
Cyclohexanone, 2-methyl-cyclohexanone, Cyclohex-2-ene-1-one, 1,2-  
10 cyclohexanedione, 1,3-cyclohexanedione, 1,4-cyclohexanedione,  
Cycloheptanone, Cyclooctanone, Cyclodecanone, Cycloundecanone,  
Cyclododecanone, Cyclotridecanone, Cyclopenta-decanone, 2-  
tridecanone, dihexyl ketone, 2-phenyl-cyclohexanone, Oxindole,  
Levoglucofenone, dimethyl sulfoxide, dimethyl-2-piperidone,  
15 Phenylboronic acid, and beta-ionone.

Alternatively it is contemplated that the enzymes of the invention  
may be used in vitro for the transformation of ketone substrates to the  
corresponding esters. The monooxygenase enzymes may be produced  
recombinantly or isolated from native sources, purified and reacted with  
20 the appropriate substrate under suitable conditions of pH and  
temperature.

Where large scale commercial production of lactones or esters is  
desired, a variety of culture methodologies may be applied. For example,  
large scale production from a recombinant microbial host may be  
25 produced by both batch or continuous culture methodologies.

A classical batch culturing method is a closed system where the  
composition of the media is set at the beginning of the culture and not  
subject to artificial alterations during the culturing process. Thus, at the  
beginning of the culturing process the media is inoculated with the desired  
30 organism or organisms and growth or metabolic activity is permitted to  
occur adding nothing to the system. Typically, however, a "batch" culture  
is batch with respect to the addition of carbon source and attempts are  
often made at controlling factors such as pH and oxygen concentration. In  
batch systems the metabolite and biomass compositions of the system  
35 change constantly up to the time the culture is terminated. Within batch  
cultures cells moderate through a static lag phase to a high growth log  
phase and finally to a stationary phase where growth rate is diminished or

halted. If untreated, cells in the stationary phase will eventually die. Cells in log phase are often responsible for the bulk of production of end product or intermediate in some systems. Stationary or post-exponential phase production can be obtained in other systems.

5           A variation on the standard batch system is the Fed-Batch system. Fed-Batch culture processes are also suitable in the present invention and comprise a typical batch system with the exception that the substrate is added in increments as the culture progresses. Fed-Batch systems are useful when catabolite repression is apt to inhibit the metabolism of the  
10       cells and where it is desirable to have limited amounts of substrate in the media. Measurement of the actual substrate concentration in Fed-Batch systems is difficult and is therefore estimated on the basis of the changes of measurable factors such as pH, dissolved oxygen and the partial pressure of waste gases such as CO<sub>2</sub>. Batch and Fed-Batch culturing  
15       methods are common and well known in the art and examples may be found in Thomas D. Brock in *Biotechnology: A Textbook of Industrial Microbiology*, Second Edition (1989) Sinauer Associates, Inc., Sunderland, MA., or Deshpande, Mukund V., *Appl. Biochem. Biotechnol.*, 36, 227, (1992), herein incorporated by reference.

20           Commercial production of lactones and esters of the present invention may also be accomplished with a continuous culture. Continuous cultures are an open system where a defined culture media is added continuously to a bioreactor and an equal amount of conditioned media is removed simultaneously for processing. Continuous cultures  
25       generally maintain the cells at a constant high liquid phase density where cells are primarily in log phase growth. Alternatively continuous culture may be practiced with immobilized cells where carbon and nutrients are continuously added, and valuable products, by-products or waste products are continuously removed from the cell mass. Cell immobilization may be  
30       performed using a wide range of solid supports composed of natural and/or synthetic materials.

          Continuous or semi-continuous culture allows for the modulation of one factor or any number of factors that affect cell growth or end product concentration. For example, one method will maintain a limiting nutrient  
35       such as the carbon source or nitrogen level at a fixed rate and allow all other parameters to moderate. In other systems a number of factors affecting growth can be altered continuously while the cell concentration, measured by media turbidity, is kept constant. Continuous systems strive

to maintain steady state growth conditions and thus the cell loss due to media being drawn off must be balanced against the cell growth rate in the culture. Methods of modulating nutrients and growth factors for continuous culture processes as well as techniques for maximizing the rate of product formation are well known in the art of industrial microbiology and a variety of methods are detailed by Brock, *supra*.  
Baeyer-Villiger monooxygenases having enhanced activity

It is contemplated that the present BVMO sequences may be used to produce gene products having enhanced or altered activity. Various methods are known for mutating a native gene sequence to produce a gene product with altered or enhanced activity including but not limited to error prone PCR (Melnikov *et al.*, *Nucleic Acids Research*, (Feb. 15, 1999) Vol. 27, No. 4, pp. 1056-1062); site directed mutagenesis (Coombs *et al.*, *Proteins* (1998), 259-311, 1 plate. Editor(s): Angeletti, Ruth Hogue. Publisher: Academic, San Diego, CA) and "gene shuffling" (US 5,605,793; US 5,811,238; US 5,830,721; and US 5,837,458, incorporated herein by reference).

The method of gene shuffling is particularly attractive due to its facile implementation, and high rate of mutagenesis and ease of screening. The process of gene shuffling involves the restriction endonuclease cleavage of a gene of interest into fragments of specific size in the presence of additional populations of DNA regions of both similarity to or difference to the gene of interest. This pool of fragments will then be denatured and reannealed to create a mutated gene. The mutated gene is then screened for altered activity.

The BVMO sequences of the present invention may be mutated and screened for altered or enhanced activity by this method. The sequences should be double stranded and can be of various lengths ranging from 50 bp to 10 kb. The sequences may be randomly digested into fragments ranging from about 10 bp to 1000 bp, using restriction endonucleases well known in the art (Maniatis *supra*). In addition to the instant microbial sequences, populations of fragments that are hybridizable to all or portions of the microbial sequence may be added. Similarly, a population of fragments which are not hybridizable to the instant sequence may also be added. Typically these additional fragment populations are added in about a 10 to 20 fold excess by weight as compared to the total nucleic acid. Generally if this process is followed the number of different specific nucleic acid fragments in the mixture will

be about 100 to about 1000. The mixed population of random nucleic acid fragments are denatured to form single-stranded nucleic acid fragments and then reannealed. Only those single-stranded nucleic acid fragments having regions of homology with other single-stranded nucleic acid fragments will reanneal. The random nucleic acid fragments may be denatured by heating. One skilled in the art could determine the conditions necessary to completely denature the double stranded nucleic acid. Preferably the temperature is from 80°C to 100°C. The nucleic acid fragments may be reannealed by cooling. Preferably the temperature is from 20°C to 75°C. Renaturation can be accelerated by the addition of polyethylene glycol ("PEG") or salt. A suitable salt concentration may range from 0 mM to 200 mM. The annealed nucleic acid fragments are then incubated in the presence of a nucleic acid polymerase and dNTP's (i.e. dATP, dCTP, dGTP and dTTP). The nucleic acid polymerase may be the Klenow fragment, the Taq polymerase or any other DNA polymerase known in the art. The polymerase may be added to the random nucleic acid fragments prior to annealing, simultaneously with annealing or after annealing. The cycle of denaturation, renaturation and incubation in the presence of polymerase is repeated for a desired number of times. Preferably the cycle is repeated from 2 to 50 times, more preferably the sequence is repeated from 10 to 40 times. The resulting nucleic acid is a larger double-stranded polynucleotide ranging from about 50 bp to about 100 kb and may be screened for expression and altered activity by standard cloning and expression protocol. (Manatis *supra*).

Furthermore, a hybrid protein can be assembled by fusion of functional domains using the gene shuffling (exon shuffling) method (Nixon *et al*, PNAS, 94:1069-1073 (1997)). The functional domain of the instant gene can be combined with the functional domain of other genes to create novel enzymes with desired catalytic function. A hybrid enzyme may be constructed using PCR overlap extension method and cloned into the various expression vectors using the techniques well known to those skilled in art.

### EXAMPLES

The present invention is further defined in the following Examples. It should be understood that these Examples, while indicating preferred embodiments of the invention, are given by way of illustration only. From the above discussion and these Examples, one skilled in the art can ascertain the essential characteristics of this invention, and without



departing from the spirit and scope thereof, can make various changes and modifications of the invention to adapt it to various usages and conditions.

#### GENERAL METHODS

5           Standard recombinant DNA and molecular cloning techniques used in the Examples are well known in the art and are described by Sambrook, J., Fritsch, E. F. and Maniatis, T. *Molecular Cloning: A Laboratory Manual*; Cold Spring Harbor Laboratory Press: Cold Spring Harbor, (1989) (Maniatis) and by T. J. Silhavy, M. L. Bennis, and L. W. Enquist, 10 Experiments with Gene Fusions, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y. (1984) and by Ausubel, F. M. *et al.*, Current Protocols in Molecular Biology, pub. by Greene Publishing Assoc. and Wiley-Interscience (1987).

15           Materials and methods suitable for the maintenance and growth of bacterial cultures are well known in the art. Techniques suitable for use in the following examples may be found as set out in Manual of Methods for General Bacteriology (Phillipp Gerhardt, R. G. E. Murray, Ralph N. Costilow, Eugene W. Nester, Willis A. Wood, Noel R. Krieg and G. Briggs Phillips, Eds., American Society for Microbiology, Washington, DC. 20 (1994)) or by Thomas D. Brock in Biotechnology: A Textbook of Industrial Microbiology, Second Ed., Sinauer Associates, Inc.: Sunderland, MA (1989). All reagents, restriction enzymes and materials used for the growth and maintenance of bacterial cells were obtained from Aldrich Chemicals (Milwaukee, WI), DIFCO Laboratories (Detroit, MI), 25 GIBCO/BRL (Gaithersburg, MD), or Sigma Chemical Company (St. Louis, MO) unless otherwise specified.

Bacterial Strains and Plasmids: *Rhodococcus erythropolis* AN12, *Brevibacterium* sp. HCU, *Arthrobacter* sp. BP2, *Rhodococcus* sp. phi1, *Rhodococcus* sp. phi2, *Acidovorax* sp. CHX, and *Acinetobacter* sp. SE19 30 were isolated from enrichment of activated sludge obtained from industrial wastewater treatment facilities. Max Efficiency competent cells of *E. coli* DH5 $\alpha$  and DH10B were purchased from GIBCO/BRL (Gaithersburg, MD). Expression plasmid pQE30 were purchased from Qiagen (Valencia, CA), while cloning vector pCR2.1 and expression vector pTrc/His2-Topo were 35 purchased from Invitrogen (San Diego, CA).

Taxonomic identification of *Rhodococcus erythropolis* AN12, *Brevibacterium* sp. HCU, *Arthrobacter* sp. BP2, *Rhodococcus* sp. phi1, *Rhodococcus* sp. phi2, *Acidovorax* sp. CHX, and *Acinetobacter* sp. SE19

- was performed by PCR amplification of 16S rDNA from chromosomal DNA using primers corresponding to conserved regions of the 16S rDNA molecule (Table 2). The following temperature program was used: 95°C (5 min) for 1 cycle followed by 25 cycles of: 95°C (1 min), 55°C (1 min), 72°C (1 min), followed by a final extension at 72°C (8 min). Following DNA sequencing (according to the method shown below), the 16S rDNA gene sequence of each isolate was used as the query sequence for a BLAST search (Altschul, *et al.*, *Nucleic Acids Res.* 25:3389-3402 (1997)) against GenBank for similar sequences.

10

Table 2  
Primers to Conserved Regions of 16s rDNA

SEQ ID NO	Primer Sequence (5'- 3')	Reference
50	GAGTTTGATCCTGGCTCAG	(HK12) Amann, R.I. et al. <i>Microbiol. Rev.</i> 59(1):143-69 (1995)
51	CAGG(A/C)GCCGCGGTAAT(A/T)C	Amann, R.I. et al. <i>Microbiol. Rev.</i> 59(1):143-69 (1995)
52	GCTGCCTCCCGTAGGAGT	(HK21) Amann, R.I. et al. <i>Microbiol. Rev.</i> 59(1):143-69 (1995)
53	CTACCAGGGTAACTAATCC	Amann, R.I. et al. <i>Microbiol. Rev.</i> 59(1):143-69 (1995)
54	ACGGGCGGTGTGTAC	Amann, R.I. et al. <i>Microbiol. Rev.</i> 59(1):143-69 (1995)
55	CACGAGCTGACGACAGCCAT	Amann, R.I. et al. <i>Microbiol. Rev.</i> 59(1):143-69 (1995)
56	TACCTTGTTACGACTT	(HK13) Amann, R.I. et al. <i>Microbiol. Rev.</i> 59(1):143-69 (1995)
57	G(A/T)ATTACCGCGGC(G/T)GCTG	Amann, R.I. et al. <i>Microbiol. Rev.</i> 59(1):143-69 (1995)
58	GGATTAGATACCCTGGTAG	Amann, R.I. et al. <i>Microbiol. Rev.</i> 59(1):143-69 (1995)
59	ATGGCTGTCGTCAGCTCGTG	Amann, R.I. et al. <i>Microbiol. Rev.</i> 59(1):143-69 (1995)
60	GCCCCCG(C/T)CAATTCCT	(HK15) Kane, M.D. et al. <i>Appl. Environ. Microbiol.</i> 59:682-686 (1993)

SEQ ID NO	Primer Sequence (5'- 3')	Reference
61	GTGCCAGCAG(C/T)(A/C) GCGGT	( <b>HK14</b> ) Kane, M.D. et al. <i>Appl. Environ. Microbiol.</i> 59:682-686 (1993)
62	GCCAGCAGCCGCGGTA	( <b>JCR15</b> ) Kane, M.D. et al. <i>Appl. Environ. Microbiol.</i> 59:682-686 (1993)

Note: Parenthetical information in bold is the original name for the primer, according to the reference provided.

### Sequencing

- 5           Sequence was generated on an ABI Automatic sequencer using dye terminator technology (U.S. Patent 5,366,860; EP 272007) using a combination of vector and insert-specific primers. Sequence editing was performed using either Sequencher (Gene Codes Corp., Ann Arbor, MI) or the Wisconsin GCG program (Wisconsin Package Version 9.0, Genetics Computer Group (GCG), Madison, WI) and the CONSED package (version 7.0). All sequences represent coverage at least two times in both directions.

- 15           Manipulations of genetic sequences were accomplished using the suite of programs available from the Genetics Computer Group Inc. (Wisconsin Package Version 9.0, Genetics Computer Group (GCG), Madison, WI). Where the GCG program "Pileup" was used, the gap creation default value of 12 and the gap extension default value of 4 were used. Where the GCG "Gap" or "Bestfit" programs were used, the default gap creation penalty of 50 and the default gap extension penalty of 3 were used. In any case where GCG program parameters were not prompted for, in these or any other GCG program, default values were used.

- 25           The meaning of abbreviations is as follows: "sec" means second(s), "min" means minute(s), "h" means hour(s), "d" means day(s), "μL" means microliter, "mL" means milliliters, "L" means liters, "μM" means micromolar, "mM" means millimolar, "M" means molar, "mmol" means millimole(s), "μmole" mean micromole, "g" means gram, "μg" means microgram, "ng" means nanogram, "U" means units, "mU" means milliunits, "ppm" means parts per million, "psi" means pounds per square inch, and "kB" means kilobase.

### EXAMPLE 1

#### Monooxygenase Gene Discovery in a Mixed Microbial Population

This Example describes the isolation of the cyclohexanone degrading organisms *Arthrobacter* sp. BP2, *Rhodococcus* sp. phi1, and  
5 *Rhodococcus* sp. phi2 by enrichment of a mixed microbial community. Differential display techniques applied to cultures containing the mixed microbial population permitted discovery of monooxygenase genes.

#### Enrichment for cyclohexanone degraders

A mixed microbial community was obtained from a wastewater  
10 bioreactor and maintained on minimal medium (50 mM KHPO<sub>4</sub> (pH 7.0), 10 mM (NH<sub>4</sub>)SO<sub>4</sub>, 2 mM MgCl<sub>2</sub>, 0.7 mM CaCl<sub>2</sub>, 50 µM MnCl<sub>2</sub>, 1 µM FeCl<sub>3</sub>, 1 µM ZnCl<sub>3</sub>, 1.72 µM CuSO<sub>4</sub>, 2.53 µM CoCl<sub>2</sub>, 2.42 µM Na<sub>2</sub>MoO<sub>2</sub>, and 0.0001% FeSO<sub>4</sub>) with trace amounts of yeast extract casamino acids and peptone (YECAAP) at 0.1% concentration with 0.1% cyclohexanol  
15 and cyclohexanone added as carbon sources. Increased culture growth in the presence of cyclohexanone indicated a microbial population with members that could convert cyclohexanone.

#### Isolation of Strains

Seven individual strains were isolated from the community by  
20 spreading culture on R2A Agar (Becton Dickinson and Company, Cockeysville, MD) at 30° C. Strains were streaked to purity on the same medium. Among these seven strains, the strain identified as *Arthrobacter* species BP2 formed large colonies of a light yellow color. One *Rhodococcus* strain, identified as species phi1, formed small colonies that  
25 were orange in color. The other *Rhodococcus* strain, designated species phi2, formed small colonies that were red in color.

Individuals strains were identified by comparing 16s rDNA sequences to known 16S rRNA sequences in the GenBank sequence database. The 16S rRNA gene sequence from strain BP2 (SEQ ID NO:1)  
30 was at least 99% homologous to the 16S rRNA gene sequences of bacteria belonging to the genus *Arthrobacter*. The 16S rRNA gene sequences from strains phi1 and phi2 were each at least 99% homologous to the 16S rRNA gene sequences of bacteria belonging to the genus of gram positive bacteria, *Rhodococcus*. The complete 16S  
35 DNA sequence of *Rhodococcus* sp. phi1 is shown as SEQ ID NO:2, while that of *Rhodococcus* sp. phi2 is listed as SEQ ID NO:3.

### Induction of cyclohexanone oxidation genes

For induction of cyclohexanone oxidation genes within members of this community, 1 ml of inoculum from a waste water bioreactor was suspended in 25 ml minimal medium with 0.1% YECAAP and incubated overnight at 30°C with agitation. The next day 10 ml of the overnight culture was resuspended in a total volume of 50 ml minimal medium with 0.1% YECAAP. The optical density of the culture was 0.29 absorbance units at 600 nm. After equilibration at 30°C for 30 min, the culture was split into two separate 25 ml volumes. To one of these cultures, 25 µl (0.1%) cyclohexanone (Sigma-Aldrich, St. Louis, MO) was added. Both cultures were incubated for an additional 3 hrs. At this time, cultures were moved onto ice, harvested by centrifugation at 4°C, washed with two volumes of minimal salts medium and diluted to an optical density of 1.0 absorbance unit (600 nm). Approximately 6 ml of culture was placed in a water jacketed respirometry cell equipped with an oxygen electrode (Yellow Springs Instruments Co., Yellow Springs, OH) at 30°C to confirm cyclohexanone enzymes were induced. After establishing the baseline respiration for each cell suspension, cyclohexanone was added to a final concentration of 0.1% and the rate of O<sub>2</sub> consumption was further monitored. For the control culture, 2 mM potassium acetate was added 200 sec after the cyclohexanone.

### Isolation of total community RNA

After the 3 hr induction period with cyclohexanone described above, the control and induced sample (2 mL each) were harvested at 1400 rpm in a 4 °C centrifuge and resuspended in 900 µl Buffer RLT (Qiagen, Valencia, CA). A 300 µl volume of zirconia beads (Biospec Products, Bartlesville, OK) was added and cells were disrupted using a bead beater (Biospec Products) at 2400 beats per min for 3 min. Each of these samples was split into six aliquots for nucleic acid isolation using the RNeasy Mini Kit (Qiagen, Valencia, CA) and each was eluted with 100 RNase-free dH<sub>2</sub>O supplied with the kit. DNA was degraded in the samples using 10 mM MgCl<sub>2</sub>, 60 mM KCl and 2 U RNase-free DNase I (Ambion, Austin, TX) at 37 °C for 4 hr. Following testing for total DNA degradation by PCR using one of the arbitrary oligonucleotides used for RT-PCR, RNA was purified using the RNeasy Mini Kit and eluted in 100 µl RNase-free dH<sub>2</sub>O as described previously.

Generation of RAPDs from arbitrarily reverse-transcribed total RNA

A set of 244 primers with the sequence CGGAGCAGATCGAVVVV (SEQ ID NO:63); where VVVV represent all the combinations of the three bases A, G and C) was used in separate RT-PCR reactions as with RNA from either the control or induced cells. The SuperScript™ One-Step™ RT-PCR System (Life Technologies Gibco BRL, Rockville, MD) reaction mixture was used with 2-5 ng of total RNA in a 25 µl total reaction volume. The PCR was conducted using the following temperature program:

- 1 cycle: 4 °C (2 min), 5 min ramp to 37 °C (1 hr), followed by 95 °C incubation (3 min);
- 1 cycle: 94 °C (1 min), 40 °C (5 min), and 72 °C (5 min);
- 40 cycles: 94 °C (1 min), 60 °C (1 min), and 72 °C (1 min);
- 1 cycle: 70 °C (5 min) and 4 °C hold until separated by electrophoresis.

Products of these PCR amplifications (essentially RAPD fragments) were separated by electrophoresis at 1 V/cm on polyacrylamide gels (Amersham Pharmacia Biotech, Piscataway, NJ). Products resulting from the control mRNA (no cyclohexanone induction) and induced mRNA fragments were visualized by silver staining using an automated gel stainer (Amersham Pharmacia Biotech, Piscataway, NJ).

Reamplification of differentially expressed DNA fragments

A 25 µl volume of a sodium cyanide elution buffer (10mg/ml NaCN, 20 mM Tris-HCl (pH 8.0), 50 mM KCl and 0.05% NP40) was incubated with an excised gel band of a differentially display fragment at 95°C for 20 min. Reamplification of this DNA fragment was achieved in a PCR reaction using 5 µl of the elution mixture in a 25 µl reaction using the primer from which the fragment was originally generated. The temperature program for reamplification was: 94 °C (5 min); 20 cycles of 94 °C (1 min), 55 °C (1 min), and 72 °C (1 min); followed by 72 °C (7 min). The reamplification products were directly cloned into the pCR2.1-TOPO vector (Invitrogen, Carlsbad, CA) and were sequenced using an ABI model 377 with ABI BigDye terminator sequencing chemistry (Perseptive Biosystems, Framingham, MA). Eight clones were submitted for sequencing from each reamplified band. The nucleotide sequence of the cloned fragments was compared against the non-redundant GenBank database using the BlastX program (NCBI).

### Sequencing of cyclohexanone oxidation pathway genes

Oligonucleotides were designed to amplify by PCR individual differentially expressed fragments. Following DNA isolation from individual strains, these oligonucleotide primers were used to determine which strain contained DNA encoding the individual differentially expressed fragments. Cosmids were screened by PCR using primers designed against differentially displayed fragments with homology to known cyclohexanone degradation genes. Each recombinant *E. coli* cell culture carrying a cosmid clone (1.0 µl) was used as the template in a 25 ul PCR reaction mixture. The primer pair A102FI (SEQ ID NO:108) and CONR (SEQ ID NO:109) was used to screen the *Arthrobacter* sp. BP2 library, primer pair A228FI (SEQ ID NO:110) and A228RI (SEQ ID NO:111) was used to screen the *Rhodococcus* sp. phi2 library, and the primer pair of A2FI (SEQ ID NO:112) and A34RI (SEQ ID NO:113) was used to screen the *Rhodococcus* sp. phi1 library. Cosmids from recombinant *E. coli* which produced the correct product size in PCR reactions were isolated, digested partially with *Sau3AI* and 10-15 kB fragments from this partial digest were sub-cloned into the blue/white screening vector pSU19 (Bartolome, B. et al. *Gene*. 102(1): 75-8 (Jun 15, 1991); Martinez, E. et al. *Gene*. 68(1): 159-62 (Aug 15, 1988)). These sub-clones were isolated using Qiagen Turbo96 Miniprep kits and re-screened by PCR as previously described. Sub-clones carrying the correct sequence fragment were transposed with pGPS1.1 using the GPS-1 Genome Priming System kit (New England Biolabs, Inc., Beverly, MA). A number of these transposed plasmids were sequenced from each end of the transposon to obtain kilobase long DNA fragments. Sequence assembly was performed with the Sequencher program (Gene Codes Corp., Ann Arbor MI).

### EXAMPLE 2

#### Isolation of *Brevibacterium* sp. HCU Monooxygenase Genes Involved In The Oxidation Of Cyclohexanone

This Example describes the isolation of the cyclohexanol and cyclohexanone degrader *Brevibacterium* sp. HCU. Discovery of BV monooxygenase genes from the organism was accomplished using differential display methods.

#### Strain Isolation

Selection for a halotolerant bacterium degrading cyclohexanol and cyclohexanone was performed on agar plates of a halophilic minimal

medium (Per liter: 15 g Agar, 100 g NaCl, 10 g MgSO<sub>4</sub>, 2 g KCl, 1 g NH<sub>4</sub>Cl, 50 mg KH<sub>2</sub>PO<sub>4</sub>, 2 mg FeSO<sub>4</sub>, 8 g, Tris-HCl (pH 7)) containing traces of yeast extract and casaminoacids (0.005% each) and incubated under vapors of cyclohexanone at 30°C. The inoculum was a  
5 resuspension of sludge from industrial wastewater treatment plant. After two weeks, beige colonies were observed and streaked to purity on fresh agar plates grown under the same conditions.

The complete 16S DNA sequence of the isolated *Brevibacterium* sp. HCU was found to be unique and is shown as SEQ ID NO:4.  
10 Comparison to other 16S rRNA sequences in the GenBank sequence database found the 16S rRNA gene sequence from strain HCU was at least 99% homologous to the 16S rRNA gene sequences of bacteria belonging to the genus *Brevibacterium*.

#### Induction of the Cyclohexanone Degradation Pathway

15 Inducibility of the cyclohexanone pathway was tested by respirometry in low salt medium. One colony of *Brevibacterium* sp. HCU was inoculated in 300 ml of S12 mineral medium (50 mM KHPO<sub>4</sub> buffer (pH 7.0), 10 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 2 mM MgCl<sub>2</sub>, 0.7 mM CaCl<sub>2</sub>, 50 µM MnCl<sub>2</sub>, 1 µM FeCl<sub>3</sub>, 1 µM ZnCl<sub>3</sub>, 1.72 µM CuSO<sub>4</sub>, 2.53 µM CoCl<sub>2</sub>, 2.42 µM  
20 Na<sub>2</sub>MoO<sub>2</sub>, and 0.0001% FeSO<sub>4</sub>) containing 0.005% yeast extract. The culture was then split into two flasks which received respectively 10 mM acetate and 10 mM cyclohexanone. Each flask was incubated for 6 hrs at 30°C to allow for the induction of the cyclohexanone degradation genes. The cultures were then chilled on ice, harvested by centrifugation and  
25 washed three times with ice-cold S12 medium lacking traces of yeast extract. Cells were finally resuspended to an optical density of 2.0 at 600 nm and kept on ice until assayed.

Half a ml of each culture was placed in a water jacketed respirometry cell equipped with an oxygen electrode (Yellow Spring  
30 Instruments Co., Yellow spring, OH) and containing 5 ml of air saturated S12 medium at 30°C. After establishing the baseline respiration for each of the cell suspensions, acetate or cyclohexanone was added to a final concentration of 0.02% and the rate of O<sub>2</sub> consumption was further monitored.

#### 35 Identification of Cyclohexanone Oxidation Genes

Identification of genes involved in the oxidation of cyclohexanone made use of the fact that this oxidation pathway is inducible. The mRNA populations of a control culture and a cyclohexanone-induced culture were



compared using a technique based on the random amplification of DNA fragments by reverse transcription followed by PCR.

#### Isolation of Total Cellular RNA

The cyclohexanone oxidation pathway was induced by addition of  
5 0.1% cyclohexanone into one of two "split" 10 ml cultures of *Brevibacterium* sp. HCU grown in S12 medium. Each culture was chilled rapidly in an ice-water bath and transferred to a 15 ml tube. Cells were collected by centrifugation for 2 min at 12,000 x g in a rotor chilled to -4°C. The supernatants were discarded, the pellets resuspended in 0.7 ml of  
10 ice-cold solution of 1% SDS and 100 mM sodium acetate at pH 5 and transferred to a 2 ml tube containing 0.7 ml of aqueous phenol pH 5 and 0.3 ml of 0.5 mm zirconia beads (Biospec Products, Bartlesville, OK). The tubes were placed in a bead beater (Biospec) and disrupted at 2,400 beats per min for two min.

15 Following the disruption of the cells, the liquid phases of the tubes were transferred to new microfuge tubes and the phases separated by centrifugation for 3 min at 15,000 x g. The aqueous phase containing total RNA was extracted twice more with phenol at pH 5 and twice with a mixture of phenol/chloroform/isoamyl alcohol pH 7.5 until a precipitate was  
20 no longer visible at the phenol/water interface. Nucleic acids were then recovered from the aqueous phase by ethanol precipitation with three volumes of ethanol and the pellet resuspended in 0.5 ml of diethyl pyrocarbonate (DEPC) treated water. DNA was digested by 6 units of RNase-free DNase (Boehringer Mannheim, Indianapolis, IN) for 1 hr at  
25 37°C. The total RNA solution was then extracted twice with phenol/chloroform/isoamyl alcohol pH 7.5, recovered by ethanol precipitation and resuspended in 1 ml of DEPC treated water to an approximate concentration of 0.5 mg per ml.

#### Generation of RAPDs Patterns From Arbitrarily Reverse-

#### 30 Transcribed Total RNA

Arbitrarily amplified DNA fragments were generated from the total RNA of control and induced cells by following the protocol described by Wong K.K. *et al.* (*Proc Natl Acad Sci U S A.* 91:639 (1994)). A series of parallel reverse transcription (RT)/PCR amplification experiments were  
35 performed using a RT-PCR oligonucleotide set. This set consisted of 81 primers, each designed with the sequence CGGAGCAGATCGAVVVV (SEQ ID NO:63) where VVVV represent all the combinations of the three bases A, G and C at the last four positions of the 3'-end.

The series of parallel RT-PCR amplification experiments were performed on the total RNA from the control and induced cells, each using a single RT-PCR oligonucleotide. Briefly, 50  $\mu$ l reverse transcription (RT) reactions were performed on 20-100 ng of total RNA using 100 U

5 Moloney Murine Leukemia Virus (MMLV) reverse transcriptase (Promega, Madison, WI) with 0.5 mM of each dNTP and 1 mM for each oligonucleotide primer. Reactions were prepared on ice and incubated at 37°C for 1 hr.

Five  $\mu$ l from each RT reaction were then used as template in a

10 50  $\mu$ l PCR reaction containing the same primer used for the RT reaction (0.25  $\mu$ M), dNTPs (0.2 mM each), magnesium acetate (4 mM) and 2.5 U of the Taq DNA polymerase Stoffel fragment (Perkin Elmer, Foster City, CA). The following temperature program was used: 94°C (5 min), 40°C (5 min), 72°C (5 min) for 1 cycle followed by 40 cycles of 94°C (1 min),

15 60°C (1 min), 72°C (5 min).

RAPD fragments were separated by electrophoresis on acrylamide gels (15 cm x 15 cm x 1.5 mm, 6% acrylamide, 29:1 acryl:bisacrylamide, 100 mM Tris, 90 mM borate, 1 mM EDTA pH 8.3). Five  $\mu$ l from each PCR reaction were analyzed with the reactions from the control and the induced

20 RNA for each primer running side by side. Electrophoresis was performed at 1 V/cm. DNA fragments were visualized by silver staining using the Plus One® DNA silver staining kit in the Hoefer automated gel stainer (Amersham Pharmacia Biotech, Piscataway, NJ).

#### Reamplification of the Differentially Expressed DNA

25 Stained gels were rinsed extensively for one hr with distilled water. Bands generated from the RNA of cyclohexanone induced cells but absent in the reaction from the RNA of control cells were excised from the gel and placed in a tube containing 50  $\mu$ l of 10 mM KCl and 10 mM Tris-HCl (pH 8.3) and heated to 95°C for 1 hr to allow some of the DNA to

30 diffuse out of the gel. Serial dilutions of the eluate over a 200 fold range were used as template for a new PCR reaction using the Taq polymerase. The primer used for each reamplification (0.25  $\mu$ M) was the one that had generated the pattern.

Each reamplified fragment was cloned into the blue/white cloning

35 vector pCR2.1 (Invitrogen, San Diego, CA) and sequenced using the universal forward and reverse primers (M13 Reverse Primer (SEQ ID NO:64) and M13 (-20) Forward Primer (SEQ ID NO:65)).

Extension of monooxygenase fragments by Out-PCR.

Kilobase-long DNA fragments extending the sequences fragments identified by differential display were generated by "Out-PCR", a PCR technique using an arbitrary primer in addition to a sequence specific primer. The first step of this PCR-based gene walking technique consisted of randomly copying the chromosomal DNA using a primer of arbitrary sequence in a single round of amplification under low stringency conditions. The primers used for Out-PCR were chosen from a primer set used for mRNA differential display and their sequences were

10 CCGAGCAGATCGAVVVV (SEQ ID NO:63) where VVVV was A, G or C. Ten Out-PCR reactions were performed, each using one primer of arbitrary sequence. The reactions (50 µl) included a 1X concentration of the rTth XL buffer provided by the manufacturer (Perkin-Elmer, Foster City, CA), 1.2 mM magnesium acetate, 0.2 mM of each dNTP, 10-100 ng

15 genomic DNA, 0.4 mM of one arbitrary primer and 1 unit of rTth XL polymerase (Perkin-Elmer). A five min annealing (45°C) and 15 min extension cycle (72°C) lead to the copying of the genomic DNA at arbitrary sites and the incorporation of a primer of arbitrary but known sequence at the 3' end.

20 After these initial low stringency annealing and replication steps, each reaction was split into two tubes. One tube received a specific primer (0.4 mM) designed against the end of the sequence to be extended and directed outward, while the second tube received water and was used as a control. Thirty additional PCR cycles were performed under higher

25 stringency conditions with denaturation at 94°C (1 min), annealing at 60°C (0.5 min) and extension at 72°C (10 min). The long extension time was designed to allow for the synthesis of long DNA fragments by the long range rTth XL DNA polymerase. The products of each pair of reactions were analyzed in adjacent lanes on an agarose gel.

30 Bands present in the sample having received the specific primer but not in the control sample were excised from the agarose gel, melted in 0.5 ml H<sub>2</sub>O and used as the template in a new set of PCR reactions. A 1X concentration of rTth XL buffer, 1.2 mM magnesium acetate, 0.2 mM of each dNTP, 0.4 mM of primers, 1/1000 dilution of the melted slice and

35 1 unit of rTth XL polymerase were used for these reactions. The PCR was performed at 94°C (1 min), 60°C (0.5 min), and 72°C (15 min) per cycle for 20 cycles. For each of these reamplification reactions, two control reactions, lacking either the arbitrary primer or the specific primer, were

included in order to confirm that the reamplification of the band of interest required both the specific and arbitrary primer. DNA fragments that required both the specific and arbitrary primer for amplification were sequenced. For sequencing, the long fragments obtained by Out-PCR  
5 were partially digested with *Mbol* and cloned into pCR2.1 (Invitrogen, Carlsbad, CA). Sequences for these partial fragments were obtained using primers designed against the vector sequence.

### EXAMPLE 3

#### Isolation of a *Acidovorax* sp. CHX Monooxygenase Gene Involved in 10 Degradation of Cyclohexane

This Example describes the isolation of the cyclohexane degrader *Acidovorax* sp. CHX. Discovery of a BVMO gene was accomplished using differential display methods.

#### Strain Isolation

15 An enrichment for bacteria growing on cyclohexane as a sole carbon source was started by adding 5 ml of an industrial wastewater sludge to 20 ml of mineral medium (50 mM KHPO<sub>4</sub> (pH 7.0), 10 mM (NH<sub>4</sub>)SO<sub>4</sub>, 2 mM MgCl<sub>2</sub>, 0.7 mM CaCl<sub>2</sub>, 50 μM MnCl<sub>2</sub>, 1 μM FeCl<sub>3</sub>, 1 μM ZnCl<sub>3</sub>, 1.72 μM CuSO<sub>4</sub>, 2.53 μM CoCl<sub>2</sub>, 2.42 μM Na<sub>2</sub>MoO<sub>2</sub>, and 0.0001%  
20 FeSO<sub>4</sub>) in a 125 ml Erlenmeyer flask sealed with a Teflon lined screw cap. A test tube containing 1 ml of a mixture of mineral oil and cyclohexane (8/1 v/v) was fitted in the flask to provide a low vapor pressure of cyclohexane (approximately 30% of the vapor pressure of pure cyclohexane). The enrichment was incubated at 30°C for a week.  
25 Periodically, 1 to 10 dilutions of the enrichment were performed in the same mineral medium supplemented with 0.005% of yeast extract under low cyclohexane vapors. After several transfers, white flocks could be seen in the enrichments under cyclohexane vapors. If cyclohexane was omitted, the flocks did not grow.

30 After several transfers, the flocks could be grown with 4 μl of liquid cyclohexanone added directly to 10 ml of medium. To isolate colonies, flocks were washed in medium and disrupted by thorough shaking in a bead beater. The cells released from the disrupted flocks were streaked onto R2A medium agar plates and incubated under cyclohexane vapors.  
35 Pinpoint colonies were picked under a dissecting microscope and inoculated in 10 ml of mineral medium supplemented with 0.01% yeast extract and 4 μl of cyclohexane. The flocks were grown, disrupted and streaked again until a pure culture was obtained.

Taxonomic identification of this isolate was performed by PCR amplification of 16S rDNA, as described in the General Methods. The 16S rRNA gene sequence from strain CHX was at least 98% homologous to the 16S rRNA gene sequence of an uncultured bacterium (Seq.

- 5 Accession number AF143840) and 95% homologous to the 16s rRNA gene sequences of the genus *Acidovorax temperans* (Accession number AF078766). The complete 16s DNA sequence of the isolated *Acidovorax* sp. CHX is shown as SEQ ID NO:5.

#### Induction of Cyclohexane Degradation Genes

- 10 For induction of cyclohexane degradation genes, colonies of *Acidovorax* sp. CHX were scraped from an R2A agar plate and inoculated into 25 ml R2A broth. This culture was incubated overnight at 30°C. The next day 25 ml of fresh R2A broth was added and growth was continued for 15 min. The culture was split into two separate flasks, each of which  
15 received 25 ml. To one of these flasks, 5 µl of pure cyclohexane was added to induce expression of cyclohexane degradation genes. The other flask was kept as a control. Differential display was used to identify the *Acidovorax* sp. CHX monooxygenase gene. Identification of cyclohexane induced gene sequences and sequencing cyclohexanone oxidation genes  
20 from strains was performed in a similar manner as described in Example 1.

#### EXAMPLE 4

##### Isolation of a *Acinetobacter* sp. SE19 Monooxygenase Gene Involved in Degradation of Cyclohexanol

- 25 This Example describes the isolation of the cyclohexanol degrader *Acinetobacter* sp. SE19. Discovery of a BV monooxygenase gene was accomplished by screening of cosmid libraries, followed by sequencing of shot-gun libraries.

##### Isolation of Strain

- 30 An enrichment for bacteria that grow on cyclohexanol was isolated from a cyclopentanol enrichment culture. The enrichment culture was established by inoculating 1 mL of activated sludge into 20 mL of S12 medium (10 mM ammonium sulfate, 50 mM potassium phosphate buffer (pH 7.0), 2 mM MgCl<sub>2</sub>, 0.7 mM CaCl<sub>2</sub>, 50 µM MnCl<sub>2</sub>, 1 µM FeCl<sub>3</sub>, 1 µM  
35 ZnCl<sub>3</sub>, 1.72 µM CuSO<sub>4</sub>, 2.53 µM CoCl<sub>2</sub>, 2.42 µM Na<sub>2</sub>MoO<sub>2</sub>, and 0.0001% FeSO<sub>4</sub>) in a sealed 125 mL screw-cap Erlenmeyer flask. The enrichment culture was supplemented with 100 ppm cyclopentanol added directly to the culture medium and was incubated at 35°C with reciprocal shaking.

The enrichment culture was maintained by adding 100 ppm cyclopentanol every 2-3 days. The culture was diluted every 2-10 days by replacing 10 mL of the culture with the same volume of S12 medium. After 15 days of incubation, serial dilutions of the enrichment culture were spread onto  
5 LB plates. Single colonies were screened for the ability to grow on S12 liquid with cyclohexanol as the sole carbon and energy source. The cultures were grown at 35°C in sealed tubes. One of the isolates, strain SE19 was selected for further characterization.

The 16s rRNA genes of SE19 isolates were amplified by PCR  
10 according to the procedures of the General Methods. Result from all isolates showed that strain SE19 has close homology to *Acinetobacter haemolyticus* and *Acinetobacter junii*, (99% nucleotide identity to each).

#### Construction Of *Acinetobacter* Cosmid Libraries

*Acinetobacter* sp. SE19 was grown in 25 ml LB medium for 6 h at  
15 37°C with aeration. Bacterial cells were centrifuged at 6,000 rpm for 10 min in a Sorvall RC5C centrifuge at 4°C. Supernatant was decanted and the cell pellet was frozen at -80°C. Chromosomal DNA was prepared as outlined below with special care taken to avoid shearing of DNA. The cell pellet was gently resuspended in 5 ml of 50 mM Tris-10 mM EDTA  
20 (pH 8) and lysozyme was added to a final concentration of 2 mg/ml. The suspension was incubated at 37°C for 1 h. Sodium dodecyl sulfate was then added to a final concentration of 1% and proteinase K was added at 100 µg/ml. The suspension was incubated at 55°C for 2 h. The suspension became clear and the clear lysate was extracted with equal  
25 volume of phenol:chloroform:isoamyl alcohol (25:24:1). After centrifuging at 12,000 rpm for 20 min, the aqueous phase was carefully removed and transferred to a new tube. Two volumes of ethanol were added and the DNA was gently spooled with a sealed glass pasteur pipet. The DNA was dipped into a tube containing 70% ethanol. After air drying, the DNA was  
30 resuspended in 400 µl of TE (10 mM Tris-1 mM EDTA, pH 8) with RNaseA (100 µg/ml) and stored at 4°C. The concentration and purity of DNA was determined spectrophotometrically by OD<sub>260</sub>/OD<sub>280</sub>. A diluted aliquot of DNA was run on a 0.5% agarose gel to determine the intact nature of DNA.

35 Chromosomal DNA was partially digested with *Sau3AI* (GIBRO/BRL, Gaithersburg, MD) as outlined by the instruction manual for the SuperCos 1 Cosmid Vector Kit. DNA (10 µg) was digested with 0.5 unit of *Sau3AI* at room temperature in 100 µl of reaction volume. Aliquots

of 20 µl were withdrawn at various time points of the digestion: e.g., 0, 3, 6, 9, 12 min. DNA loading buffer was added and samples were analyzed on a 0.5% agarose gel to determine the extent of digestion. A decrease in size of chromosomal DNA corresponded to an increase in the length of time for *Sau3AI* digestion. The preparative reaction was performed using 50 µg of DNA digested with 1 unit of *Sau3AI* for 3 min at room temperature. The digestion was terminated by addition of 8 mM of EDTA. The DNA was extracted once with phenol:chloroform:isoamyl alcohol and once with chloroform. The aqueous phase was adjusted to 0.3 M NaOAc and ethanol precipitated. The partially digested DNA was dephosphorylated with calf intestinal alkaline phosphatase and ligated to SuperCos 1 vector, which had been treated according to the instructions in the SuperCos 1 Cosmid Vector Kit. The ligated DNA was packaged into lambda phage using Gigapack III XL packaging extract, as recommended by Stratagene (manufacturer's instructions were followed). The packaged *Acinetobacter* genomic DNA library contained a phage titer of  $5.6 \times 10^4$  colony forming units per µg of DNA as determined by transfecting *E. coli* XL1-Blue MR. Cosmid DNA was isolated from six randomly chosen *E. coli* transformants and found to contain large inserts of DNA (25-40kb).

Identification and Characterization of Cosmid Clones Containing a Cyclohexanone Monooxygenase Gene

The cosmid library of *Acinetobacter* sp. SE19 was screened based on the homology of the cyclohexanone monooxygenase gene. Two primers, monoL: GAGTCTGAGCATATGTCACAAAAAATGGATTTTG (SEQ ID NO:66) and monoR: GAGTCTGAGGGATCCTTAGGCATTGGCAGGTTGCTTGAT (SEQ ID NO:67) were designed based on the published sequence of cyclohexanone monooxygenase gene of *Acinetobacter* sp. NCIB 9871. The cosmid library was screened by PCR using monoL and monoR primers. Five positive clones (5B12, 5F5, 8F6, 14B3 and 14D7) were identified among about 1000 clones screened. They all contain inserts of 35-40 kb that show homology to the cyclohexanone monooxygenase gene amplified by monoL and monoR primers. Southern hybridization using this gene fragment as a probe indicated that the cosmid clone 5B12 has about 20kb region upstream of the monooxygenase gene and cosmid clone 8F6 has about 30kb downstream of the monooxygenase gene.

Cosmid clone 14B3 contains rearranged *Acinetobacter* DNA adjacent to the monooxygenase gene.

#### Construction of shot-gun sequencing libraries

Shot gun libraries of 5B12 and 8F6 were constructed. Cosmid DNA was sheared in a nebulizer (Inhalation Plastics Inc., Chicago, IL) at 20 psi for 45 sec and the 1-3 kb portion was gel purified. Purified DNA was treated with T4 DNA polymerase and T4 polynucleotide kinase following manufacturer's (GIBCO/BRL) instructions. Polished inserts were ligated into pUC18 vectors using Ready-To-Go pUC18SmaI/BAP+Ligase (GIBCO/BRL). The ligated DNA was transformed into *E. coli* DH5 $\alpha$  cells and plated on LB with ampicillin and X-gal. A majority of the transformants were white and those containing inserts were sequenced with the universal and reverse primers of pUC18 by standard sequencing methods.

Shot gun library inserts were sequenced with pUC18 universal and reverse primers. Sequences of 200-300 clones from each library were assembled using Sequencer 3.0 program. A contig of 17419 bp containing the cyclohexanone monooxygenase gene was formed.

#### EXAMPLE 5

##### Isolation and Sequencing of *Rhodococcus erythropolis* AN12

This Example describes isolation of *Rhodococcus erythropolis* AN12 strain from wastestream sludge. A shotgun sequencing strategy approach permitted sequencing of the entire microbial genome.

##### Isolation of *Rhodococcus erythropolis* AN12

Strain AN12 of *Rhodococcus erythropolis* was isolated on the basis of ability to grow on aniline as the sole source of carbon and energy. Bacteria that grow on aniline were isolated from an enrichment culture. The enrichment culture was established by inoculating 1 ml of activated sludge into 10 ml of S12 medium (10 mM ammonium sulfate, 50 mM potassium phosphate buffer (pH 7.0), 2 mM MgCl<sub>2</sub>, 0.7 mM CaCl<sub>2</sub>, 50  $\mu$ M MnCl<sub>2</sub>, 1  $\mu$ M FeCl<sub>3</sub>, 1  $\mu$ M ZnCl<sub>2</sub>, 1.72  $\mu$ M CuSO<sub>4</sub>, 2.53  $\mu$ M CoCl<sub>2</sub>, 2.42  $\mu$ M Na<sub>2</sub>MoO<sub>4</sub>, and 0.0001% FeSO<sub>4</sub>) in a 125 ml screw cap Erlenmeyer flask. The activated sludge was obtained from a DuPont wastewater treatment facility. The enrichment culture was supplemented with 100 ppm aniline added directly to the culture medium and was incubated at 25°C with reciprocal shaking. The enrichment culture was maintained by adding 100 ppm of aniline every 2-3 days. The culture was diluted every 14 days by replacing 9.9 ml of the culture with the same volume of S12 medium.



Bacteria that utilize aniline as a sole source of carbon and energy were isolated by spreading samples of the enrichment culture onto S12 agar. Aniline was placed on the interior of each petri dish lid. The petri dishes were sealed with parafilm and incubated upside down at room temperature (25°C). Representative bacterial colonies were then tested for the ability to use aniline as a sole source of carbon and energy. Colonies were transferred from the original S12 agar plates used for initial isolation to new S12 agar plates and supplied with aniline on the interior of each petri dish lid. The petri dishes were sealed with parafilm and incubated upside down at room temperature (25°C).

A 16S rRNA gene of strain AN12 was sequenced (SEQ ID NO:6) as described in the General Methods and compared to other 16S rRNA sequences in the GenBank sequence database. The 16S rRNA gene sequence from strain AN12 was at least 98% homologous to the 16S rRNA gene sequences of high G + C Gram positive bacteria belonging to the genus *Rhodococcus*.

#### Preparation of Genomic DNA for Sequencing and Sequence Generation

Genomic DNA and library construction were prepared according to published protocols (Fraser *et al. Science* 270(5235): 397-403 (1995)). A cell pellet was resuspended in a solution containing 100 mM Na-EDTA (pH 8.0), 10 mM Tris-HCl (pH 8.0), 400 mM NaCl, and 50 mM MgCl<sub>2</sub>.

Genomic DNA preparation After resuspension, the cells were gently lysed in 10% SDS, and incubated for 30 minutes at 55°C. After incubation at room temperature, proteinase K (Boehringer Mannheim, Indianapolis, IN) was added to 100 µg/ml and incubated at 37°C until the suspension was clear. DNA was extracted twice with Tris-equilibrated phenol and twice with chloroform. DNA was precipitated in 70% ethanol and resuspended in a solution containing 10 mM Tris-HCl and 1 mM Na-EDTA (TE buffer) pH 7.5. The DNA solution was treated with a mix of RNAases, then extracted twice with Tris-equilibrated phenol and twice with chloroform. This was followed by precipitation in ethanol and resuspension in TE buffer.

Library construction 200 to 500 µg of chromosomal DNA was resuspended in a solution of 300 mM sodium acetate, 10 mM Tris-HCl, 1 mM Na-EDTA, and 30% glycerol, and sheared at 12 psi for 60 sec in an Aeromist Downdraft Nebulizer chamber (IBI Medical products, Chicago, IL). The DNA was precipitated, resuspended and treated with Bal31 nuclease (New England Biolabs, Beverly, MA). After size fractionation, a

fraction (2.0 kb, or 5.0 kb) was excised, cleaned and a two-step ligation procedure was used to produce a high titer library with greater than 99% single inserts.

- 5 Sequencing A shotgun sequencing strategy approach was adopted for the sequencing of the whole microbial genome (Fleischmann, R. *et al.* Whole-Genome Random sequencing and assembly of *Haemophilus influenzae* Rd. *Science* 269(5223): 496-512 (1995)).

#### EXAMPLE 6

##### Identification and Characterization of Bacterial Genes

- 10 Genes encoding each monooxygenase were identified by conducting BLAST (Basic Local Alignment Search Tool; Altschul, S. F., *et al.*, (1993) *J. Mol. Biol.* 215:403-410; see also [www.ncbi.nlm.nih.gov/BLAST/](http://www.ncbi.nlm.nih.gov/BLAST/)) searches for similarity to sequences contained in the BLAST "nr" database (comprising all non-redundant  
15 GenBank CDS translations, sequences derived from the 3-dimensional structure Brookhaven Protein Data Bank, the SWISS-PROT protein sequence database, EMBL, and DDBJ databases). The sequences obtained in Examples 1, 2, 3, 4, and 5 were analyzed for similarity to all publicly available DNA sequences contained in the "nr" database using the  
20 BLASTN algorithm provided by the National Center for Biotechnology Information (NCBI). The DNA sequences were translated in all reading frames and compared for similarity to all publicly available protein sequences contained in the "nr" database using the BLASTX BLOSUM62 algorithm with a gap existense cost of 11 per residue gap cost of 2,  
25 filtered, gap alignment (Gish, W. and States, D. J. *Nature Genetics* 3:266-272 (1993)) provided by the NCBI.

- All comparisons were done using either the BLASTNnr or BLASTXnr algorithm. The results of the BLAST comparisons are given in Table 3 which summarize the sequence to which each sequence has the  
30 most similarity. Table 3 displays data based on the BLASTXnr algorithm with values reported in expect values. The Expect value estimates the statistical significance of the match, specifying the number of matches, with a given score, that are expected in a search of a database of this size absolutely by chance.

35

TABLE 3

ORF Name	Gene Name and Organism of Isolation	Similarity Identified	SEQ ID base	SEQ ID Peptide	% Identity <sup>a</sup>	% Similarity <sup>b</sup>	E-value <sup>c</sup>	Citation
1	<i>chnB</i> <i>Rhodococcus</i> sp. phi 1	>gb AAG10021.1 AF282240_5 (AF282240) cyclohexanone monooxygenase [Acinetobacter sp. SE19]	7	8	55	71	e-174	Cheng, Q., et al. J. Bacteriol. 182: 4744- 4751 (2000)
2	<i>chnB</i> <i>Rhodococcus</i> sp. phi 2	>gb AAG10021.1 AF282240_5 (AF282240) cyclohexanone monooxygenase [Acinetobacter sp. SE19]	9	10	53	67	e-163	Cheng, Q., et al. J. Bacteriol. 182: 4744- 4751 (2000)
3	<i>chnB</i> <i>Arthrobacter</i> sp. BP2	>gb AAG10021.1 AF282240_5 (AF282240) cyclohexanone monooxygenase [Acinetobacter sp. SE19]	11	12	57	72	e-106	Cheng, Q., et al. J. Bacteriol. 182: 4744- 4751 (2000)
4	<i>chnB1</i> <i>Brevibacteriu</i> m sp. HCU	>pir JC7158 steroid monooxygenase (EC 1.14.99.-) - <i>Rhodococcus</i> rhodochrous dbj BAA24454.1  (AB010439) steroid monooxygenase [ <i>Rhodococcus rhodochrous</i> ]	13	14	44	59	e-122	Moril, S., et al. J. Biochem. 126 (3): 624- 631 (1999)
5	<i>chnB2</i> <i>Brevibacteriu</i> m sp. HCU	>pir JC7158 steroid monooxygenase (EC 1.14.99.-) - <i>Rhodococcus</i> rhodochrous dbj BAA24454.1  (AB010439) steroid monooxygenase [ <i>Rhodococcus rhodochrous</i> ]	15	16	38	53	2e-94	Moril, S., et al. J. Biochem. 126 (3): 624- 631 (1999)
6	<i>chnB</i> <i>Acidovorax</i> sp. CHX	>gb AAG10021.1 AF282240_5 (AF282240) cyclohexanone monooxygenase [Acinetobacter sp. SE19]	17	18	57	73	0.0	Cheng, Q., et al. J. Bacteriol. 182: 4744- 4751 (2000)

ORF Name	Gene Name and Organism of Isolation	Similarity Identified	SEQ ID base	SEQ ID Peptide	% Identity <sup>a</sup>	% Similarity <sup>b</sup>	E-value <sup>c</sup>	Citation
7	<i>chnB</i> <i>Acinetobacter</i> sp. SE19	>dbj BAA86293.1  (AB006902) cyclohexanone 1,2-monooxygenase [ <i>Acinetobacter</i> sp.] dbj BAB61738.1  (AB026668) cyclohexanone 1,2- monooxygenase [ <i>Acinetobacter</i> sp. NCIMB9871]	19	20	99	99	0.0	Chen, Y.C., et al. <i>J. Bacteriol.</i> 170 (2): 781-789 (1988)
8	ORF 8 <i>chnB</i> <i>Rhodococcus</i> <i>erythropolis</i> AN12	>pir T37052 probable flavin-containing monooxygenase - <i>Streptomyces</i> coelicolor emb CAB52349.1  (AL109747) putative flavin-containing monooxygenase [ <i>Streptomyces</i> coelicolor A3(2)]	21	22	37	50	6e-58	Seeger, K.J., et al. Direct Submission (??- AUG-1999) to the EMBL Data Library
9	ORF 9 <i>chnB</i> <i>Rhodococcus</i> <i>erythropolis</i> AN12	>emb CAB59668.1  (AL132674) monooxygenase. [ <i>Streptomyces</i> coelicolor A3(2)]	23	24	44	61	e-118	Redenbach, M., et al. <i>Mol. Microbiol.</i> 21 (1): 77-96 (1996)
10	ORF 10 <i>chnB</i> <i>Rhodococcus</i> <i>erythropolis</i> AN12	>pir JC7158 steroid monooxygenase (EC 1.14.99.-) - <i>Rhodococcus</i> rhodochrous dbj BAA24454.1  (AB010439) steroid monooxygenase [ <i>Rhodococcus</i> rhodochrous]	25	26	64	76	0.0	Morii, S., et al. <i>J. Biochem.</i> 126 (3), 624-631 (1999)
11	ORF 11 <i>chnB</i> <i>Rhodococcus</i> <i>erythropolis</i> AN12	>gb AAK22759.1  (AE005753) monooxygenase, flavin-binding family [ <i>Caulobacter crescentus</i> ]	27	28	65	74	e-176	Nierman, W.C., et al. <i>Proc. Natl. Acad. Sci. U.S.A.</i> 98 (7): 4136-4141 (2001)

ORF Name	Gene Name and Organism of Isolation	Similarity Identified	SEQ ID base	SEQ ID Peptide	% Identity <sup>a</sup>	% Similarity <sup>b</sup>	E-value <sup>c</sup>	Citation
12	ORF 12 chnB Rhodococcus erythropolis AN12	>emb CAB59668.1  (AL132674) monooxygenase. [Streptomyces coelicolor A3(2)]	29	30	45	63	e-124	Redenbach, M., et al. <i>Mol. Microbiol.</i> 21 (1): 77-96 (1996)
13	ORF 13 chnB Rhodococcus erythropolis AN12	>gb AAK24539.1  (AE005925) monooxygenase, flavin-binding family [Caulobacter crescentus]	31	32	55	68	e-159	Nierman, W.C., et al. <i>Proc. Natl. Acad. Sci. U.S.A.</i> 98 (7): 4136-4141 (2001)
14	ORF 14 chnB Rhodococcus erythropolis AN12	>pir JC7158 steroid monooxygenase (EC 1.14.99.-) - Rhodococcus rhodochrous dbj BAA24454.1  (AB010439) steroid monooxygenase [Rhodococcus rhodochrous]	33	34	51	65	e-154	Morii, S., et al. <i>J. Biochem.</i> 126 (3), 624-631 (1999)
15	ORF 15 chnB Rhodococcus erythropolis AN12	>sp P55487 Y4ID_RHISN PROBABLE MONOOXYGENASE Y4ID gb AAB91699.1  (AE000078) Y4ID [Rhizobium sp. NGR234]	35	36	39	58	e145	Freiberg, C.A., et al. <i>Nature</i> 387: 394-401 (1997).
16	ORF 16 chnB Rhodococcus erythropolis AN12	>pir A83453 probable flavin-containing monooxygenase PA1538 [imported] - Pseudomonas aeruginosa (strain PAO1) gb AAG04927.1 AE004582_5 (AE004582) probable flavin-containing monooxygenase [Pseudomonas aeruginosa]	37	38	43	59	e-119	Stover, C.K., et al. <i>Nature</i> 406 (6799): 959-964 (2000)

ORF Name	Gene Name and Organism of Isolation	Similarity Identified	SEQ ID base	SEQ ID Peptide	% Identity <sup>a</sup>	% Similarity <sup>b</sup>	E-value <sup>c</sup>	Citation
17	ORF 17 chnB Rhodococcus erythropolis AN12	>pir [G70852 hypothetical protein Rv3083 - Mycobacterium tuberculosis (strain H37RV) emb CAA16141.1  (AL021309) hypothetical protein Rv3083 [Mycobacterium tuberculosis] gb AAK47504.1  (AE007134) monooxygenase, flavin-binding family [Mycobacterium tuberculosis CDC1551]	39	40	53	70	e-150	Cole, S.T., et al. <i>Nature</i> 393 (6685): 537-544 (1998)
18	ORF 18 chnB Rhodococcus erythropolis AN12	>pir [A83453 probable flavin-containing monooxygenase PA1538 [imported] - Pseudomonas aeruginosa (strain PAO1) gb AAG04927.1 AE004582_5 (AE004582) probable flavin-containing monooxygenase [Pseudomonas aeruginosa]	41	42	44	60	e-117	Stover, C.K., et al. <i>Nature</i> 406 (6799): 959-964 (2000)
19	ORF 19 chnB Rhodococcus erythropolis AN12	>gb AAG10021.1 AF282240_5 (AF282240) cyclohexanone monooxygenase [Acinetobacter sp. SE19]	43	44	54	69	e-168	Cheng, Q., et al. <i>J. Bacteriol.</i> 182 (17): 4744-4751 (2000)
20	ORF 20 chnB Rhodococcus erythropolis AN12	>pir [JC7158 steroid monooxygenase (EC 1.14.99.-) - Rhodococcus rhodochrous dbj BAA24454.1  (AB010439) steroid monooxygenase [Rhodococcus rhodochrous]	45	46	42	60	e-123	Moril, S., et al. <i>J. Biochem.</i> 126 (3): 624-631 (1999)

<sup>a</sup>%Identity is defined as percentage of amino acids that are identical between the two proteins.

<sup>b</sup>% Similarity is defined as percentage of amino acids that are identical or conserved between the two proteins.

<sup>c</sup>Expect value. The Expect value estimates the statistical significance of the match, specifying the number of matches, with a given score, that are expected in a search of a database of this size absolutely by chance.

EXAMPLE 7Cloning and Expression Of Monooxygenase Genes into *Escherichia coli*

This example illustrates the expression in *E. coli* of isolated full length BVMO genes from *Brevibacterium* sp. HCU, *Acinetobacter* SE19, *Rhodococcus* sp. phi1, *Rhodococcus* sp. phi2, *Arthrobacter* sp. BP2 and *Acidovorax* sp. CHX.

Full length BVMO's were PCR amplified, using chromosomal DNA as the template and the primers shown below in Table 4.

10

Table 4Primers Used for Amplification of Full-Length BV Monooxygenases

Monooxygenase	Forward Primer	Reverse Primer
<i>Brevibacterium</i> sp. HCU <i>chnB1</i>	atgccaaattacacaacaacttgacc (SEQ ID NO:68)	ctatttcatacccgccgattcac (SEQ ID NO:69)
<i>Brevibacterium</i> sp. HCU <i>chnB2</i>	atgacgtcaaccaatgcctgcac (SEQ ID NO:70)	cacttaagtcgcattcagccc (SEQ ID NO:71)
<i>Acinetobacter</i> sp. SE19 <i>chnB</i>	atggatttgatgctatcgtg (SEQ ID NO:72)	ggcattggcaggttgctg (SEQ ID NO:73)
<i>Arthrobacter</i> sp. BP2 <i>chnB</i>	atgactgcacagaacactttcc (SEQ ID NO:74)	tcaaagccgcggtatccg (SEQ ID NO:75)
<i>Rhodococcus</i> sp. phi1 <i>chnB</i>	atgactgcacagatctcaccac (SEQ ID NO:76)	tcaggcggtcaccgggacagcg (SEQ ID NO:77)
<i>Rhodococcus</i> sp. phi2 <i>chnB</i>	atgaccgcacagaccatccacac (SEQ ID NO:78)	tcagaccgtgaccatclcg (SEQ ID NO:79)
<i>Acidovorax</i> sp. CHX <i>chnB</i>	atgtctcctcgccaagcagc (SEQ ID NO:80)	cagtggittgaacgcaaagcc (SEQ ID NO:81)

Following amplification, the *chnB* gene fragments were cloned into pTrcHis-TOPO TA vectors with either an N-terminal tail or C-terminal tail, as provided by the vector sequence (N-terminal tail for *Brevibacterium* sp. HCU, *Rhodococcus* sp. phi1, *Rhodococcus* sp. phi2, and *Arthrobacter* sp. BP2 monooxygenases; C-terminal tail for *Acinetobacter* sp. SE19 and *Acidovorax* sp. CHX monooxygenases). These vectors were transformed into *E. coli*, with transformants grown in Luria-Bertani broth supplemented with ampicillin (100 ug/ml) and riboflavin (0.1 ug/ml) at 30°C until the absorbance at 600 nm (A600) reached 0.5. When the A600 was reached, the temperature was shifted to 16°C.

The encoded monooxygenase sequences were expressed upon addition of IPTG to the culture media, 30 min after the temperature shift to 16°C. The cultures were grown further overnight (14 hrs) and harvested by centrifugation in a cold centrifuge. The cells were treated with  
 5 lysozyme (100 mg/ml) for 30 min on ice and sonicated. Following sonication, cell extracts were centrifuged and the supernatant was equilibrated with Ni-NTA resin (Qiagen, Valencia, CA) for 1 hr at 4°C. Protein bound resin was washed successively with increasing concentrations of imidazole buffer until the protein of interest was  
 10 released from the resin. The purified protein was concentrated and the buffer exchanged to remove the imidazole. The protein concentration was adjusted to 1 ug/ml.

#### EXAMPLE 8

Assays of *chnB* Monooxygenase Activities of *Brevibacterium* sp. HCU,  
 15 *Acinetobacter* SE19, *Rhodococcus* sp. phi1, *Rhodococcus* sp. phi2,  
*Arthrobacter* sp. BP2 and *Acidovorax* sp. CHX.

The *chnB* monooxygenase activity of each over-expressed enzyme from Example 7 was assayed against various ketone substrates: cyclobutanone, cyclopentanone, 2-methylcyclopentanone,  
 20 cyclohexanone, 2-methylcyclohexanone, cyclohex-2-ene-1-one, 1,2-cyclohexanedione, 1,3-cyclohexanedione, 1,4-cyclohexanedione, cycloheptanone, cyclooctanone, cyclodecanone, cycloundodecanone, cyclododecanone, cyclotridecanone, cyclopentadecanone, 2-tridecanone, 2-phenylcyclohexanone, diheyl ketone, norcamphor, beta-ionone,  
 25 oxindole, levoglucosenone, dimethyl sulfoxide, dimethyl-2-piperidone, and phenylboronic acid. Compounds were selected on the basis of previous observations by van der Werf (*J. Biochem.* 347:693-701 (2000)) and Miyamoto et al. (*Biochimica et Biophysica Acta* 1251: 115-124 (1995)) and by searches for the ketone substructure.

30 All compounds were obtained from Sigma-Aldrich with only two exceptions. Levoglucosenone was obtained from Toronto Research Chemicals, Inc. and dimethyl-2-piperidone was prepared according to U.S. Patent 6,077,955. For enzyme assays all compounds were dissolved to a concentration of 0.1 M in methanol, with the exceptions of  
 35 norcamphor (dissolved in ethyl acetate), cyclododecanone, cyclotridecanone and cyclopentadecanone (dissolved in propanol), and levoglucosenone (dissolved with acetone).



The monooxygenase activity of each over-expressed enzyme was assayed spectrophotometrically at 340 nm by monitoring the oxidation of NADPH. Assays were performed in individual quartz cuvettes, with a pathlength of 1 cm. The following components were added to the cuvette for the enzyme assays: 380  $\mu$ l of 33.3 mM MES-HEPES-sodium acetate buffer (pH 7.5), 5  $\mu$ l of 0.1 M substrate (1.25 mM final concentration), 10  $\mu$ l of 1  $\mu$ g/ $\mu$ l enzyme solution (10 ng total, 0.025 ng/ $\mu$ l) and 5  $\mu$ l NADPH (1.2 M, 15 mM final concentration). An Ultrospec 4000 (Pharmacia Biotech, Cambridge, England) was used to read the absorbance of the samples over a two to ten minute time period and the SWIFT (Pharmacia Biotech) program was used to calculate the slope of the reduction in absorbance over time. For the *Brevibacterium* sp. HCU *chnB2*, the rates were multiplied by a factor of 3.25 to adjust for decrease in activity due to storage as suggested by the literature (*J. Bacteriol.* 2000. 182: p.4241-4248). Monooxygenase activity of each over-expressed enzyme is shown in Table 5, with respect to each ketone substrate. The specific activity values listed are given in  $\mu$ mol/min/mg. The notation "ND" refers to "No Activity Detected".

Graphical representation of the data shown in Table 5 is also provided in Figures 1, 2, 3, 4, and 5.

**Table 5**  
**Specific Activity of Monooxygenase Enzymes Against Various Ketone Substrates**

Compound	Species						
	sp. HCU	sp. HCU	sp. SE19	sp. BP2	sp. CHX	sp. phi1	sp. phi2
	<i>chnB1</i>	<i>chnB2</i>	<i>chnB</i>	<i>chnB</i>	<i>chnB</i>	<i>chnB</i>	<i>chnB</i>
Norcamphor	0.410	1.331	4.474	2.842	0.166	1.504	2.816
Cyclobutanone	ND	0.374	0.109	0.128	ND	0.102	0.154
Cyclopentanone	ND	1.331	3.034	1.491	0.621	1.370	2.451
2-methyl-cyclopentanone	1.395	0.874	8.378	3.514	0.627	3.392	6.445
Cyclohexanone	2.765	1.726	6.349	3.565	0.397	3.680	3.750

Compound	Species						
	sp. HCU	sp. HCU	sp. SE19	sp. BP2	sp. CHX	sp. phi1	sp. phi2
	<i>chnB1</i>	<i>chnB2</i>	<i>chnB</i>	<i>chnB</i>	<i>chnB</i>	<i>chnB</i>	<i>chnB</i>
2-methyl-cyclohexanone	2.714	1.622	9.990	4.205	0.627	4.774	5.952
Cyclohex-2-ene-1-one	0.435	0.541	5.357	2.739	0.666	2.694	3.091
1,2-cyclohexanedione	0.787	0.416	0.077	0.237	0.096	0.083	ND
1,3-cyclohexanedione	0.237	0.978	0.237	0.397	0.032	ND	0.141
1,4-cyclohexanedione	3.405	1.123	8.346	3.994	0.794	3.302	6.150
Cycloheptanone	0.646	0.374	8.422	3.846	0.608	3.622	6.234
Cyclooctanone	ND	ND	1.984	0.646	0.410	0.627	0.141
Cyclodecanone	ND	ND	0.320	0.166	0.160	0.077	0.205
Cycloundecanone	ND	0.125	0.064	0.064	0.058	ND	0.051
Cyclododecanone	ND	0.229	0.122	0.198	0.051	ND	0.122
Cyclotridecanone	ND	ND	0.166	0.147	ND	ND	0.109
Cyclopentadecanone	ND	ND	0.109	0.122	ND	0.122	ND
2-tridecanone	ND	0.187	ND	ND	0.096	0.160	1.690
dihexyl ketone	ND	0.270	ND	ND	ND	0.160	ND
2-phenyl-cyclohexanone	1.459	0.104	5.370	ND	0.192	1.050	0.730
Oxindole	2.438	0.229	7.091	4.845	0.307	3.411	4.858
Levoglucosenone	ND	ND	1.126	0.525	0.147	0.461	0.506

Compound	Species						
	sp.	sp.	sp.	sp.	sp.	sp.	sp.
	HCU <i>chnB1</i>	HCU <i>chnB2</i>	SE19 <i>chnB</i>	BP2 <i>chnB</i>	CHX <i>chnB</i>	phi1 <i>chnB</i>	phi2 <i>chnB</i>
dimethyl sulfoxide	0.230	ND	0.819	0.422	0.358	0.518	0.544
dimethy-2-piperidone	2.822	0.354	8.384	4.154	0.557	3.539	6.509
Phenylboronic acid	1.606	ND	0.102	0.192	ND	ND	0.109
beta-ionone	0.109	0.374	3.347	1.485	0.544	2.707	0.544

**EXAMPLE 9**

**Cloning Of *Rhodococcus erythropolis* AN12 Monooxygenase  
Genes into *Escherichia coli***

5        This example illustrates the construction of a suite of recombinant *E. coli*, each containing a full length BVMOs from *Rhodococcus erythropolis* AN12.

Full length BV monooxygenases were PCR amplified, using chromosomal DNA as the template and the primers shown below in Table

10    6.

**Table 6**  
**Primers Used for Amplification of Full-Length BV *Rhodococcus*  
*erythropolis* AN12 Monooxygenases**

<i>chnB</i> Mono- oxygenase	Forward Primer	Reverse Primer
ORF 8	atg agc aca gag ggc aag tac gc (SEQ ID NO:82)	[tca] gtc ctt gtt cac gta gta ggc c (SEQ ID NO:83)
ORF 9	atg gtc gac atc gac cca acc tc (SEQ ID NO:84)	tta tcg gct cct cac ggt ttc tcg (SEQ ID NO:85)
ORF 10	atg acc gat cct gac ttc tcc acc (SEQ ID NO:86)	tca tgc gtg cac cgc act gtt cag (SEQ ID NO:87)
ORF 11	atg agc ccc tcc ccc ttg ccg ag (SEQ ID NO:88)	tca tgc gcg atc cgc ctt ctc gag (SEQ ID NO:89)

<i>chnB</i> Mono- oxygenase	Forward Primer	Reverse Primer
ORF 12	gtg aac aac gaa tct gac cac ttc (SEQ ID NO:90)	tca tgc ggt gta ctc cgg ttc cg (SEQ ID NO:91)
ORF 13	atg agc acc gaa cac ctc gat g (SEQ ID NO:92)	tca act ctt gct cgg tac cgg cg (SEQ ID NO:93)
ORF 14	atg aca gac gaa ttc gac gta gtg at (SEQ ID NO:94)	tca gct ctg gtt cac agg gac gg (SEQ ID NO:95)
ORF 15	atg gcg gag ata gtc aat ggt cc (SEQ ID NO:96)	tca ccc tcg cgc ggt cgg agt c (SEQ ID NO:97)
ORF 16	gtg aag ctt ccc gaa cat gtc gaa ac (SEQ ID NO:98)	tca tgc ctg gac gct ttc gat ctt g (SEQ ID NO:99)
ORF 17	atg aca cag cat gtc gac gta ctg a (SEQ ID NO:100)	cta tgc gct ggc gac ctt gct atc (SEQ ID NO:101)
ORF 18	atg tca tca cgg gtc aac gac ggc c (SEQ ID NO:102)	tca tcc ttt gcc tgt cgt cag tgc (SEQ ID NO:103)
ORF 19	atg act aca caa aag gcc ctg acc (SEQ ID NO:104)	tca ggc gtc gac ggt gtc ggc c (SEQ ID NO:105)
ORF 20	atg aca act acc gaa tcc aga act c (SEQ ID NO:106)	tca gcg cag att gaa gcc ctt gta tc (SEQ ID NO:107)

- Following amplification, the gene fragments were cloned into pTrcHis-TOPO TA vectors with either an N-terminal tail or C-terminal tail, as provided by the vector sequence. These vectors were transformed into
- 5 *E. coli*, with transformants grown in Luria-Bertani broth supplemented with ampicillin (100 ug/ml).

#### EXAMPLE 10

##### Assays of *chnB* Monooxygenase Activities of *Rhodococcus erythropolis*

##### AN12

- 10 The *chnB* monooxygenase activity of each expressed enzyme from Example 9 was tested for activity according to its ability to convert cyclohexanone to caprolactone.

##### Conversion of Cyclohexanone to Caprolactone.

- 15 Clones containing the full length monooxygenase genes were transferred from LB agar plate to 5 mL of M63 minimal media (GIBCO) containing 10 mM glycerol, 50 ug/mL ampicillin, 0.1 mM IPTG, and 500 mg/L cyclohexanone. In addition to the clones containing full length

monooxygenases, a plasmid without an insert and a "no cell" control were also assayed. The encoded monooxygenase sequences were expressed upon addition of IPTG to the culture media. The cultures were incubated overnight at room temperature (24°C). Samples (1.25 mL) for analysis  
5 were taken immediately after inoculation and after overnight incubation; cells were removed by centrifugation (4°C, 13,000 rpm).

#### GC-MS Detection of Caprolactone

Caprolactone formed by the action of the cloned monooxygenase was extracted from the aqueous phase with ethylacetate (1.0 ml  
10 aqueous/0.5 mL ethylacetate). Caprolactone was detected by gas chromatography mass spectrometry (GC-MS) analysis, using an Agilent 6890 Gas chromatograph system.

The analysis of the ethylacetate phase was performed by injecting 1 µL of the ethyl acetate phase into the GC. The inlet temperature was  
15 115°C and the column temperature profile was 50° C for 4 min and ramped to 250°C at 20°C/min, for a total run time of 14 min. The compounds were separated with an Hewlet Packard HP-5MS (5% phenyl Methyl Siloxane) column (30 m length, 250 µm diameter, and 0.25 µm film thickness). The mass spectrometer was run in Electron Ionization mode.  
20 The background mass spectra was subtracted from the spectra at the retention time of caprolactone (9.857 min). Presence of caprolactone was confirmed by comparison of the test reactions to an authentic standard obtained from Aldrich Chemical Company (St. Louis, MO).

Results of these assays are shown below in Table 7, in terms of the  
25 presence or absence of detectable caprolactone formation according to the activity of each expressed BV monooxygenase enzyme.

Table 7  
Ability of Monooxygenase Enzymes to Convert Cyclohexanone to  
Caprolactone

	Formation of Caprolactone		
	Detected	Not Detected	Not Assayed
<i>chnB</i>	ORF8	ORF 15	ORF 10
Monooxygenases	ORF9	No cell control	ORF 13
	ORF11	Plasmid control	ORF 14
	ORF12		ORF 20
	ORF16		
	ORF 17		
	ORF18		
	ORF19		

5

#### EXAMPLE 11

##### Identification of Signature Sequences Between Families of BV Monooxygenases

Sequence analysis of the 20 genes encoding Baeyer-Villiger  
monooxygenases identified in the previous examples allows definition of three  
different BV signature sequence families based on amino acid similarities. Each  
family possesses several member genes for which biochemical validation of the  
enzyme as a functional BV enzyme capable of the oxidation of cyclohexanone  
was demonstrated (Examples, *supra*). Sequence alignment of the homologues  
for each family was performed by Clustal W alignment (Higgins and Sharp (1989)  
*CABIOS*, 5:151-153). This allows the identification of a set of amino acids that  
are conserved at specific positions in the alignment created from all the  
sequences available.

The results of these Clustal W alignments are shown in Figures 7, 8, and 9  
for BV Family1, BV family 2, and BV Family 3. In all cases, an "\*" indicates a  
conserved signature amino acid position. The conserved amino acid signature  
sequence for each Family is shown in Figure 6, along with the signature  
sequence P-# positions. This conserved amino acid/ position set becomes a  
signature for each family. Any new protein with a sequence that can be aligned  
with those of the existing members of the family and which includes at the specific  
positions a at least 80% of the signature sequence amino acids can be  
considered a member of the specific family.

BV Family 1

This family comprises the *chnB* monooxygenase sequences of *Arthrobacter* sp. BP2 (SEQ ID NO:12), *Rhodococcus* sp. phi1 (SEQ ID NO:8), *Rhodococcus* sp. phi2 (SEQ ID NO:10), *Acidovorax* sp. CHX (SEQ ID NO:14), *Brevibacterium* sp. HCU (SEQ ID NOs:16 and 18), and *Rhodococcus erythropolis* AN12 ORF10, ORF14, ORF19, and ORF20 (SEQ ID NOs:26, 34, 44 and 46). Within a length of 540 amino acids, a total of 74 positions are conserved (100%). This signature sequence of Family 1 BV monooxygenases is shown beneath each alignment of proteins (Figure 7) and is listed as SEQ ID NO:47. The ability to identify the signature sequence within this family of proteins was made possible by: 1) the number of sequences of BV monooxygenases; and 2) the characterization of their activity as BV-monooxygenases.

Based on the limited number (4 total) of BV monooxygenase sequences in the public domain, for which biochemical data is also available, 3 of these sequences align with the signature sequence discovered for Family 1. These sequences are:

(1) *Acinetobacter* sp. NCIMB9871 *chnB* (NCBI Accession Number AB026668, based on Chen, Y.C. et al. (*J Bacteriol.* 170(2):781-789 (1988)). Key biochemical characterization of this protein was performed by Donogue et al. (*Eur J Biochem.* 16;63(1):175-92 (1976)), Trudgill et al. (*Methods Enzymol.* 188:70-77 (1990)), and Iwaki et al. (*Appl Environ Microbiol.* 65(11):5158-62 (1999)). This enzyme shares 72 of the 74 conserved amino acids in the signature sequence of Family 1 BV monooxygenases.

(2) *Rhodococcus erythropolis limB* (NCBI Accession Number AJ272366, based on the work of Barbirato et al. (*FEBS Lett.* 438 (3): 293-296 (1998)) and van der Werf et al. (*Biol. Chem.* 274 (37): 26296-26304 (1999)). Key biochemical characterization of this protein was performed by van der Werf, M.J. et al. (*Microbiology* 146 ( Pt 5):1129-41 (2000); *Biochem J.* 1;347 Pt 3:693-701 (2000); and *Appl Environ Microbiol.* 65(5):2092-102 (1999)). This enzyme is known as a carvone monooxygenase

(3) *Rhodococcus rhodochrous smo* (NCBI Accession Number AB010439). This enzyme was sequenced and characterized by Morii, S. et al. (*J. Biochem.* 126 (3), 624-631 (1999)). This enzyme is known as a steroid monooxygenase. It shares 74 of the 74 conserved amino acids in the signature sequence of Family 1 BV monooxygenases.

The enzymes described in the public domain having the highest sequence similarity to Group 1 have been characterized as dimethylaniline hydroxylases.

#### BV Family 2

5 This family comprises the *chnB* monooxygenase sequences of *Rhodococcus erythropolis* AN12 ORF9, ORF12, ORF15, ORF 16, and ORF18 (SEQ ID NOs:24, 30, 36, 38, and 42). Within a length of 497 amino acids, a total of 76 positions are conserved (100%). This signature sequence for Family 2 BV monooxygenases is shown beneath  
10 each alignment of proteins (Figure 8) and is listed as SEQ ID NO:48. The ability to identify the signature sequence within this family of proteins was made possible by: 1) the number of sequences of BV monooxygenases; and 2) the characterization of their activity as BV-monooxygenases.

Based on the limited number (4 total) of BV monooxygenase  
15 sequences in the public domain, for which biochemical data is also available, only 1 of these sequences align with the signature sequence discovered for Family 2. This sequence is *Pseudomonas putida* JD1 Key biochemical characterization of this protein was performed by Tanner A., et al. (*J Bacteriol.* 182(23):6565-6569 (2000)). This enzyme is known as  
20 an acetophenone monooxygenase. It shares 69 of the 76 conserved amino acids in the signature sequence of Family 2 BV monooxygenases.

#### BV Family 3

This family comprises the *chnB* monooxygenase sequences of *Rhodococcus erythropolis* AN12 ORF8, ORF 11, ORF 13, and ORF17  
25 (SEQ ID NOs:22, 28, 32, and 40). Within a length of 471 amino acids, a total of 41 positions are conserved (100%). This signature sequence for Family 3 BV monooxygenases is shown beneath each alignment of proteins (Figure 9) and is listed as SEQ ID NO:49. The ability to identify the signature sequence within this family of proteins was made possible  
30 by: 1) the number of sequences of BV monooxygenases; and 2) the characterization of their activity as BV-monooxygenases.

There are no sequences in the public domain with demonstrated BV activity that belong to this group. The dimethylaniline N-oxidase shares only 30 amino acids out of 41 conserved amino acids discovered  
35 in the signature sequence, which represents less than 80% of the conserved positions.



### CLAIMS

What is claimed is:

1. An isolated nucleic acid fragment selected from the group consisting of:
  - (a) an isolated nucleic acid fragment encoding a Baeyer-Villiger monooxygenase polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NOs:8, 10, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, and 46;
  - (b) an isolated nucleic acid molecule encoding a Baeyer-Villiger monooxygenase polypeptide that hybridizes with (a) under the following hybridization conditions: 0.1X SSC, 0.1% SDS, 65°C and washed with 2X SSC, 0.1% SDS followed by 0.1X SSC, 0.1% SDS; oran isolated nucleic acid fragment that is complementary to (a) or (b).
2. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 542 amino acids that has at least 55% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:8 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.
3. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 541 amino acids that has at least 53% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:10 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.
4. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 439 amino acids that has at least 37% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:22 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.
5. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 518 amino acids that has at least 44% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ

ID NO:24 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.

6. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 541 amino acids that has at least 64% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:26 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.

7. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 462 amino acids that has at least 65% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:28 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.

8. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 523 amino acids that has at least 45% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:30 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.

9. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 493 amino acids that has at least 55% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:32 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.

10. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 539 amino acids that has at least 51% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:34 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.

11. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 649 amino acids that has at least 39% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:36 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.

12. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 494 amino acids that has at least 43% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:38 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.

13. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 499 amino acids that has at least 53% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:40 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.

14. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 493 amino acids that has at least 44% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:42 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.

15. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 541 amino acids that has at least 54% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:44 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.

16. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 545 amino acids that has at least 42% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:46 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.

17. The isolated nucleic acid fragment of Claim 1 selected from the group consisting of SEQ ID NOs:7, 9, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, and 45.

18. An isolated nucleic acid fragment of Claim 1 isolated from *Rhodococcus*.

19. A polypeptide encoded by the isolated nucleic acid fragment of Claim 1.

20. The polypeptide of Claim 19 selected from the group consisting of SEQ ID NOs:8, 10, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, and 46.

21. An isolated nucleic acid fragment selected from the group consisting of:

- (a) an isolated nucleic acid fragment encoding a Baeyer-Villiger monooxygenase polypeptide having an amino acid sequence as set forth in SEQ ID NO:12;
- (b) an isolated nucleic acid molecule encoding a Baeyer-Villiger monooxygenase polypeptide that hybridizes with (a) under the following hybridization conditions: 0.1X SSC, 0.1% SDS, 65°C and washed with 2X SSC, 0.1% SDS followed by 0.1X SSC, 0.1% SDS; or

an isolated nucleic acid fragment that is complementary to (a), or (b).

22. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 532 amino acids that has at least 57% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:11 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.

23. An isolated nucleic acid fragment of Claim 21 isolated from *Arthrobacter*.

24. A polypeptide encoded by the isolated nucleic acid fragment of Claim 21.

25. The polypeptide of Claim 24 as set forth in SEQ ID NO:12.

26. An isolated nucleic acid fragment selected from the group consisting of:

- (a) an isolated nucleic acid fragment encoding a Baeyer-Villiger monooxygenase polypeptide having an amino acid sequence as set forth in SEQ ID NO:18;
- (b) an isolated nucleic acid molecule encoding a Baeyer-Villiger monooxygenase polypeptide that hybridizes with (a) under the following hybridization conditions: 0.1X SSC, 0.1% SDS, 65°C and washed with 2X SSC, 0.1% SDS followed by 0.1X SSC, 0.1% SDS; or

an isolated nucleic acid fragment that is complementary to (a), or (b).

27. An isolated nucleic acid molecule comprising a first nucleotide sequence encoding a polypeptide of at least 538 amino acids that has at

least 57% identity based on the Smith-Waterman method of alignment when compared to a polypeptide having the sequence as set forth in SEQ ID NO:17 or a second nucleotide sequence comprising the complement of the first nucleotide sequence.

28. An isolated nucleic acid fragment of Claim 26 isolated from *Acidovorax*.

29. A polypeptide encoded by the isolated nucleic acid fragment of Claim 26.

30. The polypeptide of Claim 29 selected from the group consisting of SEQ ID NO:18.

31. A chimeric gene comprising the isolated nucleic acid fragment of any one of Claims 1, 19, 25, 30, or 35 operably linked to suitable regulatory sequences.

32. A transformed host cell comprising a host cell and the chimeric gene of Claim 31.

33. The transformed host cell of Claim 32 wherein the host cell is selected from the group consisting of bacteria, yeast, filamentous fungi, and green plants.

34. The transformed host cell of Claim 33 wherein the host cell is selected from the group consisting of proteobacteria and actinomycetes.

35. The transformed host cell of Claim 34 wherein the host cell is selected from the group consisting of *Burkholderia*, *Alcaligenes*, *Pseudomonas*, *Sphingomonas*, *Pandoraea*, *Delftia* and *Comamonas*.

36. The transformed host cell of Claim 33 wherein the host cell is selected from the group consisting of *Rhodococcus*, *Acinetobacter*, *Mycobacteria*, *Nocardia*, *Arthrobacter*, *Brevibacterium*, *Acidovorax*, *Bacillus*, *Streptomyces*, *Escherichia*, *Salmonella*, *Pseudomonas*, *Aspergillus*, *Saccharomyces*, *Pichia*, *Candida*, *Comyebacterium*, and *Hansenula*.

37. The transformed host cell of Claim 33 wherein the host cell is selected from the group consisting of soybean, rapeseed, sunflower, cotton, corn, tobacco, alfalfa, wheat, barley, oats, sorghum, rice, *Arabidopsis*, cruciferous vegetables, melons, carrots, celery, parsley, tomatoes, potatoes, strawberries, peanuts, grapes, grass seed crops, sugar beets, sugar cane, beans, peas, rye, flax, hardwood trees, softwood trees, and forage grasses

38. A method of obtaining a nucleic acid fragment encoding a Baeyer-Villiger monooxygenase polypeptide comprising:

- (a) probing a genomic library with the nucleic acid fragment of any one of Claims 1, 21, or 26;
- (b) identifying a DNA clone that hybridizes with the nucleic acid fragment of any one of Claims 1, 21, or 26;
- (c) sequencing the genomic fragment that comprises the clone identified in step (b);

wherein the sequenced genomic fragment encodes a Baeyer-Villiger monooxygenase polypeptide.

39. A method of obtaining a nucleic acid fragment encoding a Baeyer-Villiger monooxygenase polypeptide comprising:

- (a) synthesizing at least one oligonucleotide primer corresponding to a portion of the isolated nucleic acid sequence of any one of Claims 1, 21, or 26; and
- (b) amplifying an insert present in a cloning vector using the oligonucleotide primer of step (a);

wherein the amplified insert encodes a Baeyer-Villiger monooxygenase polypeptide.

40. A method for the identification of a polypeptide having monooxygenase activity comprising:

- (a) obtaining the amino acid sequence of a polypeptide suspected of having monooxygenase activity; and
- (b) aligning the amino acid sequence of step (a) with the amino acid sequence of a Baeyer-Villiger monooxygenase consensus sequence selected from the group consisting of SEQ ID NO:47, SEQ ID NO:48 and SEQ ID NO:49;

wherein where at least 80% of the amino acid residues at positions p1-p74 of SEQ ID NO:47, or at least 80% of the amino acid residues at p1-p76 of SEQ ID NO:48 or at least 80% of the amino acid residues of p1-p41 of SEQ ID NO:49 are completely conserved, the polypeptide of (a) is identified as having monooxygenase activity.

41. A method according to Claim 40 wherein least 100% of the amino acid residues at positions p1-p74 of SEQ ID NO:47, or at least 100% of the amino acid residues at p1-p76 of SEQ ID NO:48 or at least 100% of the amino acid residues of p1-p41 of SEQ ID NO:49 are completely conserved.

42. A method for identifying a gene encoding a Baeyer-Villiger monooxygenase polypeptide comprising:

- (a) probing a genomic library with a nucleic acid fragment encoding a polypeptide wherein where at least 80% of the amino acid residues at positions p1- p74 of SEQ ID NO:47, or at least 80% of the amino acid residues at p1-p76 of SEQ ID NO:48 or at least 80% of the amino acid residues of p1-p41 of SEQ ID NO:49 are completely conserved;
- (b) identifying a DNA clone that hybridizes with a nucleic acid fragment of step (a);
- (c) sequencing the genomic fragment that comprises the clone identified in step (b);

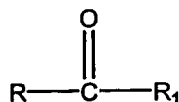
wherein the sequenced genomic fragment encodes a Baeyer-Villiger monooxygenase polypeptide.

43. A method according to Claim 42 wherein least 100% of the amino acid residues at positions p1- p74 of SEQ ID NO:47, or at least 100% of the amino acid residues at p1-p76 of SEQ ID NO:48 or at least 100% of the amino acid residues of p1-p41 of SEQ ID NO:49 are completely conserved.

44. The product of either of Claims 40 or 42.

45. A method for the biotransformation of a ketone substrate to the corresponding ester, comprising: contacting a transformed host cell under suitable growth conditions with an effective amount of ketone substrate whereby the corresponding ester is produced, said transformed host cell comprising a nucleic acid fragment encoding an isolated nucleic acid fragment of any of Claims 1, 21, 26 or 44; under the control of suitable regulatory sequences.

46. The method of Claim 45 wherein the ketone substrate is selected from the group consisting of cyclic ketones and ketoterpenes having the general formula:

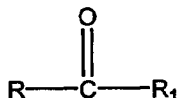


wherein R and R<sub>1</sub> are independently selected from substituted or unsubstituted phenyl, substituted or unsubstituted alkyl, or substituted or unsubstituted alkenyl or substituted or unsubstituted alkylidene.

47. The method of Claim 46 wherein the ketone substrate is selected from the group consisting of Norcamphor, Cyclobutanone, Cyclopentanone, 2-methyl-cyclopentanone, Cyclohexanone, 2-methyl-cyclohexanone, Cyclohex-2-ene-1-one, 1,2-cyclohexanedione, 1,3-cyclohexanedione, 1,4-cyclohexanedione, Cycloheptanone, Cyclooctanone, Cyclodecanone, Cycloundecanone, Cyclododecanone, Cyclotridecanone, Cyclopenta-decanone, 2-tridecanone, dihexyl ketone, 2-phenyl-cyclohexanone, Oxindole, Levoglucosenone, dimethyl sulfoxide, dimethyl-2-piperidone, Phenylboronic acid, and beta-ionone.

48. A method for the *in vitro* transformation of a ketone substrate to the corresponding ester, comprising: contacting a ketone substrate under suitable reaction conditions with an effective amount of a Baeyer-Villiger monooxygenase enzyme, the enzyme having an amino acid sequence selected from the group consisting of SEQ ID NOs:8, 10, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44, and 46.

49. A method according to Claim 49 wherein the ketone substrate is selected from the group consisting of cyclic ketones and ketoterpenes having the general formula:



wherein R and R<sub>1</sub> are independently selected from substituted or unsubstituted phenyl, substituted or unsubstituted alkyl, or substituted or unsubstituted alkenyl or substituted or unsubstituted alkylidene.

50. A method according to Claim 48 wherein the ketone substrate is selected from the group consisting of Norcamphor, Cyclobutanone, Cyclopentanone, 2-methyl-cyclopentanone, Cyclohexanone, 2-methyl-cyclohexanone, Cyclohex-2-ene-1-one, 1,2-cyclohexanedione, 1,3-cyclohexanedione, 1,4-cyclohexanedione, Cycloheptanone, Cyclooctanone, Cyclodecanone, Cycloundecanone, Cyclododecanone, Cyclotridecanone, Cyclopenta-decanone, 2-tridecanone, dihexyl ketone, 2-phenyl-cyclohexanone, Oxindole, Levoglucosenone, dimethyl sulfoxide, dimethyl-2-piperidone, Phenylboronic acid, and beta-ionone.



51. A mutated microbial gene encoding a protein having an altered biological activity produced by a method comprising the steps of
- (i) digesting a mixture of nucleotide sequences with restriction endonucleases wherein said mixture comprises:
    - a) a native microbial gene selected from the group consisting of SEQ ID NOs:7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43, and 45;
    - b) a first population of nucleotide fragments which will hybridize to said native microbial sequence;
    - c) a second population of nucleotide fragments which will not hybridize to said native microbial sequence;wherein a mixture of restriction fragments are produced;
  - (ii) denaturing said mixture of restriction fragments;
  - (iii) incubating the denatured said mixture of restriction fragments of step (ii) with a polymerase;
  - (iv) repeating steps (ii) and (iii) wherein a mutated microbial gene is produced encoding a protein having an altered biological activity.
52. An *Acidovorax* sp. comprising the 16s rDNA sequence as set forth in SEQ ID NO:5
53. An *Arthrobacter* sp. comprising the 16s rDNA sequence as set forth in SEQ ID NO:1
54. A *Rhodococcus* sp. comprising the 16s rDNA sequence as set forth in SEQ ID NO:6
55. An isolated nucleic acid useful for the identification of a BV monooxygenase selected from the group consisting of SEQ ID 70-113.

Figure 1

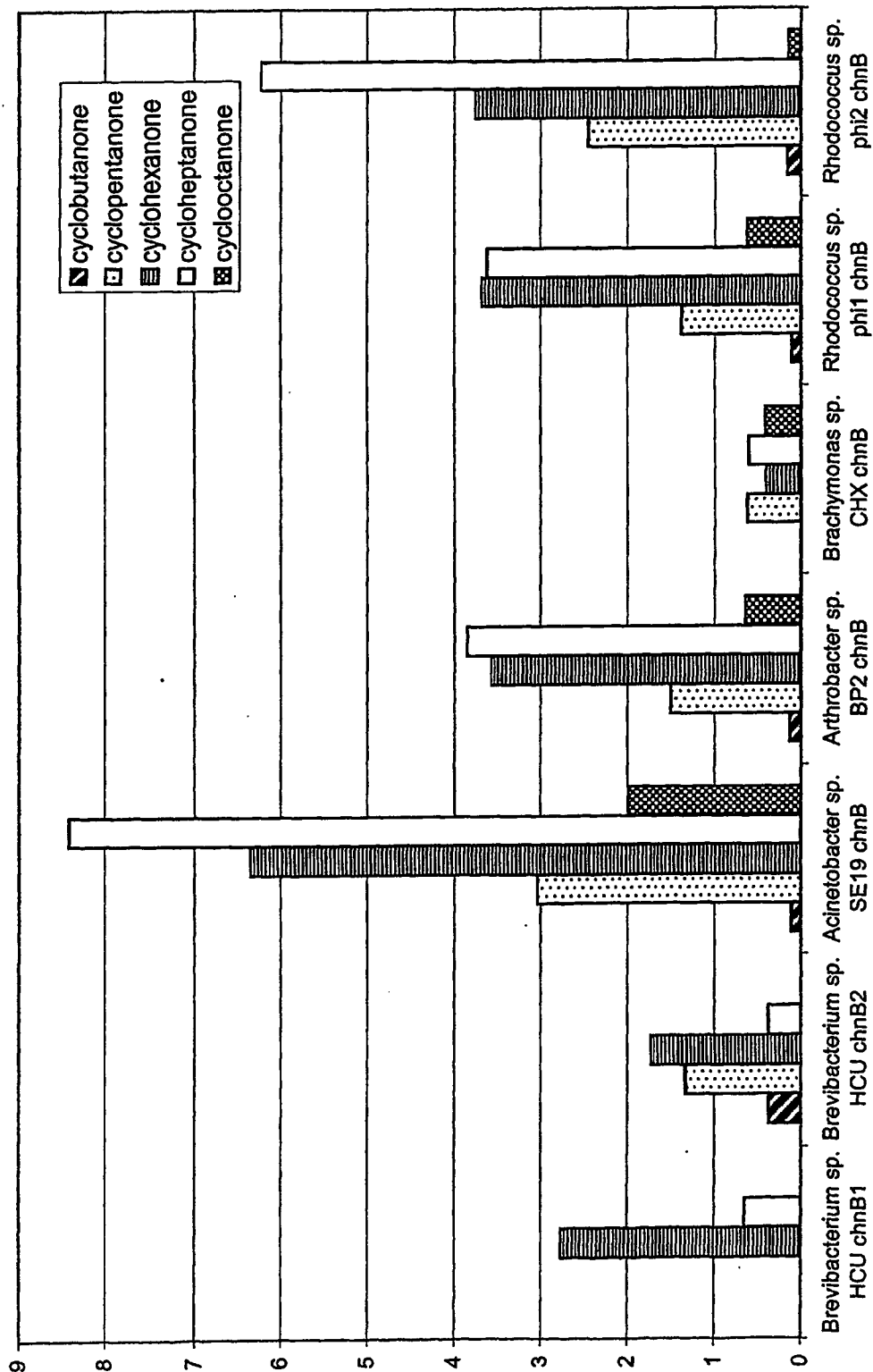


Figure 2

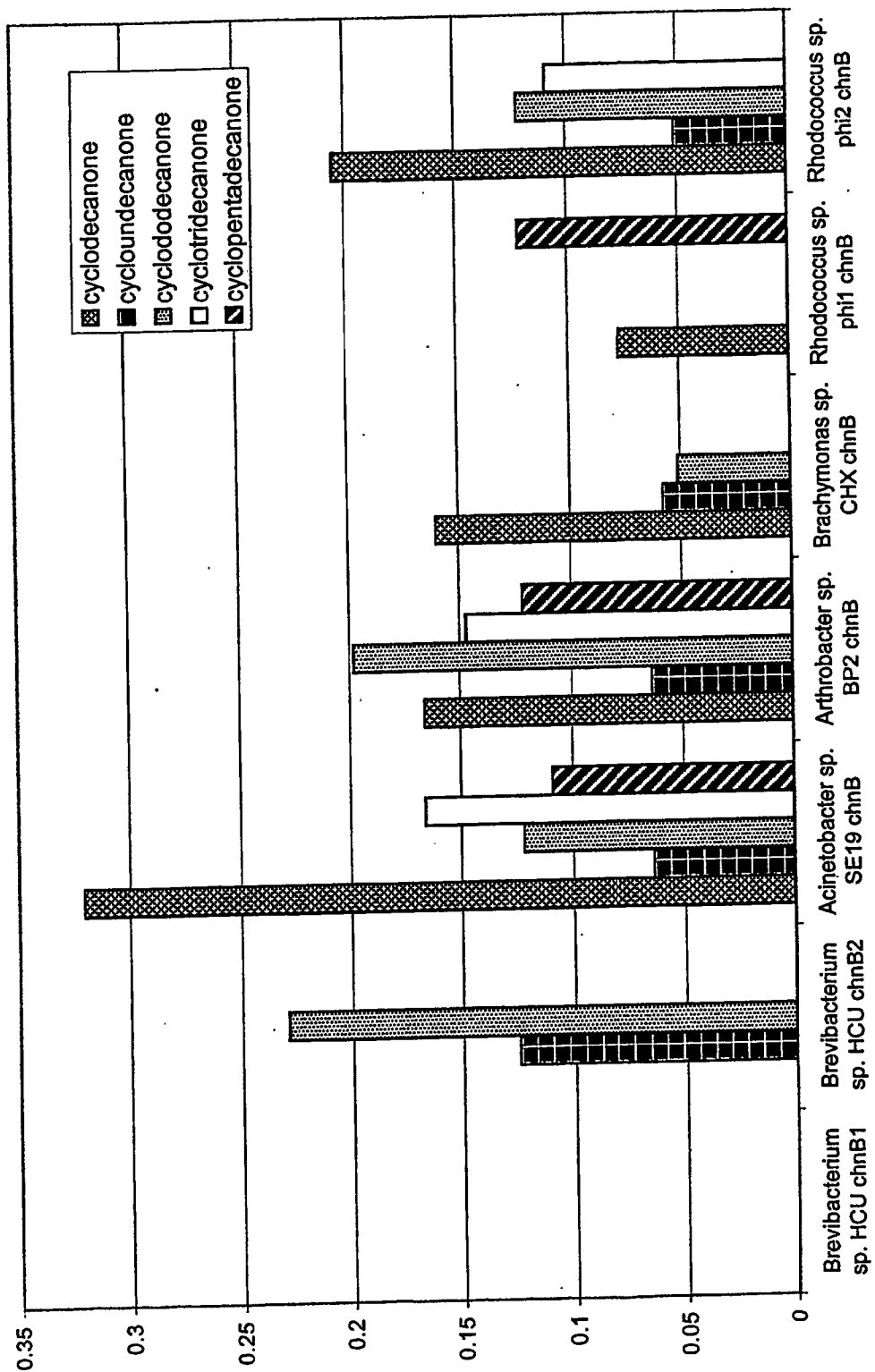


Figure 3

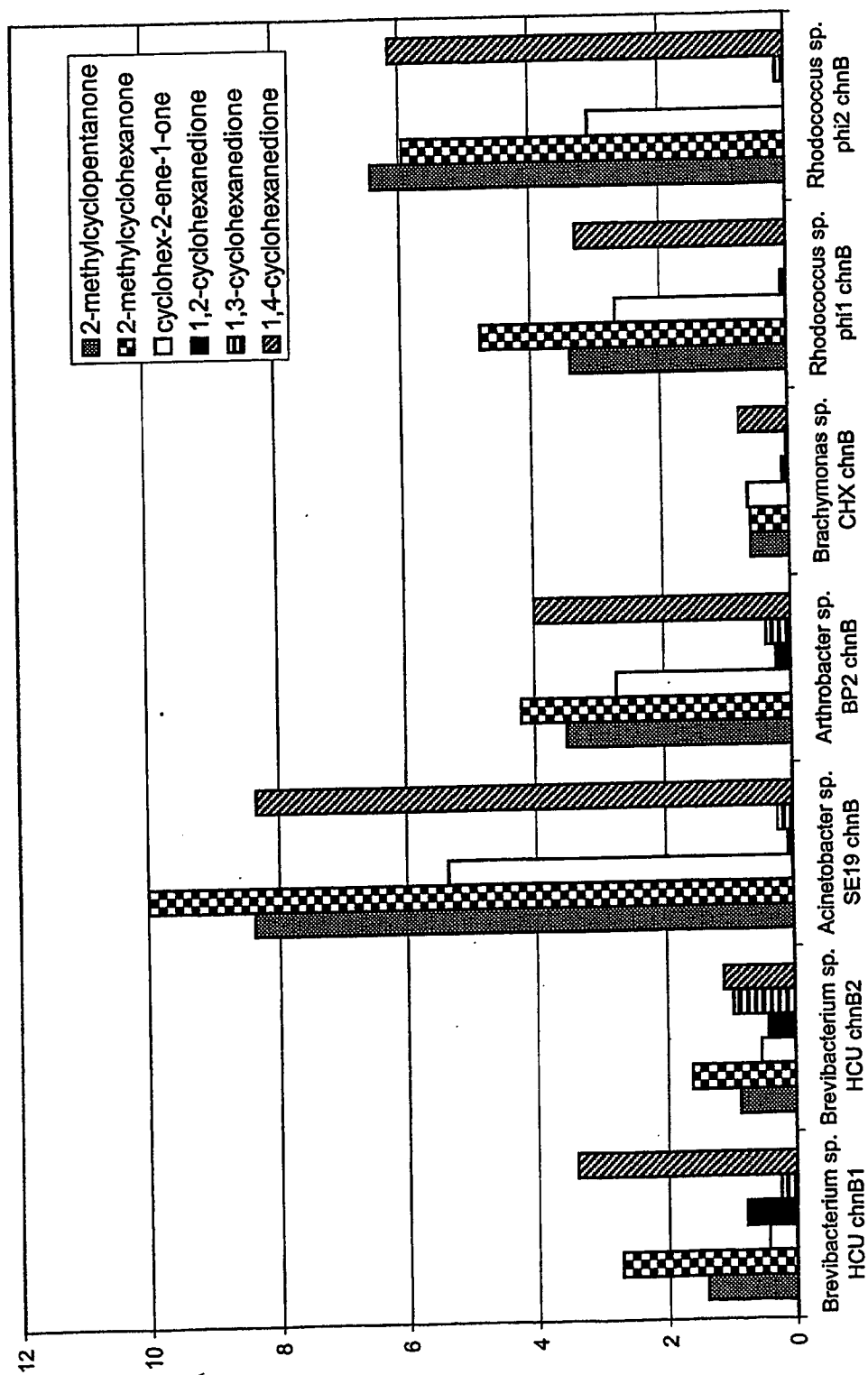


Figure 4

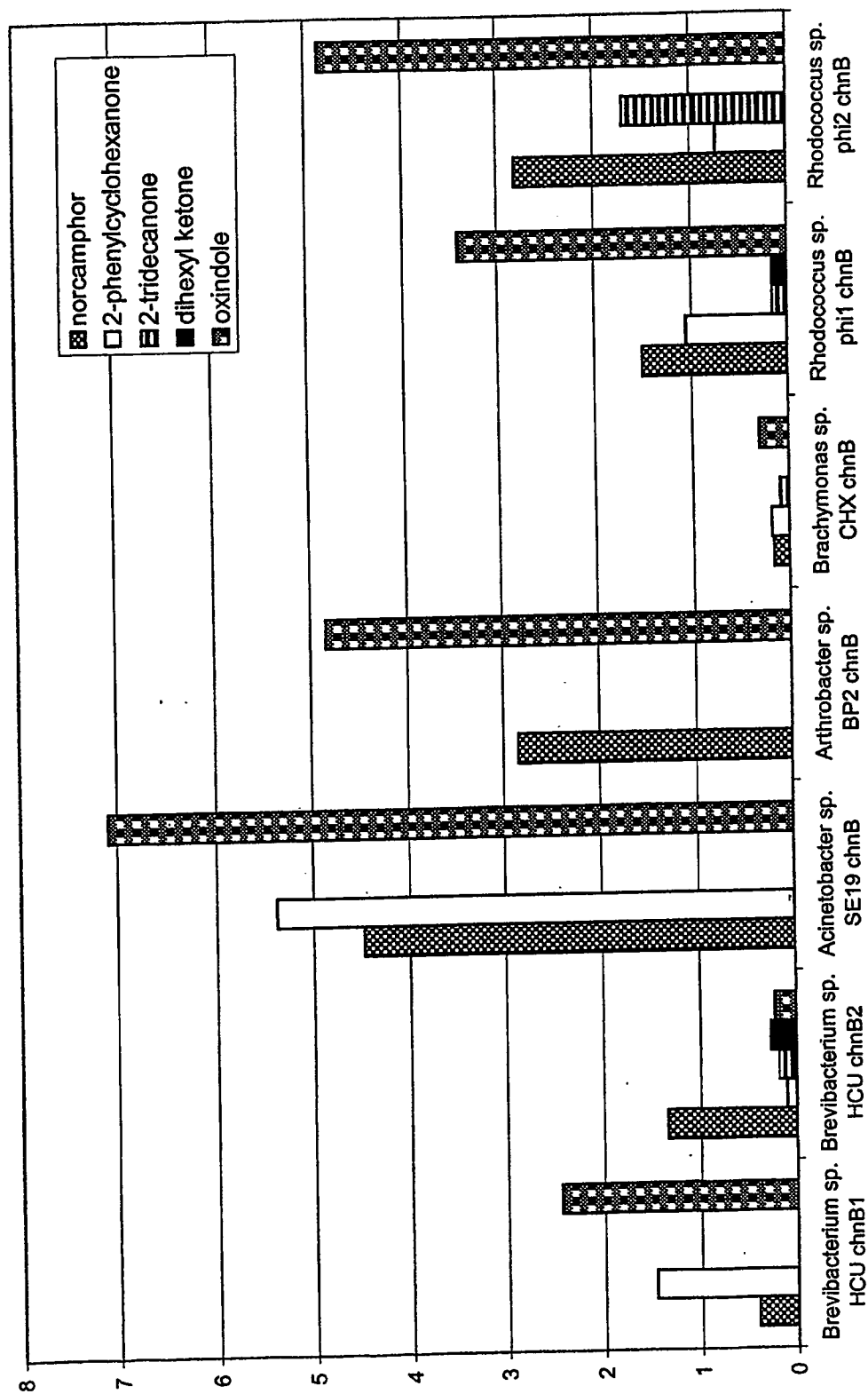


Figure 5

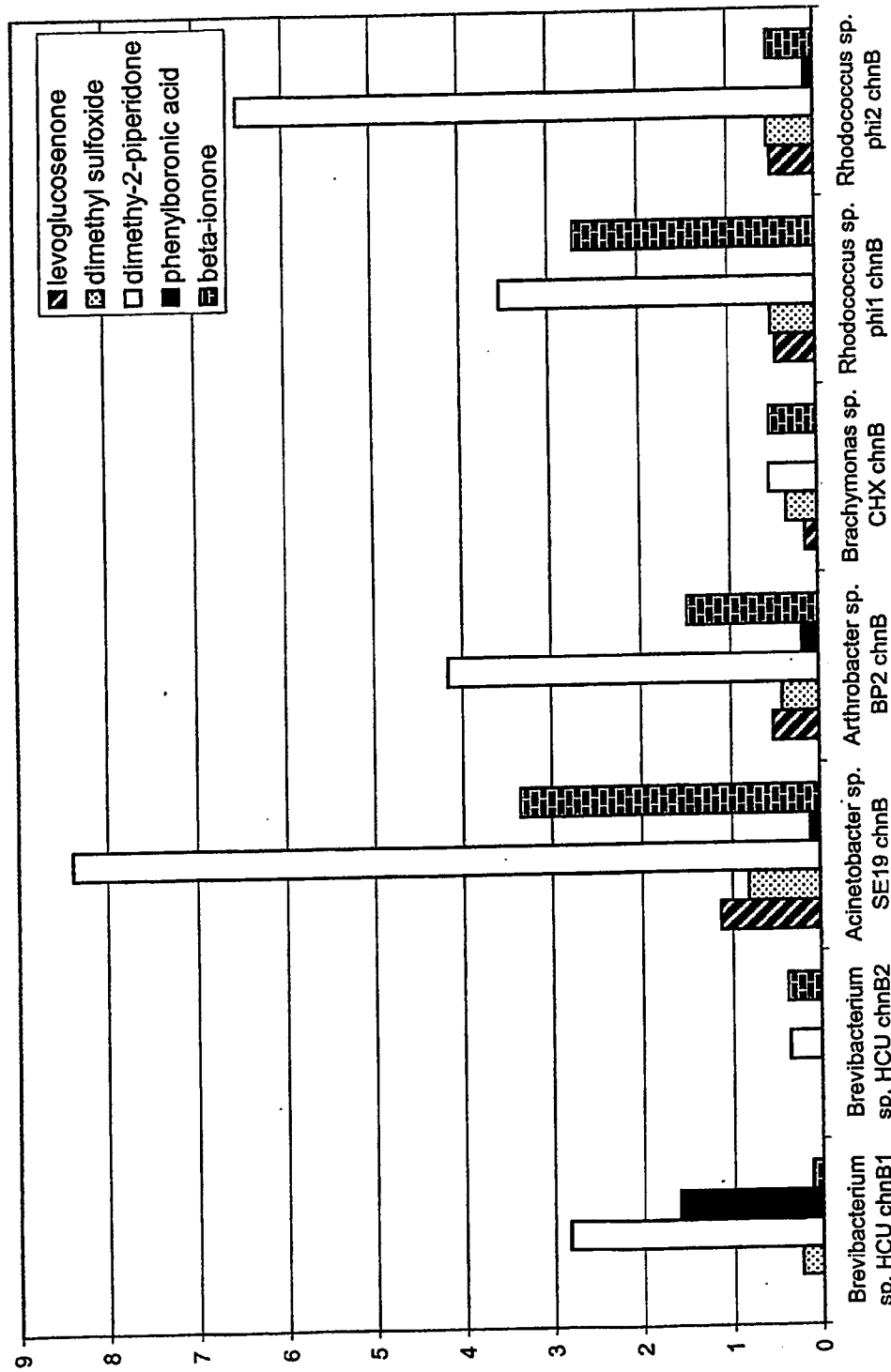


FIGURE 6

**BVMO Family 1 consensus:**

MTAQESLTVVDAVVIGAGFGGIYAVHKLREQGLTVVGFDAAADGPGGTWYWNRYPGALSDTESHVYRFSFDEDLLQDWTWKE  
 TYPTQPEILEYLEDVDRFDLRRDFRFGTEVTSATYLEDENLWEVTTDGGEVYRARFVVNAVGLLSAINFPNIPGLDTFEG  
 ETIHTAAWPEGVDLTGKRVGVI GTGSTGIQVITALAPEVEHLTVFVRTPQYSVPVGNRPVTAEQIDA KADYDEIWAQVKF  
 SGVAFGFREESTVPAMSVSEBERNRVFEEAWEEGGGFRFMFGTFGDIATDEAANETAASFIRSKIREIVKDPETARKLTPTG  
 LFARRRLCDDGYEYVYNRPNVEAVDIKENPIREITAKGVVTEGVLHELDVLVFATGFDVAVDGNYRRIDIRGRGGLSLNDE  
 WDGQPTSYLGLSTAGFPNWFVVLGPNGPFTNLPPSIETQVEWISDTIAYAEENGIRAIPTPEAEDEWTATCTDIANATLF  
 TKADSWIFGANVPGKKPSVLFYLGGLGNRAVLADVAAAGYRGFALKSADAVTA (SEQ ID NO:47)

**Signature Sequence Positions**  
**BVMO Family 1**

<i>Amino acid</i>	<i>Consensus position</i>	<i>Signature Position</i>	<i>Amino acid</i>	<i>Consensus position</i>	<i>Signature Position</i>	<i>Amino acid</i>	<i>Consensus position</i>	<i>Signature Position</i>
D	11	P-1	G	178	P-26	P	354	P-51
G	16	P-2	V	181	P-27	I	355	P-52
G	18	P-3	V	183	P-28	D	374	P-53
G	21	P-4	G	185	P-29	A	379	P-54
G	32	P-5	G	187	P-30	T	380	P-55
G	45	P-6	G	190	P-31	G	381	P-56
G	46	P-7	Q	192	P-32	D	383	P-57
W	48	P-8	I	194	P-33	G	387	P-58
N	51	P-9	A	198	P-34	G	399	P-59
Y	53	P-10	L	204	P-35	W	406	P-60
P	54	P-11	V	206	P-36	G	415	P-61
G	55	P-12	F	207	P-37	P	422	P-62
D	59	P-13	R	209	P-38	N	423	P-63
Y	65	P-14	R	265	P-39	P	430	P-64
D	101	P-15	G	276	P-40	P	433	P-65
L	102	P-16	F	286	P-41	N	436	P-66
W	124	P-17	F	302	P-42	E	464	P-67
G	144	P-18	K	306	P-43	W	473	P-68
G	156	P-19	D	313	P-44	W	492	P-69
F	160	P-20	L	320	P-45	G	495	P-70
G	162	P-21	P	322	P-46	N	497	P-71
H	166	P-22	R	329	P-47	P	499	P-72
T	167	P-23	Y	336	P-48	G	500	P-73
W	170	P-24	N	344	P-49	K	501	P-74
P	171	P-25	V	345	P-50			

**BVMO Family 2 consensus:**

MVXIPXRHXEVVIIIGAGFAGIGAAVELKRXGIDDFVLLERADDVGGTWRDNTYPGAACDVPSXLYSYSFAP  
 NPNWTRLFAXQPEIYDYLEDVAAAXGLXXHVRFGVEVTEARWDESAQLWRVXTASGELTAXFLVAATGPLS  
 XPKIPDLPGLESFEGXXFHSAXWNHDLDLRGERVAVVGTGASAVQFVPEIADXAXTLTVFQRTQWVLP  
 DXTLPPXAXRAVFSRVPGTQKWLKRLYGIFEALGSGFVXPXWLLPXXXALARAHLRRQVRDPELRXKLT  
 YTPGCKRMLLSNDWYPALXKPNVSLVTSGVVEVTEXGVVDADGVEHEVDTIIFATGFHXTDXPXAMKIFGR  
 EGRSLADHWNGSAXAYLGTAVSGFPNLFXLLGPNTGLGHTSIVXILEAQAEYIASALXXMRREGLGALDVR  
 AEVQXXFNXAVQERLATTVWNAGGCSSWYXDPDGRNSTXWPWSTXXFRARTRRFDPDSDYXPSSTPETXXG  
 (SEQ ID NO:48)

**Signature Sequence Positions****BVMO Family 2**

<i>Amino acid</i>	<i>Consensus position</i>	<i>Signature Position</i>	<i>Amino acid</i>	<i>Consensus position</i>	<i>Signature Position</i>	<i>Amino acid</i>	<i>Consensus position</i>	<i>Signature Position</i>
G	15	P-1	F	155	P-27	R	291	P-53
G	17	P-2	G	157	P-28	L	302	P-54
G	20	P-3	F	160	P-29	V	307	P-55
E	39	P-4	H	161	P-30	G	321	P-56
G	45	P-5	W	165	P-31	D	333	P-57
G	46	P-6	G	173	P-32	T	339	P-58
W	48	P-7	G	180	P-33	G	340	P-59
N	51	P-8	G	182	P-34	F	341	P-60
Y	53	P-9	A	183	P-35	G	357	P-61
P	54	P-10	S	184	P-36	W	364	P-62
G	55	P-11	A	185	P-37	G	373	P-63
D	59	P-12	Q	187	P-38	F	379	P-64
P	61	P-13	P	190	P-39	P	380	P-65
L	64	P-14	Q	203	P-40	N	381	P-66
Y	65	P-15	R	204	P-41	G	387	P-67
S	66	P-16	W	208	P-42	P	388	P-68
S	68	P-17	P	211	P-43	S	396	P-69
W	75	P-18	D	214	P-44	E	402	P-70
E	84	P-19	P	229	P-45	Q	404	P-71
Y	88	P-20	R	236	P-46	Y	407	P-72
W	120	P-21	L	268	P-47	V	429	P-73
G	139	P-22	Q	271	P-48	V	445	P-74
P	144	P-23	D	274	P-49	G	460	P-75
P	147	P-24	L	277	P-50	R	461	P-76
P	150	P-25	P	283	P-51	P	467	P-77
G	151	P-26	K	290	P-52			



**BVMO Family 3 consensus:**

MSTEHLDVLIIGAGLSGIGAAXRLXREXGIXFAILEARDNVGGTWDLFNYPGIRSDSDHLTXGKGAFRPFPPXAKYLADGPS  
 HELXXYVRDTAXEXGLRXHIXFGTKVVAAXXXAXSLWTVTVXXXGETEVXTYNVLXXANGYYSDKGNIPDFPGEFXGXLV  
 HPQXYPEXLDYRGKKVVVIGSGASGXTLAPXMXXXAXHVTMLQRSGTYIALPSDAVVPXQLAGXRXXXXXLQXXQLRXPPW  
 XAKRLXLLLIIRQLGKNVXLXGFPTPSYXPWDQHLCVVPNGDLLKXLGSGDAXIXTDIDTFTGKGVXFASGREXDADVVT  
 ATGLNXXXGGPFTXXDGLLVLDLXXRXALFYXXXXXSDNLNFLGXVGYTNASWTLRADLAXLVACRLXXMXXRSAXXXXXH  
 AXAEXXXLLASGYKXRXGXMPXQGXKXXWXXXNXXDRXLXXXXXXXXXXXXXFSKXXXXXXX (SEQ  
 ID NO 49)

**Signature Sequence Positions**  
**BVMO Family 1**

<i>Amino acid</i>	<i>Consensus position</i>	<i>Signature Position</i>	<i>Amino acid</i>	<i>Consensus position</i>	<i>Signature Position</i>
G	12	P-1	G	159	P-22
A	13	P-2	H	163	P-23
G	14	P-3	K	176	P-24
G	17	P-4	V	178	P-25
A	21	P-5	V	180	P-26
E	36	P-6	G	182	P-27
G	42	P-7	G	184	P-28
G	43	P-8	A	198	P-29
W	45	P-9	R	206	P-30
S	57	P-10	P	220	P-31
F	67	P-11	P	242	P-32
D	78	P-12	P	269	P-33
Y	87	P-13	G	293	P-34
V	107	P-14	G	314	P-35
W	118	P-15	D	320	P-36
V	120	P-16	A	325	P-37
T	121	P-17	T	326	P-38
G	141	P-18	G	327	P-39
P	151	P-19	D	361	P-40
G	155	P-20	L	415	P-41
F	157	P-21	Y	419	P-42

```

-----MTDE-----FDVVIVGAGLAGMOMLHEVR-MVGLTAKV
---MTDPDFSTAP----LDVVVIGAGVAGMYAMHRLR-EQGLRVHG
---MTAQNTFQT----VDAVVIGAGFGGIYAVHKLHNEQGGLTVVG
---MTTQKALTT----VDAIVIGAGFGGIYAVHKLANELGLTTVG
---MTAQTIHT----VDAVVIGAGFGGIYAVHKLHHELGLTTVG
---MTAQISPTV----VDAVVIGAGFGGIYAVHKLHNEQGGLTVVG
---MSSSPSSAIH----FDAIVVGAGFGGMYMLHKLRLDQLGLKVKV
-----MPITQLD-----HDAIVIGAGFSGLAILHHLR-EIGLDTQI
---MTTTESRTQTDKAGAVTDLALIIGAGVAGLYQLHMLR-EQGLNVRA
MTSTMPAPTAAQAN-ADETEVLDALIVGGGSGPVSVDRLR-EDGFKVKV

```

```

FEAGGGAGGTTWYNRYPGARCDVESLEYSYQFSEVLQOEWEWTRRYADQA
FEAGSGVGGTWYFNRYPGARCDVESFDYSYFSFEELQDDWDWSEKYAAQP
FDKADGPGGTWYWNRYPGALSDTESHVYRFSFDKGLLQDGTWKHTYITQP
FDKADGPGGTWYWNRYPGALSDTESHVYRFSFDRDLLQDGTWKHTYITQP
FDKADGPGGTWYWNRYPGALSDTESHLRYRFSFDRDLLQDGTWKHTYITQP
FDKADGPGGTWYWNRYPGALSDTESHLRYRFSFDRDLLQDGTWKHTYITQP
FDTAGGIGGTWYWNRYPGALSDTHSHVYQYSFDEAMLQEWTKNKNYLTQP
VEATDGIIGGTWINRYPGVVRTDSEFHYYFSFSKEVRDEWWTQRYPDGE
YDAAEDVGGTWYWNRYPGARFDS EAYIYQYLFSEDLYKNWSWSQRPPAQ
WDAAGGFGGIWNNWCYPGARTDSGTQIYQFY - KDLWKDFDFKELYPDFN

```

EIMRYISHVVETFDLARDIRFHTRVEAMTYEETRTARWTVQTD SAGEVVAK  
EILSYLDHVADRFDLRGTFTDTRVL SAQFDEGTATWRVQTDGGHDVTSR  
EILEYLEDVDRFDLRHRHFRFGTEVKSATYLEDEGLWEVTTGGGAVYRAK  
EILEYLEDVVSFRFDLRHRHFHFGTAVESAVYLEDEQLWEVTTDGTGEYRAT  
EILEYLEDVDRFDLRHRHFRFGTEVTSAYIYLEDENLWEVTTDGGVYRAT  
EILEYLESVDRFDLRHRHFRFGTEVTSAYIYLEDENLWEVSTDKGEVYRAK  
EILAYLEYVADRDLDRPDQIQLNTTVMTHFNEVHNIVEVTRDRGGYYTAR  
EVCAYLNFADRLDLRKDIQLNSRVNTARWNETEKYWDVTFEDGSSKRAR  
EIERWMRYVADTDLDRSIQFSTTITSAEFDEVAERWTIRTRDGRGEISTR  
GVREYFEYVDSQLDRSDVTFTNTFAESCTWDDAAKEWTVRSSEGREQRAR  
\* \* \* \* \*

FVIMATGCLSENPVPIPGVETPAGDVLHTGRWPQDPVDFTGKRVGVIGT  
FVVCATGSLSTANVPNIAGRETFGGDVHTGFWPHEGVDFTGKRVGVIGT  
YVINAVGLLSAINFPNLPGIDTFEGETIHTAAWP-QGKSLAGRVRGVIGT  
YVVNAVGLLSAINRPNLPLGDTFEGETIHTAAWP-EGKDLTGRVRGVIGT  
YVVNAVGLLSAINFPNLPGDTPFEGETIHTAAWP-EGKSLAGRVRGVIGT  
YVVNAVGLLSAINFPDLPLGDTFEGETIHTAAWP-EGKNLAGKRVGVIGT  
FIVTALGLLSAINWPNIPGRESFQGEYHTAAWP-KDVELRGKRVGVIGT  
FLISAMGALSQAIFPAIDGIDFNGAKYHTAAWPADGVDFTGKRVGVIGV  
FFITCCGMLSAPMEDLFPQQDFRQIPIHTRSRWPHGDVELTGKRVGVVGV  
AVIVATGFGAKPLYPNIEGLDSFEGECHHTARWPQGGLDMTGKRVVVMGT  
\* \* \* \* \*

GSSGVQAIPLIARQAAELVVFFQRTPAYTLPAVDEPLDPELQAAIKADYRG  
GSSGIQSIPLIAEQADHLYVFFQRSANYSVPGAGNTPLDDKRRAEIKAGYAE  
GSTGQQVITALAPEVEHLTVFVRTPQYSVPVVGKRPVTTQQIDEIKADYDN  
GSTGQQVITALAPTVEHLTVFVRTPQYSVPVVGKRAVTTDEQIDAVKADYEN  
GSTGQQVITALAPEVEHLTVFVRTPQYSVPVGNRPVTPQIDAIKADYDR  
GSTGQQVITALAPEVEHLTVFVRTPQYSVPVGNRPVTKQIDAIAKADYDG  
GSTGVQLITAIAPEVKHLTVFQRTPQYSVPTGNRPVSAQIEAEVKRNFESK  
GASGIQIIPELAKLAGELFVFQRTPNYVVESNNDKVDAEWMQYVVRDNYDE  
GATGIQVIQTIADEVQDLKFVRTPQYALPMKNPQYDSSDDVAAYKDRFEE  
GASGIQVIEAAVAEHLTVFQRTPNLALPMRQRLSADDNDRYENIED  
\* \* \* \* \*

FRARNNEVPTAGLSRFPPTNPNSVFLFSTKERDAILEHNWNRGG--PLMLR  
RRALSK--RSGGGSPFVSDPRSALEVSEAERNAAEERWKLGG--VLFAK  
IWAQVK--RSGVAFGFEESTVPAMSVTEEERROVYEKAWEYGGGRFMFE

2005  
1273  
Arthrobacter  
2082  
Rhodococcus-phi2-Mono  
Rhodococcus-phi1-Mono  
Acidovorax  
Brevibacterium-Mono1  
2093  
Brevibacterium-Mono2

IWTQVK--RSSVAFGEESTLPAHSVSA[REDACTED]RVYEEAWEGGGGFRFMFG  
IWEQAK--NSAVAFGEESTLPAHSVSEEEERNRIQEAWDHGGGFRFMFG  
IWDSVK--KSAVAFGEESTLPAHSVSEEEERNRIQEAWDHGGGFRFMFG  
VWQQVR--ESAVAFGEESTLPAHSVSEAEQRVVFQEAWNQNGFYFMFG  
IFERAS--KHPFGVDMMEYPTDSAVEVSEEEKRVFESKWEEGG--FHFANE  
LRTTLP--HTFTGFEYDFEYVWADLAPE-QRREVLNIYEYGS-LKLWLS  
RFQIRD--NSFAGFDYFIPQNAADTPEDERTAIYEKMWDEGG--FPLWLG  
\* \* \* : : \*

2005  
1273  
Arthrobacter  
2082  
Rhodococcus-phi2-Mono  
Rhodococcus-phi1-Mono  
Acidovorax  
Brevibacterium-Mono1  
2093  
Brevibacterium-Mono2

AFGDLVDSAANEVVAEFVRNKIRQIVTDPEVAAKLTP-T--HVIGCKRI  
TFADQTSNIEANGTAAFAERKIRSEVQDQAIADLLIPND--HPIGTRKI  
TFSDIATDEEANETAASFIRNKIVETIKDPETARKLTP----TGLFARRP  
TFGDIATDEEANETAASFIRSKITAMIEDPETARKLTP----TGLFARRP  
TFGDIATDEAANEAASFIKSKIAEIIEDPETARKLMP----TGLFAKRP  
TFGDIATDEAANEAASFIKSKIAEIIEDPETARKLMP----TGLYAKRP  
TFCDIATDPQANEAAATFIRNKIAEIVKDPETARKLTP----TDVYARRP  
CFTDLGTSPEASELASEFIRSKIREVVKDPATADLLCPKS--YSFNGKRV  
SFAEMFFDEQVSEISEFVREKMRARLIDPELCDLLIPTD--YGFQTHRV  
NFQGLLTDEAANHTFYNFWRSKVHDRVKDPKTAEMPLAPTPPHFGVKRP  
\* . . . \* : \* . \* . \* : \*

2005  
1273  
Arthrobacter  
2082  
Rhodococcus-phi2-Mono  
Rhodococcus-phi1-Mono  
Acidovorax  
Brevibacterium-Mono1  
2093  
Brevibacterium-Mono2

CLSDGYETYNRVNVLVDIKRHPIEEITPTTARTGE-DSHDLMLVFAT  
VTDNTYYQSYNRDNVSLVDLKSAPIEAIDEAGIKTAD-AHYELDALVFAT  
LCDDGYFQVFNRPNVEAVAIAKENPIREVTAAGVVTEDGVLHLDVIVFAT  
LCDDGYFQVFNRPNVEAVAIAKENPIREVTAAGVVTEDGVLHLDVIVFAT  
LCDAGYHQVFNRPNVEAVAIAKENPIREVTAAGVVTEDGVLHLDVIVFAT  
LCDNGYEVYFNRPNVEAVAIAKENPIREVTAAGVVTEDGVLHLDVIVFAT  
LCDSGYYRTYFNRSNVSLVDVKATPISAMTPRGIRTADGVEHLDMLILAT  
PTGHGYETFPNRTNVHLLDARGTPITRISSEKGIHGD-TEYELDAIVFAT  
PLETNYLEVYHRPNVTAIGVKNPFIARIVPQGIELTDGTFHELDVILAT  
SLEQNYFDVYNQDNVLDISNATPITRVLPGVETPD-GVVECDVIVLAT  
\* : : \* : . \* : : : \* : : \*

2005  
1273  
Arthrobacter  
2082  
Rhodococcus-phi2-Mono  
Rhodococcus-phi1-Mono  
Acidovorax  
Brevibacterium-Mono1  
2093  
Brevibacterium-Mono2

GYDAITGALSRIIDIRGRAGLSLQEAWS-DGPRTYLGLGVSGFPNLFIMTG  
GFDAMTGALDRIEIRGRNGETLRENWH-AGPRTYLGLGVHGFNLFIVTG  
GFDVADGNYRMEISGRDGVNINDHWD-GQPTSYLGVSTAKFPNWFMLVG  
GFDVADGNYRMTISGRGGLNINDHWD-GQPTSYLGVSTANFPNWFMLVG  
GFDVADGNYRRIEIRGRDGLHINDHWD-GQPTSYLGVSTANFPNWFMLVG  
GFDVADGNYRRIEIRGRNGLHINDHWD-GQPTSYLGVSTANFPNWFMLVG  
GYDAVADGNYRRIIDIRGRGGQTINEHWN-DTPTSYGVSTANFPNMFILG  
GFDAMTGTLNIDIVGRDGVILRDKWAQDGLRTNIGLTVNGFPNFMLSLG  
GFDAGTGALTIRIDIRGRGGRSLKEDWG-RDIRTMGLMVHGYPNMLTTAV  
GFDNNSGGINAIDIKA-GGQLLRDKWA-TGVDTYMGLSTHGFNLMFLYG  
\* : \* : : \* : : \* : : \* : : \*

2005  
1273  
Arthrobacter  
2082  
Rhodococcus-phi2-Mono  
Rhodococcus-phi1-Mono  
Acidovorax  
Brevibacterium-Mono1  
2093  
Brevibacterium-Mono2

PGSPSV-LTNVLVAIHQHATWIGECLEKHMNDNDIRTMEATPEAEQNWGDH  
PGSPSV-LSNMILAAEQHVDWIAGAINHLDASAGIDTIEPSAEAVDNWLDL  
PNGP---FTNLPPSIETQVEWISDTVAYAEENGIRAIPTPEAEAEWTET  
PNGP---FTNLPPSIETQVEWISDTIGYVERTGVRAIPTPEAEASAWTAT  
PNGP---FTNLPPSIETQVEWISDTIGYAEENGVRRAIPTPEAEAEWTET  
PNGP---FTNLPPSIETQVEWISDTVAYAEERNEIRAIPTPEAEAEWTET  
PNGP---FTNLPPSIEAQVEWITDLVAHMRQHGLATAEPTRAEDAWGRT  
PQTP---YSLNVVPIQLGAQWQRFKFIQERGIEVFESSREAEIWNAE  
PLAPSAALCNMTTCLQQQTEWISEAIRYMQRDLTVIPTKEAEDAWVAH  
PQSPSG-FCNGTDFGGAPGDMVADFLIWLKDNIGISRFESTEVEEREWRAH  
\* \* \* : : \* : : \* : : \*

2005  
1273  
Arthrobacter  
2082  
Rhodococcus-phi2-Mono  
Rhodococcus-phi1-Mono  
Acidovorax  
Brevibacterium-Mono1  
2093  
Brevibacterium-Mono2

VRDLAEQTLSS----CGSWYLGANIPGKRQVFMPLVG-FPDYAKKCAEI  
CSRRASATLFPS----ANSWYMGANIPGKPRIFMPPFISGGFVYSDICADV  
CTQIANMTVFTK----VDSWIFGANVPGKKPSVLFYLGGLGNRYRGLVLDV  
CTDIANMTVFTK----VDSWIFGANVPGKKPSVLFYLGGLGNRYRAVLADV  
CTAIANATLFTK----GDSWIFGANIPGKTFSVLFYLGGLGNRYRAVLAEV  
CTDIANATLFTK----GDSWIFGANVPGKKPSVLFYLGGLGNRYRNLVAGV  
CAEIAEQTLFGQ----VESWIFGANSPGKKHTLMFYLAGLGNRYRKLQADV  
TIRGAESTVMSIEGPKAGANFIGGNI PGKSREYQVYMGGGQVYQDWCREA  
HDETAAVNLISK----TDSWYVGSNVPGKPRRVLSYTGGVGAYREKAQEI  
VDDIFVNSLFPK----AKSWYGANVPGKPAQMLNYSSEASPHI-----  
\* : : \* : \* : \*

2005	ASAGYPGFAPQYDP <sup>2</sup> --VPVNQS----	[SEQ ID NO:34]
1273	AAAGYRGFELN-----SAVHA-----	[SEQ ID NO:26]
Arthrobacter	TANGYRGFELKS-E--AAVAA-----	[SEQ ID NO:12]
2082	TEGGYQGFKLKT-A--DTVDA-----	[SEQ ID NO:44]
Rhodococcus-phi2-Mono	ATDGYRGFDVKS-A--EMVTV-----	[SEQ ID NO:10]
Rhodococcus-phi1-Mono	VADSYRGFELKS-A--VPVTAZ----	[SEQ ID NO:8]
Acidovorax	ANAQYQGFAFQP-L-----	[SEQ ID NO:18]
Brevibacterium-Mono1	EESDYATFLNADSIDGEKVRRESAGMK	[SEQ ID NO:14]
2093	ADAGYKGFNLR-----	[SEQ ID NO:46]
Brevibacterium-Mono2	-----	[SEQ ID NO:16]

12/14

2022	-----	[SEQ ID NO:38]
1985	--TARETVRSR-----	[SEQ ID NO:24]
1294	-----	[SEQ ID NO:42]
2035	--TARG-----	[SEQ ID NO:36]

1861 --MSTEHLVDLIVGAGLSGIGAAYRLQTELPKGSYAILEARANSGGTWDLFKY--PGIRSD  
 1976 --MTQHVDVLIIGAGLSGIGAACHLIREQTGSTYAILERRENIGGTWDLFKY--PGIRSD  
 1413 --MSTEGKYALIGAGPSGLAGARNLDR--AGIAFDGFEHDDVGGGLWDIDNP--HSTVYE  
 2034 MSPSPLPSVCIIGAGPTGITAKRMKE--FGIPFDCYEASDEVGGNWWYKPNPGMSACYQ  
 : . : : \* \* : : \* . : \* : \* \* : . : :  
 1861 SDMFTLGYP---FRPWTDAKAIADGDS---ILRYVRDTARENGIDKKIRYNRKVTAASWS  
 1976 SDMLTFGFG---FRPWIGTKVLADGAS---IRDYVEETAKEYGVTDHINFGRKVAMDFD  
 1413 SAHLISSKGTAFAEFPMADSVADYPSHIELAEYFRDYADTHDLRRHPAFG--TTVIDVL  
 2034 SLHIDTSKWRLAFEDFPVSADLPDFPHHSELFQYFKDYVEHFGLRESIIFN-TSVVAAER  
 \* : . \* : : : \* : \* . : . : : : :  
 1861 SATSTWTVTVTTGD--EDETLCNPLYLCSGYYSYDGGYTPDFPGRESFAGEVVPQFWP  
 1976 RTAAQWSVTVLVEATGETETWTANVLVGACGYNYDKGYRPAFPGEDDFRGQIVHPQHWP  
 1413 PVDSLWQVTTSRRS-GETSVARYRGVILANGTLSKPN--IPTFRG--DFTGTLMHTSEYR  
 2034 DANGLTWVTRSDGE-----VRTYDVLVCMNGHHWDFN--IPDYPG--EFDGVLMSHSYV  
 . . \* \* . : . \* \* : \* . \* \* : : :  
 1861 ---EELDYSDDKVVVIGSGATAVTLVPTMSRDASHVTMLQRSPTYILALPSSDKLSDTIR  
 1976 ---EDLDYTGGKVVVIGSGATAITLIPSMAPTAGHVTMLQRSPTWIQALPSEDPVAKGLK  
 1413 ---SAEIFRGKRVLVIGAGNSG---CDIAYDAVH-----QAECDLSVRRGY  
 2034 DFPDPIIDMRGKKVVVVGNGNSG---LDIASSELGQR-----YLADKLIVSARRGVW  
 . \* : : \* \* : : : : \* : :  
 1861 -AVLPNQLAHSIARWKSVVVNLSFYQLCRRSPARAKRMLNLAI SRQLPKDIPLDP---HF  
 1976 LARVPDQIAYKIGRARNIALQRASFQLSRTNPKLAKKLFLAQIRLQLGKNVDLR---HF  
 1413 --FVPKYL---GR-PSDTLN-----QGKPLPPWIKQRVDTLKKQFTGDPVRFG---FP  
 2034 --VLPKYL---GV-PGDKLI-----T--PPWMPRGLRLFLSRRFLGKNLGTMEGYGLP  
 : \* . : . : : \* : : : :  
 1861 TPSYDPWDQRLCVVPGDLFKALRSGKASIEDHIDTFTETGILLASGRELEADIIVTAT  
 1976 TPSYNPDQRLCVVPGDLFKVLKSGKADIVTDRIATFTEKGIVTESGRELEADIVTAT  
 1413 APDYKIYES-HPVV-NSLILHHIGHGDVHVRAD-VDRFEGKTVRFVDGSSADYDLVLCAT  
 2034 KPDHRPFEA-HPSA-SGEFLGRAGSGDITFKPA--ITKLDGKQVHFADGTAEDVDVVVCAT  
 \* . : : . . : : \* . : : : : \* : \* : : \*  
 1861 GLKMEACGMSIEVDGELVTLGDRYAYKGMMSDVPNFAMCVGYTNASWTLRADLTSMYV  
 1976 GLNVQIILGGATMSIDGEPVKLNETVAYKSVLYSDIPNFMILGYTNASWTLKADLAASYL  
 1413 GYHLDYFFIAREDLWNSGAAPDLFLNVASRRH-DNLFLVLMVEASGLGWQGR-YQQAELV  
 2034 GYNISFPFFDDENLLPDKDNRFPLFKRMMKPGIDNLFFMGLAQPMPTLVNFA-EQQSKLV  
 \* : : . : . \* . : : : : : :  
 1861 CRLLTEMDKRDYSKCVPHAT-EEMDQRPILD--LASGYVMRAVEQFPKQGSKSPWNMRQN  
 1976 CRVLKIMRDRSYTTFEVHAEPEDFAEESLMGGALTSGYIQRGDGEMPRQGARGAWKVVNN  
 1413 AKLITARTEAPAAAREFSAA---AAGPPPD--LSGGYK-----YLLKLG-----RMA  
 2034 AAYLTGKYQLPSANEMQEIT---KADEAYF--LAPYYK--S-PRHTIQLEFDPYVRNMN  
 . : . : : \* : \*  
 1861 YILDR-LHSTFGSINDHMTFSKAPARHSTFVPSKS- [SEQ ID NO:32]  
 1976 YYRDRKLMHDAEIEDGVLQFSKVDIAVVPDSKVASA [SEQ ID NO:40]  
 1413 YYVVK---D----- [SEQ ID NO:22]  
 2034 KEIAKGTKRAAASGNKLPVAARAAHELEKADRA- [SEQ ID NO:28]

## SEQUENCE LISTING

&lt;110&gt; E. I. DU PONT DE NEMOURS &amp; COMPANY

&lt;120&gt; GENES ENCODING BAEYER-VILLIGER MONOOXYGENASES

&lt;130&gt; CL1789 PCT

&lt;150&gt; 60/315,546

&lt;151&gt; 2001-08-29

&lt;160&gt; 113

&lt;170&gt; Microsoft Office 97

&lt;210&gt; 1

&lt;211&gt; 791

&lt;212&gt; DNA

&lt;213&gt; Arthrobacter sp. BP2

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&lt;210&gt; 2



&lt;211&gt; 1303

&lt;212&gt; DNA

<213> *Rhodococcus* sp. phil

&lt;400&gt; 2

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ctaataccgg atatgacctc gggatgcatg tcctgggggtg gaaagttttt cgggtgcagga      180
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&lt;211&gt; 1296

&lt;212&gt; DNA

<213> Rhodococcus sp. phi2

<400> 3

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cgaggtggag cgaatccctt aaagccggtc tcagttcgga tcgggggtctg caactcgacc     1260
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<210> 4

<211> 1388

<212> DNA

<213> Brevibacterium sp. HCU

<400> 4

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&lt;210&gt; 5

&lt;211&gt; 895

&lt;212&gt; DNA

&lt;213&gt; Brachymonas sp. CHX

&lt;400&gt; 5

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cgggtttcct ccgcatactc acgcatttca ctgctacacg cgggaattcca tccccctcg 780
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<210> 6

<211> 1439

<212> DNA

<213> Rhodococcus erythropolis AN12

<220>

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<222> (1417)..(1417)

<223> N = G or A or T or C

<400> 6

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ttatcgggtg aggatgggcc cgcggcctat cagcttggtg gtggggtaat ggcctaccaa 240
ggcgacaacg ggtacccgac ctgaaagggt gaccggccac actgggactg aaacacggcc 300

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<210> 7

<211> 1626

<212> DNA

<213> *Rhodococcus* sp. phil

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<210> 8

<211> 542

<212> PRT

<213> Rhodococcus sp. phil

<400> 8

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 35 40 45

Trp Tyr Trp Asn Arg Tyr Pro Gly Ala Leu Ser Asp Thr Glu Ser His  
 50 55 60

Leu Tyr Arg Phe Ser Phe Asp Arg Asp Leu Leu Gln Asp Gly Thr Trp  
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Lys Thr Thr Tyr Ile Thr Gln Pro Glu Ile Leu Glu Tyr Leu Glu Ser  
 85 90 95

Val Val Asp Arg Phe Asp Leu Arg Arg His Phe Arg Phe Gly Thr Glu  
 100 105 110

Val Thr Ser Ala Ile Tyr Leu Glu Asp Glu Asn Leu Trp Glu Val Ser  
 115 120 125

Thr Asp Lys Gly Glu Val Tyr Arg Ala Lys Tyr Val Val Asn Ala Val  
 130 135 140

Gly Leu Leu Ser Ala Ile Asn Phe Pro Asp Leu Pro Gly Leu Asp Thr  
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Phe Glu Gly Glu Thr Ile His Thr Ala Ala Trp Pro Glu Gly Lys Asn  
 165 170 175

Leu Ala Gly Lys Arg Val Gly Val Ile Gly Thr Gly Ser Thr Gly Gln  
 180 185 190

Gln Val Ile Thr Ala Leu Ala Pro Glu Val Glu His Leu Thr Val Phe  
 195 200 205

Val Arg Thr Pro Gln Tyr Ser Val Pro Val Gly Asn Arg Pro Val Thr  
 210 215 220

Lys Glu Gln Ile Asp Ala Ile Lys Ala Asp Tyr Asp Gly Ile Trp Asp  
 225 230 235 240

Ser Val Lys Lys Ser Ala Val Ala Phe Gly Phe Glu Glu Ser Thr Leu  
 245 250 255

Pro Ala Met Ser Val Ser Glu Glu Glu Arg Asn Arg Ile Phe Gln Glu  
 260 265 270

Ala Trp Asp His Gly Gly Gly Phe Arg Phe Met Phe Gly Thr Phe Gly  
 275 280 285

Asp Ile Ala Thr Asp Glu Ala Ala Asn Glu Ala Ala Ala Ser Phe Ile  
 290 295 300

Arg Ser Lys Ile Ala Glu Ile Ile Glu Asp Pro Glu Thr Ala Arg Lys  
 305 310 315 320

Leu Met Pro Thr Gly Leu Tyr Ala Lys Arg Pro Leu Cys Asp Asn Gly  
 325 330 335

Tyr Tyr Glu Val Tyr Asn Arg Pro Asn Val Glu Ala Val Ala Ile Lys  
 340 345 350

Glu Asn Pro Ile Arg Glu Val Thr Ala Lys Gly Val Val Thr Glu Asp  
 355 360 365

Gly Val Leu His Glu Leu Asp Val Leu Val Phe Ala Thr Gly Phe Asp  
 370 375 380

Ala Val Asp Gly Asn Tyr Arg Arg Ile Glu Ile Arg Gly Arg Asn Gly  
 385 390 395 400

Leu His Ile Asn Asp His Trp Asp Gly Gln Pro Thr Ser Tyr Leu Gly  
 405 410 415

Val Thr Thr Ala Asn Phe Pro Asn Trp Phe Met Val Leu Gly Pro Asn  
 420 425 430

Gly Pro Phe Thr Asn Leu Pro Pro Ser Ile Glu Thr Gln Val Glu Trp  
 435 440 445

Ile Ser Asp Thr Val Ala Tyr Ala Glu Arg Asn Glu Ile Arg Ala Ile  
 450 455 460

Glu Pro Thr Pro Glu Ala Glu Glu Glu Trp Thr Gln Thr Cys Thr Asp  
 465 470 475 480

Ile Ala Asn Ala Thr Leu Phe Thr Arg Gly Asp Ser Trp Ile Phe Gly  
 485 490 495

Ala Asn Val Pro Gly Lys Lys Pro Ser Val Leu Phe Tyr Leu Gly Gly



500

505

510

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&lt;210&gt; 9

&lt;211&gt; 1623

&lt;212&gt; DNA

&lt;213&gt; Rhodococcus sp. phi2

&lt;400&gt; 9

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 gacctgcgcc gccacttccg gttcggcacc gaggtcacct cggcgatcta tctcgacgac 360  
 gagaacctct gggaggtcac caccgacggc ggcgacgtct atcgggcgac ctacgtcgtc 420  
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 gaggcgagaga ccatccacac cgccgcctgg ccggagggca agagcctcgc cgggcgccgc 540  
 gtcggcgta tcggtaccgg ttccaccggc cagcaggtca tcacggcgct ggcgccggag 600  
 gtcgagcacc tcaccgtctt cgtccggacc ccgcagtact ccgtaccggt cggcaaccgt 660  
 cccgtgaccc cggagcagat cgacgcgac aaggccgact acgaccgaat ctgggagcag 720  
 gccagaact ccgcggtggc cttcggcttc gaggagtcca ccctgccggc catgtccgtc 780  
 tcggaggagg agcgcaaccg gatcttccag gaggcctggg accacggcgg cggattccgt 840  
 ttcatgttcg gcaccttcgg tgacatcgcc accgacgagg ccgccaacga agccgcgcg 900  
 tcgttcatcc gctccaagat cgccgagatc atcgaggatc cggagaccgc ccgcaagctg 960  
 atgccgaccg gtctgttcgc caagcgcccg ctgtgcgacg ccggctacca ccaggtcttc 1020  
 aaccggccga acgtggaagc ggttgccatc aaggagaacc ccatccgcga ggtcaccgcg 1080  
 aaggcgctgg tgaccgagga cggcgtcctg cacgagttgg acgtgctcgt ctccgccacc 1140  
 ggcttcgacg ccgtggacgg caactaccgg cgcacgcaga tccgcggccg ggacggcctg 1200

cacatcaacg accactggga cggccagccg accagctacc tgggcgtctc cacggcgaac 1260  
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 aagacgccga gcgtactgtt ctacctgggc ggctgcgca actaccgtgc cgtcctcgcc 1560  
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 tga 1623

<210> 10

<211> 541

<212> PRT

<213> Rhodococcus sp. phi2

<400> 10

Met Thr Ala Gln Thr Ile His Thr Val Asp Ala Val Val Ile Gly Ala  
1 5 10 15

Gly Phe Gly Gly Ile Tyr Ala Val His Lys Leu His His Glu Leu Gly  
20 25 30

Leu Thr Thr Val Gly Phe Asp Lys Ala Asp Gly Pro Gly Gly Thr Trp  
35 40 45

Tyr Trp Asn Arg Tyr Pro Gly Ala Leu Ser Asp Thr Glu Ser His Leu  
50 55 60

Tyr Arg Phe Ser Phe Asp Arg Asp Leu Leu Gln Asp Gly Thr Trp Lys  
65 70 75 80

Asn Thr Tyr Val Thr Gln Pro Glu Ile Leu Glu Tyr Leu Glu Asp Val  
85 90 95

Val Asp Arg Phe Asp Leu Arg Arg His Phe Arg Phe Gly Thr Glu Val  
100 105 110

Thr Ser Ala Ile Tyr Leu Asp Asp Glu Asn Leu Trp Glu Val Thr Thr  
115 120 125

Asp Gly Gly Asp Val Tyr Arg Ala Thr Tyr Val Val Asn Ala Val Gly  
 130 135 140

Leu Leu Ser Ala Ile Asn Phe Pro Asn Leu Pro Gly Leu Asp Thr Phe  
 145 150 155 160

Glu Gly Glu Thr Ile His Thr Ala Ala Trp Pro Glu Gly Lys Ser Leu  
 165 170 175

Ala Gly Arg Arg Val Gly Val Ile Gly Thr Gly Ser Thr Gly Gln Gln  
 180 185 190

Val Ile Thr Ala Leu Ala Pro Glu Val Glu His Leu Thr Val Phe Val  
 195 200 205

Arg Thr Pro Gln Tyr Ser Val Pro Val Gly Asn Arg Pro Val Thr Pro  
 210 215 220

Glu Gln Ile Asp Ala Ile Lys Ala Asp Tyr Asp Arg Ile Trp Glu Gln  
 225 230 235 240

Ala Lys Asn Ser Ala Val Ala Phe Gly Phe Glu Glu Ser Thr Leu Pro  
 245 250 255

Ala Met Ser Val Ser Glu Glu Glu Arg Asn Arg Ile Phe Gln Glu Ala  
 260 265 270

Trp Asp His Gly Gly Gly Phe Arg Phe Met Phe Gly Thr Phe Gly Asp  
 275 280 285

Ile Ala Thr Asp Glu Ala Ala Asn Glu Ala Ala Ala Ser Phe Ile Arg  
 290 295 300

Ser Lys Ile Ala Glu Ile Ile Glu Asp Pro Glu Thr Ala Arg Lys Leu  
 305 310 315 320

Met Pro Thr Gly Leu Phe Ala Lys Arg Pro Leu Cys Asp Ala Gly Tyr  
 325 330 335

His Gln Val Phe Asn Arg Pro Asn Val Glu Ala Val Ala Ile Lys Glu  
 340 345 350

Asn Pro Ile Arg Glu Val Thr Ala Lys Gly Val Val Thr Glu Asp Gly  
 355 360 365

Val Leu His Glu Leu Asp Val Leu Val Phe Ala Thr Gly Phe Asp Ala  
 370 375 380

Val Asp Gly Asn Tyr Arg Arg Ile Glu Ile Arg Gly Arg Asp Gly Leu  
 385 390 395 400

His Ile Asn Asp His Trp Asp Gly Gln Pro Thr Ser Tyr Leu Gly Val  
 405 410 415

Ser Thr Ala Asn Phe Pro Asn Trp Phe Met Val Leu Gly Pro Asn Gly  
 420 425 430

Pro Phe Thr Asn Leu Pro Pro Ser Ile Glu Thr Gln Val Glu Trp Ile  
 435 440 445

Ser Asp Thr Ile Gly Tyr Ala Glu Arg Asn Gly Val Arg Ala Ile Glu  
 450 455 460

Pro Thr Pro Glu Ala Glu Ala Glu Trp Thr Glu Thr Cys Thr Ala Ile  
 465 470 475 480

Ala Asn Ala Thr Leu Phe Thr Lys Gly Asp Ser Trp Ile Phe Gly Ala  
 485 490 495

Asn Ile Pro Gly Lys Thr Pro Ser Val Leu Phe Tyr Leu Gly Gly Leu  
 500 505 510

Arg Asn Tyr Arg Ala Val Leu Ala Glu Val Ala Thr Asp Gly Tyr Arg  
 515 520 525

Gly Phe Asp Val Lys Ser Ala Glu Met Val Thr Val Glx  
 530 535 540

<210> 11

<211> 1596

<212> DNA

<213> *Arthrobacter* sp. BP2

<400> 11

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 ggcattctacg ccgtccacaa gcttcacaac gagcagggtc tgaccgttgt cggcttcgac 120  
 aaggccgacg gtcccggcgg cacctggtac tggaaccgct acccgggcgc tctctctgac 180

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accgagagcc acgtctaccg cttctctttc gataagggcc tcctgcagga cggcacctgg 240
aagcacacct acatcaccca gcccagatc ctcgagtacc ttgaggacgt cgttgaccgc 300
tttgacctgc ggcgccactt ccgcttttgt accgaggtca agtccgccac ctacctgaa 360
gacgagggcc tgtgggaagt gaccaccggc ggcggcgcgg tgtaccgggc taagtacgtc 420
atcaacgccg tggggctgct gtcagccatc aacttcccga acctgcccgg gatcgacacc 480
tttgagggcg agaccatcca caccgccgc tgccgcagg gcaagtcctt cgccggtcgc 540
cgctgggtg tgatcggcac cggttccacc ggccagcagg tcatcacggc gctggcaccg 600
gaagttgaac acctgaccgt ctctgtcagg accccgcagt actccgtccc ggtgggcaag 660
cgccccgtga ccaccagca gattgacgag atcaaggccg actacgacaa catctgggca 720
caggtcaagc gttccggcgt agccttcggc ttcgaggaaa gcaccgtgcc ggccatgagc 780
gtcaccgaag aagaacgccg ccaggtctac gagaaggcct gggaatacgg cggcggcttc 840
cgcttcatgt tcgaaacctt cagcgacatc gccaccgacg aggaggccaa cgagactgcg 900
gcatccttca tccggaacaa gatcgtcgag accatcaagg atccggagac ggcacggaaa 960
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ggccaagggg gtggtgacgg aggacggcgt gctgcacgag ctggacgtca tcgtcttcgc 1140
gaccggtttc gacgccgtgg acggcaatta ccgccgatg gagatcagcg ggcgcgacgg 1200
cgtgaacatc aacgaccact gggacgggca gccaccagc tacctgggcg tttccacagc 1260
gaagttcccc aactggttca tgggtgctggg acccaacggc ccgttcacga acctgccgcc 1320
gagcatcgag acgcaggtcg aatggatcag cgacacggtg gcctacgcgg aggaaaacgg 1380
aatccggggc atcgagccga ccccgaggc cgaagccgag tggaccgaga cgtgcacaca 1440
gatcggaac atgacggtgt tcaccaaggt cgattcatgg atcttcggcg cgaacgttcc 1500
gggcaagaag ccagcgtgc tgttctatct gggcggcctg ggcaactacc gcggcgtcct 1560
ggacgatgtc accgacaacg gataccgcgg ctttga 1596

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&lt;210&gt; 12

&lt;211&gt; 532

&lt;212&gt; PRT

<213> *Arthrobacter* sp. BP2

&lt;400&gt; 12

Met Thr Ala Gln Asn Thr Phe Gln Thr Val Asp Ala Val Val Ile Gly  
 1 5 10 15  
 Ala Gly Phe Gly Gly Ile Tyr Ala Val His Lys Leu His Asn Glu Gln  
 20 25 30  
 Gly Leu Thr Val Val Gly Phe Asp Lys Ala Asp Gly Pro Gly Gly Thr  
 35 40 45  
 Trp Tyr Trp Asn Arg Tyr Pro Gly Ala Leu Ser Asp Thr Glu Ser His  
 50 55 60  
 Val Tyr Arg Phe Ser Phe Asp Lys Gly Leu Leu Gln Asp Gly Thr Trp  
 65 70 75 80  
 Lys His Thr Tyr Ile Thr Gln Pro Glu Ile Leu Glu Tyr Leu Glu Asp  
 85 90 95  
 Val Val Asp Arg Phe Asp Leu Arg Arg His Phe Arg Phe Gly Thr Glu  
 100 105 110  
 Val Lys Ser Ala Thr Tyr Leu Glu Asp Glu Gly Leu Trp Glu Val Thr  
 115 120 125  
 Thr Gly Gly Gly Ala Val Tyr Arg Ala Lys Tyr Val Ile Asn Ala Val  
 130 135 140  
 Gly Leu Leu Ser Ala Ile Asn Phe Pro Asn Leu Pro Gly Ile Asp Thr  
 145 150 155 160  
 Phe Glu Gly Glu Thr Ile His Thr Ala Ala Trp Pro Gln Gly Lys Ser  
 165 170 175  
 Leu Ala Gly Arg Arg Val Gly Val Ile Gly Thr Gly Ser Thr Gly Gln  
 180 185 190  
 Gln Val Ile Thr Ala Leu Ala Pro Glu Val Glu His Leu Thr Val Phe  
 195 200 205  
 Val Arg Thr Pro Gln Tyr Ser Val Pro Val Gly Lys Arg Pro Val Thr  
 210 215 220  
 Thr Gln Gln Ile Asp Glu Ile Lys Ala Asp Tyr Asp Asn Ile Trp Ala  
 225 230 235 240  
 Gln Val Lys Arg Ser Gly Val Ala Phe Gly Phe Glu Glu Ser Thr Val

16

Arg Glu Arg Ser Gly Gln Glu Ala Gln Arg Ala Val Leu Ser Gly Arg  
                   500                                  505                                  510

Pro Gly Gln Leu Pro Arg Arg Pro Gly Arg Cys His Arg Gln Arg Ile  
                   515                                  520                                  525

Pro Arg Leu Glx  
                   530

<210> 13

<211> 1662

<212> DNA

<213> Brevibacterium sp. HCU

<220>

<221> CDS

<222> (1)..(1662)

<223>

<400> 13  
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 Met Pro Ile Thr Gln Gln Leu Asp His Asp Ala Ile Val Ile Gly Ala  
 1                  5                                  10                                  15

ggc ttc tcc gga cta gcc att ctg cac cac ctg cgt gaa atc ggc cta 96  
 Gly Phe Ser Gly Leu Ala Ile Leu His His Leu Arg Glu Ile Gly Leu  
                   20                                  25                                  30

gac act caa atc gtc gaa gca acc gac ggc att gga gga act tgg tgg 144  
 Asp Thr Gln Ile Val Glu Ala Thr Asp Gly Ile Gly Gly Thr Trp Trp  
                   35                                  40                                  45

atc aac cgc tac ccg ggg gtg cgg acc gac agc gag ttc cac tac tac 192  
 Ile Asn Arg Tyr Pro Gly Val Arg Thr Asp Ser Glu Phe His Tyr Tyr  
                   50                                  55                                  60

tct ttc agc ttc agc aag gaa gtt cgt gac gag tgg aca tgg act caa 240  
 Ser Phe Ser Phe Ser Lys Glu Val Arg Asp Glu Trp Thr Trp Thr Gln  
 65                                  70                                  75                                  80

cgc tac cca gac ggt gaa gaa gtt tgc gcc tat ctc aat ttc att gct 288  
 Arg Tyr Pro Asp Gly Glu Glu Val Cys Ala Tyr Leu Asn Phe Ile Ala  
                   85                                  90                                  95

gat cga ctt gat ctt cgg aag gac att cag ctc aac tca cga gtg aat 336  
 Asp Arg Leu Asp Leu Arg Lys Asp Ile Gln Leu Asn Ser Arg Val Asn



100	105	110	
act gcc cgt tgg aat gag acg gaa aag tac tgg gac gtc att ttc gaa Thr Ala Arg Trp Asn Glu Thr Glu Lys Tyr Trp Asp Val Ile Phe Glu 115 120 125			384
gac ggg tcc tcg aaa cgc gct cgc ttc ctc atc agc gca atg ggt gca Asp Gly Ser Ser Lys Arg Ala Arg Phe Leu Ile Ser Ala Met Gly Ala 130 135 140			432
ctt agc cag gcg att ttc ccg gcc atc gac gga atc gac gaa ttc aac Leu Ser Gln Ala Ile Phe Pro Ala Ile Asp Gly Ile Asp Glu Phe Asn 145 150 155 160			480
ggc gcg aaa tat cac act gcg gct tgg cca gct gat ggc gta gat ttc Gly Ala Lys Tyr His Thr Ala Ala Trp Pro Ala Asp Gly Val Asp Phe 165 170 175			528
acg ggc aag aag gtt gga gtc att ggg gtt ggg gcc tcg gga att caa Thr Gly Lys Lys Val Gly Val Ile Gly Val Gly Ala Ser Gly Ile Gln 180 185 190			576
atc att ccc gag ctc gcc aag ttg gct ggc gaa cta ttc gta ttc cag Ile Ile Pro Glu Leu Ala Lys Leu Ala Gly Glu Leu Phe Val Phe Gln 195 200 205			624
cga act ccg aac tat gtg gtt gag agc aac aac gac aaa gtt gac gcc Arg Thr Pro Asn Tyr Val Val Glu Ser Asn Asn Asp Lys Val Asp Ala 210 215 220			672
gag tgg atg cag tac gtt cgc gac aac tat gac gaa att ttc gaa cgc Glu Trp Met Gln Tyr Val Arg Asp Asn Tyr Asp Glu Ile Phe Glu Arg 225 230 235 240			720
gca tcc aag cac ccg ttc ggg gtc gat atg gag tat ccg acg gat tcc Ala Ser Lys His Pro Phe Gly Val Asp Met Glu Tyr Pro Thr Asp Ser 245 250 255			768
gcc gtc gag gtt tca gaa gaa gaa cgt aag cga gtc ttt gaa agc aaa Ala Val Glu Val Ser Glu Glu Glu Arg Lys Arg Val Phe Glu Ser Lys 260 265 270			816
tgg gag gag gga ggc ttc cat ttt gca aac gag tgt ttc acg gac ctg Trp Glu Glu Gly Gly Phe His Phe Ala Asn Glu Cys Phe Thr Asp Leu 275 280 285			864
ggt acc agt cct gag gcc agc gag ctg gcg tca gag ttc ata cgt tcg Gly Thr Ser Pro Glu Ala Ser Glu Leu Ala Ser Glu Phe Ile Arg Ser 290 295 300			912
aag att cgg gag gtc gtt aag gac ccc gct acg gca gat ctc ctt tgt Lys Ile Arg Glu Val Lys Asp Pro Ala Thr Ala Asp Leu Leu Cys 305 310 315 320			960
ccc aag tcg tac tcg ttc aac ggt aag cga gtg ccg acc ggc cac ggc Pro Lys Ser Tyr Ser Phe Asn Gly Lys Arg Val Pro Thr Gly His Gly 325 330 335			1008
tac tac gag acg ttc aat cgc acg aat gtg cac ctt ttg gat gcc agg Tyr Tyr Glu Thr Phe Asn Arg Thr Asn Val His Leu Leu Asp Ala Arg 340 345 350			1056

ggc act cca att act cgg atc agc agc aaa ggt atc gtt cac gga gac 1104  
 Gly Thr Pro Ile Thr Arg Ile Ser Ser Lys Gly Ile Val His Gly Asp  
 355 360 365

acc gaa tac gaa cta gat gca atc gtg ttc gca acc ggc ttc gac gcg 1152  
 Thr Glu Tyr Glu Leu Asp Ala Ile Val Phe Ala Thr Gly Phe Asp Ala  
 370 375 380

atg aca ggt acg ctc acc aac att gac atc gtc ggc cgc gac gga gtc 1200  
 Met Thr Gly Thr Leu Thr Asn Ile Asp Ile Val Gly Arg Asp Gly Val  
 385 390 395 400

atc ctc cgc gac aag tgg gcc cag gat ggg ctt agg aca aac att ggt 1248  
 Ile Leu Arg Asp Lys Trp Ala Gln Asp Gly Leu Arg Thr Asn Ile Gly  
 405 410 415

ctt act gta aac ggc ttc ccg aac ttc ctg atg tct ctt gga cct cag 1296  
 Leu Thr Val Asn Gly Phe Pro Asn Phe Leu Met Ser Leu Gly Pro Gln  
 420 425 430

acc ccg tac tcc aac ctt gtt gtt cct att cag ttg gga gcc caa tgg 1344  
 Thr Pro Tyr Ser Asn Leu Val Val Pro Ile Gln Leu Gly Ala Gln Trp  
 435 440 445

atg cag cga ttc ctt aag ttc att cag gaa cgc ggc att gaa gtg ttc 1392  
 Met Gln Arg Phe Leu Lys Phe Ile Gln Glu Arg Gly Ile Glu Val Phe  
 450 455 460

gag tcg tcg aga gaa gct gaa gaa atc tgg aat gcc gaa acc att cgc 1440  
 Glu Ser Ser Arg Glu Ala Glu Glu Ile Trp Asn Ala Glu Thr Ile Arg  
 465 470 475 480

ggc gct gaa tct acg gtc atg tcc atc gaa gga ccc aaa gcc ggc gca 1488  
 Gly Ala Glu Ser Thr Val Met Ser Ile Glu Gly Pro Lys Ala Gly Ala  
 485 490 495

tgg ttc atc ggc ggc aac att ccc ggt aaa tca cgt gag tac cag gtg 1536  
 Trp Phe Ile Gly Gly Asn Ile Pro Gly Lys Ser Arg Glu Tyr Gln Val  
 500 505 510

tat atg ggc ggc ggt cag gtc tac cag gac tgg tgc cgc gag gcg gaa 1584  
 Tyr Met Gly Gly Gly Gln Val Tyr Gln Asp Trp Cys Arg Glu Ala Glu  
 515 520 525

gaa tcc gac tac gcc act ttt ctg aat gct gac tcc att gac ggc gaa 1632  
 Glu Ser Asp Tyr Ala Thr Phe Leu Asn Ala Asp Ser Ile Asp Gly Glu  
 530 535 540

aag gtt cgt gaa tcg gcg ggt atg aaa tag 1662  
 Lys Val Arg Glu Ser Ala Gly Met Lys  
 545 550

&lt;210&gt; 14

&lt;211&gt; 553

&lt;212&gt; PRT

<213> Brevibacterium sp. HCU

<400> 14

Met Pro Ile Thr Gln Gln Leu Asp His Asp Ala Ile Val Ile Gly Ala  
1 5 10 15

Gly Phe Ser Gly Leu Ala Ile Leu His His Leu Arg Glu Ile Gly Leu  
20 25 30

Asp Thr Gln Ile Val Glu Ala Thr Asp Gly Ile Gly Gly Thr Trp Trp  
35 40 45

Ile Asn Arg Tyr Pro Gly Val Arg Thr Asp Ser Glu Phe His Tyr Tyr  
50 55 60

Ser Phe Ser Phe Ser Lys Glu Val Arg Asp Glu Trp Thr Trp Thr Gln  
65 70 75 80

Arg Tyr Pro Asp Gly Glu Glu Val Cys Ala Tyr Leu Asn Phe Ile Ala  
85 90 95

Asp Arg Leu Asp Leu Arg Lys Asp Ile Gln Leu Asn Ser Arg Val Asn  
100 105 110

Thr Ala Arg Trp Asn Glu Thr Glu Lys Tyr Trp Asp Val Ile Phe Glu  
115 120 125

Asp Gly Ser Ser Lys Arg Ala Arg Phe Leu Ile Ser Ala Met Gly Ala  
130 135 140

Leu Ser Gln Ala Ile Phe Pro Ala Ile Asp Gly Ile Asp Glu Phe Asn  
145 150 155 160

Gly Ala Lys Tyr His Thr Ala Ala Trp Pro Ala Asp Gly Val Asp Phe  
165 170 175

Thr Gly Lys Lys Val Gly Val Ile Gly Val Gly Ala Ser Gly Ile Gln  
180 185 190

Ile Ile Pro Glu Leu Ala Lys Leu Ala Gly Glu Leu Phe Val Phe Gln  
195 200 205

Arg Thr Pro Asn Tyr Val Val Glu Ser Asn Asn Asp Lys Val Asp Ala  
210 215 220

Glu Trp Met Gln Tyr Val Arg Asp Asn Tyr Asp Glu Ile Phe Glu Arg  
 225 230 235 240  
 Ala Ser Lys His Pro Phe Gly Val Asp Met Glu Tyr Pro Thr Asp Ser  
 245 250 255  
 Ala Val Glu Val Ser Glu Glu Glu Arg Lys Arg Val Phe Glu Ser Lys  
 260 265 270  
 Trp Glu Glu Gly Gly Phe His Phe Ala Asn Glu Cys Phe Thr Asp Leu  
 275 280 285  
 Gly Thr Ser Pro Glu Ala Ser Glu Leu Ala Ser Glu Phe Ile Arg Ser  
 290 295 300  
 Lys Ile Arg Glu Val Val Lys Asp Pro Ala Thr Ala Asp Leu Leu Cys  
 305 310 315 320  
 Pro Lys Ser Tyr Ser Phe Asn Gly Lys Arg Val Pro Thr Gly His Gly  
 325 330 335  
 Tyr Tyr Glu Thr Phe Asn Arg Thr Asn Val His Leu Leu Asp Ala Arg  
 340 345 350  
 Gly Thr Pro Ile Thr Arg Ile Ser Ser Lys Gly Ile Val His Gly Asp  
 355 360 365  
 Thr Glu Tyr Glu Leu Asp Ala Ile Val Phe Ala Thr Gly Phe Asp Ala  
 370 375 380  
 Met Thr Gly Thr Leu Thr Asn Ile Asp Ile Val Gly Arg Asp Gly Val  
 385 390 395 400  
 Ile Leu Arg Asp Lys Trp Ala Gln Asp Gly Leu Arg Thr Asn Ile Gly  
 405 410 415  
 Leu Thr Val Asn Gly Phe Pro Asn Phe Leu Met Ser Leu Gly Pro Gln  
 420 425 430  
 Thr Pro Tyr Ser Asn Leu Val Val Pro Ile Gln Leu Gly Ala Gln Trp  
 435 440 445  
 Met Gln Arg Phe Leu Lys Phe Ile Gln Glu Arg Gly Ile Glu Val Phe  
 450 455 460

Glu Ser Ser Arg Glu Ala Glu Glu Ile Trp Asn Ala Glu Thr Ile Arg  
 465 470 475 480

Gly Ala Glu Ser Thr Val Met Ser Ile Glu Gly Pro Lys Ala Gly Ala  
 485 490 495

Trp Phe Ile Gly Gly Asn Ile Pro Gly Lys Ser Arg Glu Tyr Gln Val  
 500 505 510

Tyr Met Gly Gly Gly Gln Val Tyr Gln Asp Trp Cys Arg Glu Ala Glu  
 515 520 525

Glu Ser Asp Tyr Ala Thr Phe Leu Asn Ala Asp Ser Ile Asp Gly Glu  
 530 535 540

Lys Val Arg Glu Ser Ala Gly Met Lys  
 545 550

<210> 15

<211> 1590

<212> DNA

<213> Brevibacterium sp. HCU

<220>

<221> CDS

<222> (1)..(1590)

<223>

<400> 15

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 Met Thr Ser Thr Met Pro Ala Pro Thr Ala Ala Gln Ala Asn Ala Asp  
 1 5 10 15

gag acc gag gtc ctc gac gca ctc atc gtg ggt ggc gga ttc tcg ggg 96  
 Glu Thr Glu Val Leu Asp Ala Leu Ile Val Gly Gly Gly Phe Ser Gly  
 20 25 30

cct gta tct gtc gac cgc ctg cgt gaa gac ggg ttc aag gtc aag gtc 144  
 Pro Val Ser Val Asp Arg Leu Arg Glu Asp Gly Phe Lys Val Lys Val  
 35 40 45

tgg gac gcc gcc ggc gga ttc ggc ggc atc tgg tgg tgg aac tgc tac 192  
 Trp Asp Ala Ala Gly Gly Phe Gly Gly Ile Trp Trp Trp Asn Cys Tyr  
 50 55 60

ccg ggt gct cgt acg gac agc acc gga cag atc tat cag ttc cag tac Pro Gly Ala Arg Thr Asp Ser Thr Gly Gln Ile Tyr Gln Phe Gln Tyr 65 70 75 80	240
aag gac ctg tgg aag gac ttc gac ttc aag gag ctc tac ccc gac ttc Lys Asp Leu Trp Lys Asp Phe Asp Phe Lys Glu Leu Tyr Pro Asp Phe 85 90 95	288
aac ggg gtt cgg gag tac ttc gag tac gtc gac tcg cag ctc gac ctg Asn Gly Val Arg Glu Tyr Phe Glu Tyr Val Asp Ser Gln Leu Asp Leu 100 105 110	336
tcc cgc gac gtc aca ttc aac acc ttt gcg gag tcc tgc aca tgg gac Ser Arg Asp Val Thr Phe Asn Thr Phe Ala Glu Ser Cys Thr Trp Asp 115 120 125	384
gac gct gcc aag gag tgg acg gtg cga tcg tcg gaa gga cgt gag cag Asp Ala Ala Lys Glu Trp Thr Val Arg Ser Ser Glu Gly Arg Glu Gln 130 135 140	432
cgg gcc cgt gcg gtc atc gtc gcc acc ggc ttc ggt gcg aag ccc ctc Arg Ala Arg Ala Val Ile Val Ala Thr Gly Phe Gly Ala Lys Pro Leu 145 150 155 160	480
tac ccg aac atc gag ggc ctc gac agc ttc gaa ggc gag tgc cat cac Tyr Pro Asn Ile Glu Gly Leu Asp Ser Phe Glu Gly Glu Cys His His 165 170 175	528
acc gca cgc tgg ccg cag ggt ggc ctc gac atg acg ggc aag cga gtc Thr Ala Arg Trp Pro Gln Gly Gly Leu Asp Met Thr Gly Lys Arg Val 180 185 190	576
gtc gtc atg ggc acc ggt gct tcc ggc atc cag gtc att caa gaa gcc Val Val Met Gly Thr Gly Ala Ser Gly Ile Gln Val Ile Gln Glu Ala 195 200 205	624
gcg gcg gtt gcc gaa cac ctc acc gtc ttc cag cgc acc ccg aac ctt Ala Ala Val Ala Glu His Leu Thr Val Phe Gln Arg Thr Pro Asn Leu 210 215 220	672
gcc ctg ccg atg cgg cag cag cgg ctg tcg gcc gat gac aac gat cgc Ala Leu Pro Met Arg Gln Gln Arg Leu Ser Ala Asp Asp Asn Asp Arg 225 230 235 240	720
tac cga gag aac atc gaa gat cgt ttc caa atc cgt gac aat tcg ttt Tyr Arg Glu Asn Ile Glu Asp Arg Phe Gln Ile Arg Asp Asn Ser Phe 245 250 255	768
gcc gga ttc gac ttc tac ttc atc ccg cag aac gcc gcg gac acc ccc Ala Gly Phe Asp Phe Tyr Phe Ile Pro Gln Asn Ala Ala Asp Thr Pro 260 265 270	816
gag gac gag cgg acc gcg atc tac gaa aag atg tgg gac gaa ggc gga Glu Asp Glu Arg Thr Ala Ile Tyr Glu Lys Met Trp Asp Glu Gly Gly 275 280 285	864
ttc cca ctg tgg ctc gga aac ttc cag gga ctc ctc acc gat gag gca Phe Pro Leu Trp Leu Gly Asn Phe Gln Gly Leu Leu Thr Asp Glu Ala 290 295 300	912
gcc aac cac acc ttc tac aac ttc tgg cgt tcg aag gtg cac gat cgt	960

Ala Asn His Thr Phe Tyr Asn Phe Trp Arg Ser Lys Val His Asp Arg	
305 310 315 320	
gtg aag gat ccc aag acc gcc gag atg ctc gca ccg gcg acc cca ccg	1008
Val Lys Asp Pro Lys Thr Ala Glu Met Leu Ala Pro Ala Thr Pro Pro	
325 330 335	
cac ccg ttc ggc gtc aag cgt ccc tcg ctc gaa cag aac tac ttc gac	1056
His Pro Phe Gly Val Lys Arg Pro Ser Leu Glu Gln Asn Tyr Phe Asp	
340 345 350	
gta tac aac cag gac aat gtc gat ctc atc gac tcg aat gcc acc ccg	1104
Val Tyr Asn Gln Asp Asn Val Asp Leu Ile Asp Ser Asn Ala Thr Pro	
355 360 365	
atc acc cgg gtc ctt ccg aac ggg gtc gaa acc ccg gac gga gtc gtc	1152
Ile Thr Arg Val Leu Pro Asn Gly Val Glu Thr Pro Asp Gly Val Val	
370 375 380	
gaa tgc gat gtc ctc gtg ctg gcc acc ggc ttc gac aac aac agc ggc	1200
Glu Cys Asp Val Leu Val Leu Ala Thr Gly Phe Asp Asn Asn Ser Gly	
385 390 395 400	
ggc atc aac gcc atc gat atc aaa gcc ggc ggg cag ctg ctg cgt gac	1248
Gly Ile Asn Ala Ile Asp Ile Lys Ala Gly Gly Gln Leu Leu Arg Asp	
405 410 415	
aag tgg gcg acc ggc gtg gac acc tac atg ggg ctg tcg acg cac gga	1296
Lys Trp Ala Thr Gly Val Asp Thr Tyr Met Gly Leu Ser Thr His Gly	
420 425 430	
ttc ccc aat ctc atg ttc ctc tac ggc ccg cag agc cct tcg ggc ttc	1344
Phe Pro Asn Leu Met Phe Leu Tyr Gly Pro Gln Ser Pro Ser Gly Phe	
435 440 445	
tgc aat ggg acc gac ttc ggc gga gcg cca ggc gat atg gtc gcc gac	1392
Cys Asn Gly Thr Asp Phe Gly Gly Ala Pro Gly Asp Met Val Ala Asp	
450 455 460	
ttc ctc atc tgg ctc aag gac aac ggc atc tcg cgg ttc gaa tcc acc	1440
Phe Leu Ile Trp Leu Lys Asp Asn Gly Ile Ser Arg Phe Glu Ser Thr	
465 470 475 480	
gaa gag gtc gag cgg gaa tgg cgc gcc cat gtc gac gac atc ttc gtc	1488
Glu Glu Val Glu Arg Glu Trp Arg Ala His Val Asp Asp Ile Phe Val	
485 490 495	
aac tcg ctg ttc ccc aag gcg aag tcc tgg tac tgg ggc gcc aac gtc	1536
Asn Ser Leu Phe Pro Lys Ala Lys Ser Trp Tyr Trp Gly Ala Asn Val	
500 505 510	
ccc ggc aag ccg gcg cag atg ctc aac tat tcg gag gcg tcc ccg cat	1584
Pro Gly Lys Pro Ala Gln Met Leu Asn Tyr Ser Glu Ala Ser Pro His	
515 520 525	
atc tag	1590
Ile	

&lt;210&gt; 16

&lt;211&gt; 529

&lt;212&gt; PRT

&lt;213&gt; Brevibacterium sp. HCU

&lt;400&gt; 16

Met Thr Ser Thr Met Pro Ala Pro Thr Ala Ala Gln Ala Asn Ala Asp  
 1 5 10 15

Glu Thr Glu Val Leu Asp Ala Leu Ile Val Gly Gly Gly Phe Ser Gly  
 20 25 30

Pro Val Ser Val Asp Arg Leu Arg Glu Asp Gly Phe Lys Val Lys Val  
 35 40 45

Trp Asp Ala Ala Gly Gly Phe Gly Gly Ile Trp Trp Trp Asn Cys Tyr  
 50 55 60

Pro Gly Ala Arg Thr Asp Ser Thr Gly Gln Ile Tyr Gln Phe Gln Tyr  
 65 70 75 80

Lys Asp Leu Trp Lys Asp Phe Asp Phe Lys Glu Leu Tyr Pro Asp Phe  
 85 90 95

Asn Gly Val Arg Glu Tyr Phe Glu Tyr Val Asp Ser Gln Leu Asp Leu  
 100 105 110

Ser Arg Asp Val Thr Phe Asn Thr Phe Ala Glu Ser Cys Thr Trp Asp  
 115 120 125

Asp Ala Ala Lys Glu Trp Thr Val Arg Ser Ser Glu Gly Arg Glu Gln  
 130 135 140

Arg Ala Arg Ala Val Ile Val Ala Thr Gly Phe Gly Ala Lys Pro Leu  
 145 150 155 160

Tyr Pro Asn Ile Glu Gly Leu Asp Ser Phe Glu Gly Glu Cys His His  
 165 170 175

Thr Ala Arg Trp Pro Gln Gly Gly Leu Asp Met Thr Gly Lys Arg Val  
 180 185 190

Val Val Met Gly Thr Gly Ala Ser Gly Ile Gln Val Ile Gln Glu Ala



195	200	205
Ala Ala Val Ala Glu His Leu Thr Val Phe Gln Arg Thr Pro Asn Leu		
210	215	220
Ala Leu Pro Met Arg Gln Gln Arg Leu Ser Ala Asp Asp Asn Asp Arg		
225	230	235 240
Tyr Arg Glu Asn Ile Glu Asp Arg Phe Gln Ile Arg Asp Asn Ser Phe		
245	250	255
Ala Gly Phe Asp Phe Tyr Phe Ile Pro Gln Asn Ala Ala Asp Thr Pro		
260	265	270
Glu Asp Glu Arg Thr Ala Ile Tyr Glu Lys Met Trp Asp Glu Gly Gly		
275	280	285
Phe Pro Leu Trp Leu Gly Asn Phe Gln Gly Leu Leu Thr Asp Glu Ala		
290	295	300
Ala Asn His Thr Phe Tyr Asn Phe Trp Arg Ser Lys Val His Asp Arg		
305	310	315 320
Val Lys Asp Pro Lys Thr Ala Glu Met Leu Ala Pro Ala Thr Pro Pro		
325	330	335
His Pro Phe Gly Val Lys Arg Pro Ser Leu Glu Gln Asn Tyr Phe Asp		
340	345	350
Val Tyr Asn Gln Asp Asn Val Asp Leu Ile Asp Ser Asn Ala Thr Pro		
355	360	365
Ile Thr Arg Val Leu Pro Asn Gly Val Glu Thr Pro Asp Gly Val Val		
370	375	380
Glu Cys Asp Val Leu Val Leu Ala Thr Gly Phe Asp Asn Asn Ser Gly		
385	390	395 400
Gly Ile Asn Ala Ile Asp Ile Lys Ala Gly Gly Gln Leu Leu Arg Asp		
405	410	415
Lys Trp Ala Thr Gly Val Asp Thr Tyr Met Gly Leu Ser Thr His Gly		
420	425	430
Phe Pro Asn Leu Met Phe Leu Tyr Gly Pro Gln Ser Pro Ser Gly Phe		
435	440	445

Cys Asn Gly Thr Asp Phe Gly Gly Ala Pro Gly Asp Met Val Ala Asp  
 450 455 460

Phe Leu Ile Trp Leu Lys Asp Asn Gly Ile Ser Arg Phe Glu Ser Thr  
 465 470 475 480

Glu Glu Val Glu Arg Glu Trp Arg Ala His Val Asp Asp Ile Phe Val  
 485 490 495

Asn Ser Leu Phe Pro Lys Ala Lys Ser Trp Tyr Trp Gly Ala Asn Val  
 500 505 510

Pro Gly Lys Pro Ala Gln Met Leu Asn Tyr Ser Glu Ala Ser Pro His  
 515 520 525

Ile

<210> 17

<211> 1614

<212> DNA

<213> Brachymonas sp. CHX

<400> 17

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ggcggcatgt atatgctgca caaactgcgc gaccagctcg gactcaaggt caagggttttc	120
gacacagccg gcggcatcgg cggcacctgg tattggaatc gctatcctgg agccttgctc	180
gacacgcaca gtcattgtcta tcagtattct ttgcagcaag cgatgctcca agaattggaca	240
tggaagaaca aatacctcac gcagccagaa atactggctt atctggagta tgtagcagac	300
cggtctgatc tgcgcccgga cattcagttg aacacgaccg tgacatcgat gcatttcaat	360
gaagtccaca acatctggga agtgcgcacg gaccggggcg ggtactacac cgcgcgcttt	420
atcgtgacgg cactgggttt gttatccgcg atcaactggc ccaacattcc gggccgcgaa	480
agcttccaag gcgagatgta tcacacagcc gcctggccaa aagatgtcga actgcgcggc	540
aaacgcgtcg gcgtgatcgg caccggctcg acgggtgtgc agctgattac cgccatcgct	600
ccagaggtca aacacctgac ggtcttccag cgtacaccgc aatacagcgt gccgacggga	660
aatcgctctg tctccgcgca agaaatcgca gaagtcaagc gaaacttcag caagggtatgg	720

caacaagtac gtgaatccgc cgtcgcattc ggcttcgagg aaagcacagt gcccgcgatg 780  
 agcgtctccg aagccgaacg ccagcgcgtc ttccaggaag cctggaacca aggcaacggc 840  
 ttttactaca tggtcggcac attttgcgac atcgccaccg acccgcaggc caacgaagcc 900  
 gcagccacct tcatacgcaa caaaatcgcc gagatcgtca aagacccgga aaccgcccgc 960  
 aagctcacgc ctacggatgt ttacgcccga cgcccgtttt gcgacagtgg ctactatcgc 1020  
 acctacaacc gcagcaacgt ctactgggtg gatgtgaagg cgacaccaat cagtgcgatg 1080  
 acgccccggg gcattcgcac cgccgacggt gtcgagcacg agttggatat gttgatcctt 1140  
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 ggccaaacca tcaatgagca ctggaacgac actcctacca gttatgtagg ggtcagcacc 1260  
 gccaaacttc ccaacatggt catgatcctg ggcccgaat gccattcac gaacctgccg 1320  
 ccgtcgatcg aagcacaggt cgaatggatc accgacctgg ttgcccacat gcgccagcac 1380  
 gggctcgcga cggccgaacc aacgcgcgat gctgaagatg cctggggccg cacctgcgcg 1440  
 gaaatcgccg agcagacgct ttttggccag gttgaatcat ggatcttcgg tgccaacagc 1500  
 cccgggaaga aacatacttt gatgttctat ctggccggcc tggggaacta ccgcaagcag 1560  
 ctgcgcgacg tagcgaacgc gcaataccaa ggctttgcgt tccaaccact gtaa 1614

<210> 18

<211> 538

<212> PRT

<213> Brachymonas sp. CHX

<400> 18

Met Ser Ser Ser Pro Ser Ser Ala Ile His Phe Asp Ala Ile Val Val  
 1 5 10 15

Gly Ala Gly Phe Gly Gly Met Tyr Met Leu His Lys Leu Arg Asp Gln  
 20 25 30

Leu Gly Leu Lys Val Lys Val Phe Asp Thr Ala Gly Gly Ile Gly Gly  
 35 40 45

Thr Trp Tyr Trp Asn Arg Tyr Pro Gly Ala Leu Ser Asp Thr His Ser  
 50 55 60

His Val Tyr Gln Tyr Ser Phe Asp Glu Ala Met Leu Gln Glu Trp Thr  
 65 70 75 80

Trp Lys Asn Lys Tyr Leu Thr Gln Pro Glu Ile Leu Ala Tyr Leu Glu  
 85 90 95

Tyr Val Ala Asp Arg Leu Asp Leu Arg Pro Asp Ile Gln Leu Asn Thr  
 100 105 110

Thr Val Thr Ser Met His Phe Asn Glu Val His Asn Ile Trp Glu Val  
 115 120 125

Arg Thr Asp Arg Gly Gly Tyr Tyr Thr Ala Arg Phe Ile Val Thr Ala  
 130 135 140

Leu Gly Leu Leu Ser Ala Ile Asn Trp Pro Asn Ile Pro Gly Arg Glu  
 145 150 155 160

Ser Phe Gln Gly Glu Met Tyr His Thr Ala Ala Trp Pro Lys Asp Val  
 165 170 175

Glu Leu Arg Gly Lys Arg Val Gly Val Ile Gly Thr Gly Ser Thr Gly  
 180 185 190

Val Gln Leu Ile Thr Ala Ile Ala Pro Glu Val Lys His Leu Thr Val  
 195 200 205

Phe Gln Arg Thr Pro Gln Tyr Ser Val Pro Thr Gly Asn Arg Pro Val  
 210 215 220

Ser Ala Gln Glu Ile Ala Glu Val Lys Arg Asn Phe Ser Lys Val Trp  
 225 230 235 240

Gln Gln Val Arg Glu Ser Ala Val Ala Phe Gly Phe Glu Glu Ser Thr  
 245 250 255

Val Pro Ala Met Ser Val Ser Glu Ala Glu Arg Gln Arg Val Phe Gln  
 260 265 270

Glu Ala Trp Asn Gln Gly Asn Gly Phe Tyr Tyr Met Phe Gly Thr Phe  
 275 280 285

Cys Asp Ile Ala Thr Asp Pro Gln Ala Asn Glu Ala Ala Ala Thr Phe  
 290 295 300

Ile Arg Asn Lys Ile Ala Glu Ile Val Lys Asp Pro Glu Thr Ala Arg  
 305 310 315 320

Lys Leu Thr Pro Thr Asp Val Tyr Ala Arg Arg Pro Leu Cys Asp Ser  
325 330 335

Gly Tyr Tyr Arg Thr Tyr Asn Arg Ser Asn Val Ser Leu Val Asp Val  
340 345 350

Lys Ala Thr Pro Ile Ser Ala Met Thr Pro Arg Gly Ile Arg Thr Ala  
355 360 365

Asp Gly Val Glu His Glu Leu Asp Met Leu Ile Leu Ala Thr Gly Tyr  
370 375 380

Asp Ala Val Asp Gly Asn Tyr Arg Arg Ile Asp Leu Arg Gly Arg Gly  
385 390 395 400

Gly Gln Thr Ile Asn Glu His Trp Asn Asp Thr Pro Thr Ser Tyr Val  
405 410 415

Gly Val Ser Thr Ala Asn Phe Pro Asn Met Phe Met Ile Leu Gly Pro  
420 425 430

Asn Gly Pro Phe Thr Asn Leu Pro Pro Ser Ile Glu Ala Gln Val Glu  
435 440 445

Trp Ile Thr Asp Leu Val Ala His Met Arg Gln His Gly Leu Ala Thr  
450 455 460

Ala Glu Pro Thr Arg Asp Ala Glu Asp Ala Trp Gly Arg Thr Cys Ala  
465 470 475 480

Glu Ile Ala Glu Gln Thr Leu Phe Gly Gln Val Glu Ser Trp Ile Phe  
485 490 495

Gly Ala Asn Ser Pro Gly Lys Lys His Thr Leu Met Phe Tyr Leu Ala  
500 505 510

Gly Leu Gly Asn Tyr Arg Lys Gln Leu Ala Asp Val Ala Asn Ala Gln  
515 520 525

Tyr Gln Gly Phe Ala Phe Gln Pro Leu Glx  
530 535

<210> 19

<211> 1644

&lt;212&gt; DNA

&lt;213&gt; Acinetobacter sp. SE19

&lt;220&gt;

&lt;221&gt; CDS

&lt;222&gt; (1) .. (1644)

&lt;223&gt;

&lt;400&gt; 19

atg	gag	att	atc	atg	tca	caa	aaa	atg	gat	ttt	gat	gct	atc	gtg	att	48
Met	Glu	Ile	Ile	Met	Ser	Gln	Lys	Met	Asp	Phe	Asp	Ala	Ile	Val	Ile	
1				5					10					15		

ggc	gga	ctt	tat	gca	gtc	aaa	aaa	tta	aga	gac	gag	96				
Gly	Gly	Gly	Phe	Gly	Gly	Leu	Tyr	Ala	Val	Lys	Lys	Leu	Arg	Asp	Glu	
		20				25						30				

ctc	gaa	ctt	aag	gtt	cag	gct	ttt	gat	aaa	gcc	acg	gat	gtc	gca	ggc	144
Leu	Glu	Leu	Lys	Val	Gln	Ala	Phe	Asp	Lys	Ala	Thr	Asp	Val	Ala	Gly	
		35				40						45				

act	tgg	tac	tgg	aac	cgt	tac	cca	ggc	gca	ttg	tcg	gat	aca	gaa	acc	192
Thr	Trp	Tyr	Trp	Asn	Arg	Tyr	Pro	Gly	Ala	Leu	Ser	Asp	Thr	Glu	Thr	
	50					55					60					

cac	ctc	tac	tgc	tat	tct	tgg	gat	aaa	gaa	tta	cta	caa	tcg	cta	gaa	240
His	Leu	Tyr	Cys	Tyr	Ser	Trp	Asp	Lys	Glu	Leu	Leu	Gln	Ser	Leu	Glu	
65					70					75				80		

atc	aag	aaa	aaa	tat	gtg	caa	ggc	cct	gat	gta	cgc	aag	tat	tta	cag	288
Ile	Lys	Lys	Lys	Tyr	Val	Gln	Gly	Pro	Asp	Val	Arg	Lys	Tyr	Leu	Gln	
				85					90					95		

caa	gtg	gct	gaa	aag	cat	gat	tta	aag	aag	agc	tat	caa	ttc	aat	acc	336
Gln	Val	Ala	Glu	Lys	His	Asp	Leu	Lys	Lys	Ser	Tyr	Gln	Phe	Asn	Thr	
			100					105					110			

gcg	gtt	caa	tcg	gct	cat	tac	aac	gaa	gca	gat	gcc	ttg	tgg	gaa	gtc	384
Ala	Val	Gln	Ser	Ala	His	Tyr	Asn	Glu	Ala	Asp	Ala	Leu	Trp	Glu	Val	
		115					120					125				

acc	act	gaa	tat	ggc	gat	aag	tac	acg	gcg	cgt	ttc	ctc	atc	act	gct	432
Thr	Thr	Glu	Tyr	Gly	Asp	Lys	Tyr	Thr	Ala	Arg	Phe	Leu	Ile	Thr	Ala	
		130				135					140					

tta	ggc	tta	ttg	tct	gcg	cct	aac	ttg	cca	aac	atc	aaa	ggc	att	aat	480
Leu	Gly	Leu	Leu	Ser	Ala	Pro	Asn	Leu	Pro	Asn	Ile	Lys	Gly	Ile	Asn	
145					150					155				160		

cag	ttt	aaa	ggc	gag	ctg	cat	cat	acc	agc	cgc	tgg	cca	gat	gac	gta	528
Gln	Phe	Lys	Gly	Glu	Leu	His	His	Thr	Ser	Arg	Trp	Pro	Asp	Asp	Val	
				165					170					175		

agt ttt gaa ggt aaa cgt gtc ggc gtg att ggt acg ggt tcc acc ggt Ser Phe Glu Gly Lys Arg Val Gly Val Ile Gly Thr Gly Ser Thr Gly 180 185 190	576
gtt cag gtt att acg gct gtg gca cct ctg gct aaa cac ctc act gtc Val Gln Val Ile Thr Ala Val Ala Pro Leu Ala Lys His Leu Thr Val 195 200 205	624
ttc cag cgt tct gca caa tac agc gtt cca att ggc aat gat cca ctg Phe Gln Arg Ser Ala Gln Tyr Ser Val Pro Ile Gly Asn Asp Pro Leu 210 215 220	672
tct gaa gaa gat gtt aaa aag atc aaa gac aat tat gac aaa att tgg Ser Glu Glu Asp Val Lys Lys Ile Lys Asp Asn Tyr Asp Lys Ile Trp 225 230 235 240	720
gat ggt gta tgg aat tca gcc ctt gcc ttt ggc ctg aat gaa agc aca Asp Gly Val Trp Asn Ser Ala Leu Ala Phe Gly Leu Asn Glu Ser Thr 245 250 255	768
gtg cca gca atg agc gta tca gct gaa gaa cgc aag gca gtt ttt gaa Val Pro Ala Met Ser Val Ser Ala Glu Glu Arg Lys Ala Val Phe Glu 260 265 270	816
aag gca tgg caa aca ggt ggc ggt ttc cgt ttc atg ttt gaa act ttc Lys Ala Trp Gln Thr Gly Gly Gly Phe Arg Phe Met Phe Glu Thr Phe 275 280 285	864
ggt gat att gcc acc aat atg gaa gcc aat atc gaa gcg caa aat ttc Gly Asp Ile Ala Thr Asn Met Glu Ala Asn Ile Glu Ala Gln Asn Phe 290 295 300	912
att aag ggt aaa att gct gaa atc gtc aaa gat cca gcc att gca cag Ile Lys Gly Lys Ile Ala Glu Ile Val Lys Asp Pro Ala Ile Ala Gln 305 310 315 320	960
aag ctt atg cca cag gat ttg tat gca aaa cgt ccg ttg tgt gac agt Lys Leu Met Pro Gln Asp Leu Tyr Ala Lys Arg Pro Leu Cys Asp Ser 325 330 335	1008
ggt tac tac aac acc ttt aac cgt gac aat gtc cgt tta gaa gat gtg Gly Tyr Tyr Asn Thr Phe Asn Arg Asp Asn Val Arg Leu Glu Asp Val 340 345 350	1056
aaa gcc aat ccg att gtt gaa att acc gaa aac ggt gtg aaa ctc gaa Lys Ala Asn Pro Ile Val Glu Ile Thr Glu Asn Gly Val Lys Leu Glu 355 360 365	1104
aat ggc gat ttc gtt gaa tta gac atg ctg ata tgt gcc aca ggt ttt Asn Gly Asp Phe Val Glu Leu Asp Met Leu Ile Cys Ala Thr Gly Phe 370 375 380	1152
gat gcc gtc gat ggc aac tat gtg cgc atg gac att caa ggt aaa aac Asp Ala Val Asp Gly Asn Tyr Val Arg Met Asp Ile Gln Gly Lys Asn 385 390 395 400	1200
ggc ttg gcc atg aaa gac tac tgg aaa gaa ggt ccg tcg agc tat atg Gly Leu Ala Met Lys Asp Tyr Trp Lys Glu Gly Pro Ser Ser Tyr Met 405 410 415	1248
ggt gtc acc gta aat aac tat cca aac atg ttc atg gtg ctt gga ccg	1296

Gly Val Thr Val Asn Asn Tyr Pro Asn Met Phe Met Val Leu Gly Pro	
420 425 430	
aat ggc ccg ttt acc aac ctg ccg cca tca att gaa tca cag gtg gaa	1344
Asn Gly Pro Phe Thr Asn Leu Pro Pro Ser Ile Glu Ser Gln Val Glu	
435 440 445	
tgg atc agt gat acc att caa tac acg gtt gaa aac aat gtt gaa tcc	1392
Trp Ile Ser Asp Thr Ile Gln Tyr Thr Val Glu Asn Asn Val Glu Ser	
450 455 460	
att gaa gcg aca aaa gaa gcg gaa gaa caa tgg act caa act tgc gcc	1440
Ile Glu Ala Thr Lys Glu Ala Glu Glu Gln Trp Thr Gln Thr Cys Ala	
465 470 475 480	
aat att gcg gaa atg acc tta ttc cct aaa gcg caa tcc tgg att ttt	1488
Asn Ile Ala Glu Met Thr Leu Phe Pro Lys Ala Gln Ser Trp Ile Phe	
485 490 495	
ggt gcg aat atc ccg ggc aag aaa aac acg gtt tac ttc tat ctc ggt	1536
Gly Ala Asn Ile Pro Gly Lys Lys Asn Thr Val Tyr Phe Tyr Leu Gly	
500 505 510	
ggt tta aaa gaa tat cgc agt gcg cta gcc aac tgc aaa aac cat gcc	1584
Gly Leu Lys Glu Tyr Arg Ser Ala Leu Ala Asn Cys Lys Asn His Ala	
515 520 525	
tat gaa ggt ttt gat att caa tta caa cgt tca gat atc aag caa cct	1632
Tyr Glu Gly Phe Asp Ile Gln Leu Gln Arg Ser Asp Ile Lys Gln Pro	
530 535 540	
gcc aat gcc taa	1644
Ala Asn Ala	
545	

&lt;210&gt; 20

&lt;211&gt; 547

&lt;212&gt; PRT

&lt;213&gt; Acinetobacter sp. SE19

&lt;400&gt; 20

Met Glu Ile Ile Met Ser Gln Lys Met Asp Phe Asp Ala Ile Val Ile
1 5 10 15

Gly Gly Gly Phe Gly Gly Leu Tyr Ala Val Lys Lys Leu Arg Asp Glu
20 25 30

Leu Glu Leu Lys Val Gln Ala Phe Asp Lys Ala Thr Asp Val Ala Gly
35 40 45

Thr Trp Tyr Trp Asn Arg Tyr Pro Gly Ala Leu Ser Asp Thr Glu Thr



50	55	60
His Leu Tyr Cys Tyr Ser Trp Asp Lys Glu Leu Leu Gln Ser Leu Glu		
65	70	75 80
Ile Lys Lys Lys Tyr Val Gln Gly Pro Asp Val Arg Lys Tyr Leu Gln		
	85	90 95
Gln Val Ala Glu Lys His Asp Leu Lys Lys Ser Tyr Gln Phe Asn Thr		
	100	105 110
Ala Val Gln Ser Ala His Tyr Asn Glu Ala Asp Ala Leu Trp Glu Val		
	115	120 125
Thr Thr Glu Tyr Gly Asp Lys Tyr Thr Ala Arg Phe Leu Ile Thr Ala		
	130	135 140
Leu Gly Leu Leu Ser Ala Pro Asn Leu Pro Asn Ile Lys Gly Ile Asn		
	145	150 155 160
Gln Phe Lys Gly Glu Leu His His Thr Ser Arg Trp Pro Asp Asp Val		
	165	170 175
Ser Phe Glu Gly Lys Arg Val Gly Val Ile Gly Thr Gly Ser Thr Gly		
	180	185 190
Val Gln Val Ile Thr Ala Val Ala Pro Leu Ala Lys His Leu Thr Val		
	195	200 205
Phe Gln Arg Ser Ala Gln Tyr Ser Val Pro Ile Gly Asn Asp Pro Leu		
	210	215 220
Ser Glu Glu Asp Val Lys Lys Ile Lys Asp Asn Tyr Asp Lys Ile Trp		
	225	230 235 240
Asp Gly Val Trp Asn Ser Ala Leu Ala Phe Gly Leu Asn Glu Ser Thr		
	245	250 255
Val Pro Ala Met Ser Val Ser Ala Glu Glu Arg Lys Ala Val Phe Glu		
	260	265 270
Lys Ala Trp Gln Thr Gly Gly Gly Phe Arg Phe Met Phe Glu Thr Phe		
	275	280 285
Gly Asp Ile Ala Thr Asn Met Glu Ala Asn Ile Glu Ala Gln Asn Phe		
	290	295 300

Ile Lys Gly Lys Ile Ala Glu Ile Val Lys Asp Pro Ala Ile Ala Gln  
 305 310 315 320

Lys Leu Met Pro Gln Asp Leu Tyr Ala Lys Arg Pro Leu Cys Asp Ser  
 325 330 335

Gly Tyr Tyr Asn Thr Phe Asn Arg Asp Asn Val Arg Leu Glu Asp Val  
 340 345 350

Lys Ala Asn Pro Ile Val Glu Ile Thr Glu Asn Gly Val Lys Leu Glu  
 355 360 365

Asn Gly Asp Phe Val Glu Leu Asp Met Leu Ile Cys Ala Thr Gly Phe  
 370 375 380

Asp Ala Val Asp Gly Asn Tyr Val Arg Met Asp Ile Gln Gly Lys Asn  
 385 390 395 400

Gly Leu Ala Met Lys Asp Tyr Trp Lys Glu Gly Pro Ser Ser Tyr Met  
 405 410 415

Gly Val Thr Val Asn Asn Tyr Pro Asn Met Phe Met Val Leu Gly Pro  
 420 425 430

Asn Gly Pro Phe Thr Asn Leu Pro Pro Ser Ile Glu Ser Gln Val Glu  
 435 440 445

Trp Ile Ser Asp Thr Ile Gln Tyr Thr Val Glu Asn Asn Val Glu Ser  
 450 455 460

Ile Glu Ala Thr Lys Glu Ala Glu Glu Gln Trp Thr Gln Thr Cys Ala  
 465 470 475 480

Asn Ile Ala Glu Met Thr Leu Phe Pro Lys Ala Gln Ser Trp Ile Phe  
 485 490 495

Gly Ala Asn Ile Pro Gly Lys Lys Asn Thr Val Tyr Phe Tyr Leu Gly  
 500 505 510

Gly Leu Lys Glu Tyr Arg Ser Ala Leu Ala Asn Cys Lys Asn His Ala  
 515 520 525

Tyr Glu Gly Phe Asp Ile Gln Leu Gln Arg Ser Asp Ile Lys Gln Pro  
 530 535 540

Ala Asn Ala  
545

<210> 21

<211> 1320

<212> DNA

<213> *Rhodococcus erythropolis* AN12

<400> 21

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atgagcacag agggcaagta cgcgctgata ggagcgggtc cgtctggatt ggccggcgcg      60
cgaaacctcg atcgagccgg catagcgctc gacggcttcg agagccacga cgacgtcggg      120
gggctctggg acatcgacaa cccgcacagc accgtctacg agtcggcgca cctcatttcg      180
tcgaagggca ccaccgcatt cgcggagttc ccgatggcgg attcggttgc cgactaccgc      240
agccacatcg aacttgccga gtatttcgcg gactacgcgc ataccacga tcttcgcagg      300
cactttgcct tcggcactac cgtcatcgac gttttgccgg tcgattcgct gtggcaggtc      360
accacgcgta gtgcgagcgg tgagaactca gtcgcggcgg atcgaggcgt gatcatcgcg      420
aacggaacgc tgtcgaagcc gaacataccg acgttcgggg gcgacttcac cggcacgttg      480
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ggagcgggca acagtggatg cgacatcgcc gtcgatgccg tccaccaggc cgagtgcgtc      600
gatttgagcg ttccggcgagg ctactacttc gtccccaagt atctgttcgg gcgaccctcg      660
gacacgttga atcagggaaa gccgttgccg ccgtggatca aacaacgcgt cgacaccttg      720
ttactcaagc agttcacggg agatccgggt cggttccgat ttccggcacc ggactacaag      780
atctacgaat cgcattccgt cgtgaactcg ttgatcctgc accacatcgg gcacgggtgac      840
gtgcacgtgc gcgccgacgt cgaccggttc gaggggaaga cggtgcggtt tgtcgacgga      900
tcgtctgccg actacgacct cgttctctgc gccacggggg atcacctcga ctatcccttc      960
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cagggtcggt accagcaggc cgagttgggt gccaaattga tcaccgcacg caccgaagcc     1140
cccgcgcgcg cgcgcgaatt ctccgcagcg gcggccggcc ctctctccga tctgtccggg     1200
ggatacaagt acctgaagct gggacgaatg gcctactacg tgaacaagga cgcctaccga     1260
tcggcgatca gacggcacat cggactgctc gatgccgctc tgacgaaggg aggtcagtga     1320

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&lt;210&gt; 22

&lt;211&gt; 439

&lt;212&gt; PRT

&lt;213&gt; Rhodococcus erythropolis AN12

&lt;400&gt; 22

Met Ser Thr Glu Gly Lys Tyr Ala Leu Ile Gly Ala Gly Pro Ser Gly  
 1 5 10 15

Leu Ala Gly Ala Arg Asn Leu Asp Arg Ala Gly Ile Ala Phe Asp Gly  
 20 25 30

Phe Glu Ser His Asp Asp Val Gly Gly Leu Trp Asp Ile Asp Asn Pro  
 35 40 45

His Ser Thr Val Tyr Glu Ser Ala His Leu Ile Ser Ser Lys Gly Thr  
 50 55 60

Thr Ala Phe Ala Glu Phe Pro Met Ala Asp Ser Val Ala Asp Tyr Pro  
 65 70 75 80

Ser His Ile Glu Leu Ala Glu Tyr Phe Arg Asp Tyr Ala Asp Thr His  
 85 90 95

Asp Leu Arg Arg His Phe Ala Phe Gly Thr Thr Val Ile Asp Val Leu  
 100 105 110

Pro Val Asp Ser Leu Trp Gln Val Thr Thr Arg Ser Arg Ser Gly Glu  
 115 120 125

Thr Ser Val Ala Arg Tyr Arg Gly Val Ile Ile Ala Asn Gly Thr Leu  
 130 135 140

Ser Lys Pro Asn Ile Pro Thr Phe Arg Gly Asp Phe Thr Gly Thr Leu  
 145 150 155 160

Met His Thr Ser Glu Tyr Arg Ser Ala Glu Ile Phe Arg Gly Lys Arg  
 165 170 175

Val Leu Val Ile Gly Ala Gly Asn Ser Gly Cys Asp Ile Ala Val Asp  
 180 185 190

Ala Val His Gln Ala Glu Cys Val Asp Leu Ser Val Arg Arg Gly Tyr

195	200	205
Tyr Phe Val Pro Lys Tyr Leu Phe Gly Arg Pro Ser Asp Thr Leu Asn		
210	215	220
Gln Gly Lys Pro Leu Pro Pro Trp Ile Lys Gln Arg Val Asp Thr Leu		
225	230	235
Leu Leu Lys Gln Phe Thr Gly Asp Pro Val Arg Phe Gly Phe Pro Ala		
	245	250
Pro Asp Tyr Lys Ile Tyr Glu Ser His Pro Val Val Asn Ser Leu Ile		
	260	265
Leu His His Ile Gly His Gly Asp Val His Val Arg Ala Asp Val Asp		
	275	280
Arg Phe Glu Gly Lys Thr Val Arg Phe Val Asp Gly Ser Ser Ala Asp		
290	295	300
Tyr Asp Leu Val Leu Cys Ala Thr Gly Tyr His Leu Asp Tyr Pro Phe		
305	310	315
Ile Ala Arg Glu Asp Leu Asp Trp Ser Gly Ala Ala Pro Asp Leu Phe		
	325	330
Leu Asn Val Ala Ser Arg Arg His Asp Asn Leu Phe Val Leu Gly Met		
	340	345
Val Glu Ala Ser Gly Leu Gly Trp Gln Gly Arg Tyr Gln Gln Ala Glu		
	355	360
Leu Val Ala Lys Leu Ile Thr Ala Arg Thr Glu Ala Pro Ala Ala Ala		
	370	375
Arg Glu Phe Ser Ala Ala Ala Ala Gly Pro Pro Pro Asp Leu Ser Gly		
385	390	395
Gly Tyr Lys Tyr Leu Lys Leu Gly Arg Met Ala Tyr Tyr Val Asn Lys		
	405	410
Asp Ala Tyr Arg Ser Ala Ile Arg Arg His Ile Gly Leu Leu Asp Ala		
	420	425
Ala Leu Thr Lys Gly Gly Gln		
435		

&lt;210&gt; 23

&lt;211&gt; 1557

&lt;212&gt; DNA

<213> *Rhodococcus erythropolis* AN12

&lt;400&gt; 23

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cgaacacgag tcgtcgtcat cggagccggt ttcggcggca tcggaacggc tgtccgcttg      120
aagcagtcgc ggatcgacga cttcgtcggt ctggaacgtg ccgcgaggcc cgggggggacc      180
tggcaggtca atacctaccc cggtgcacag tgcgacatcc cgtcgattct gtactcgttc      240
tcgtttgcgc ccaatccgaa ctggacgcgg ctgtatcccc tgcagcccgga gatctacgac      300
tatctccggg attgcgtcca tcgcttcgga ctggccgggtc atttccactg caaccaggac      360
gtgacagaag cttcgtggga cgagcaagcc cagatctggc gggtagacac tgcggaaacc      420
gtctgggagg cacagtctct ggtcgcggcc accggcccggt tcagtgcgcc cgccacaccc      480
gaccttcccg ggctcgaatc gtttcgtggt cagatgttcc acaccgcgga ctggaaccac      540
gaccacgacc ttcgcggtga gcggatagcc gtggtcggca ccggcgcctc tgcggtgcag      600
atcatcccca gactgcaacc gtcgcgggac acgttgaccg tgttccagcg gacaccgacg      660
tggatcctgc cgcacccgga tcagccgatg accggctggc caagcgctct cttcgagcgg      720
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cccggattcg tgtacaagcc gtcactgctc aaagggtggc ccgactcgg ccgagcacac      840
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gaggtggtga cggacggaat cgtcgaggtg caggagcgcg gagttctcac cgcggacggc     1020
gccttccggg aagtcgacac catagtcatg ggaaccggct ttcggatggg agacaacccg     1080
tcgttcgaca ccatccgagg ccaggacggc cgcagcctcg cacagacgtg gaacggcagt     1140
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gtgcaacgcg agttcgtacg cgcgacagac cgcgactcg ccaccagcgt gtggaacgcc     1380
ggcgggtgca gtagttacta cctcgtcgac ggcggtcgca actacacctt ctatcccgga     1440

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 caaccctgtct cgtccgcagc actcaccact gctcgagaaa ccgtgaggag ccgataa 1557

<210> 24

<211> 518

<212> PRT

<213> Rhodococcus erythropolis AN12

<400> 24

Met Val Asp Ile Asp Pro Thr Ser Gly Pro Ser Ala Gly Asp Glu Glu  
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Thr Arg Thr Arg Arg Thr Arg Val Val Val Ile Gly Ala Gly Phe Gly  
 20 25 30

Gly Ile Gly Thr Ala Val Arg Leu Lys Gln Ser Gly Ile Asp Asp Phe  
 35 40 45

Val Val Leu Glu Arg Ala Ala Glu Pro Gly Gly Thr Trp Gln Val Asn  
 50 55 60

Thr Tyr Pro Gly Ala Gln Cys Asp Ile Pro Ser Ile Leu Tyr Ser Phe  
 65 70 75 80

Ser Phe Ala Pro Asn Pro Asn Trp Thr Arg Leu Tyr Pro Leu Gln Pro  
 85 90 95

Glu Ile Tyr Asp Tyr Leu Arg Asp Cys Val His Arg Phe Gly Leu Ala  
 100 105 110

Gly His Phe His Cys Asn Gln Asp Val Thr Glu Ala Ser Trp Asp Glu  
 115 120 125

Gln Ala Gln Ile Trp Arg Val His Thr Ala Glu Thr Val Trp Glu Ala  
 130 135 140

Gln Phe Leu Val Ala Ala Thr Gly Pro Phe Ser Ala Pro Ala Thr Pro  
 145 150 155 160

Asp Leu Pro Gly Leu Glu Ser Phe Arg Gly Gln Met Phe His Thr Ala  
 165 170 175

Asp Trp Asn His Asp His Asp Leu Arg Gly Glu Arg Ile Ala Val Val  
 180 185 190

Gly Thr Gly Ala Ser Ala Val Gln Ile Ile Pro Arg Leu Gln Pro Leu  
 195 200 205

Ala Asp Thr Leu Thr Val Phe Gln Arg Thr Pro Thr Trp Ile Leu Pro  
 210 215 220

His Pro Asp Gln Pro Met Thr Gly Trp Pro Ser Ala Leu Phe Glu Arg  
 225 230 235 240

Val Pro Leu Thr Gln Arg Leu Ala Arg Lys Gly Leu Asp Leu Leu Gln  
 245 250 255

Glu Ala Leu Val Pro Gly Phe Val Tyr Lys Pro Ser Leu Leu Lys Gly  
 260 265 270

Leu Ala Ala Leu Gly Arg Ala His Leu Arg Arg Gln Val Arg Asp Pro  
 275 280 285

Glu Leu Arg Ala Lys Leu Leu Pro His Tyr Ala Phe Gly Cys Lys Arg  
 290 295 300

Pro Thr Phe Ser Asn Thr Tyr Tyr Pro Ala Leu Ala Ser Pro Asn Val  
 305 310 315 320

Glu Val Val Thr Asp Gly Ile Val Glu Val Gln Glu Arg Gly Val Leu  
 325 330 335

Thr Ala Asp Gly Ala Phe Arg Glu Val Asp Thr Ile Val Met Gly Thr  
 340 345 350

Gly Phe Arg Met Gly Asp Asn Pro Ser Phe Asp Thr Ile Arg Gly Gln  
 355 360 365

Asp Gly Arg Ser Leu Ala Gln Thr Trp Asn Gly Ser Ala Glu Ala Phe  
 370 375 380

Leu Gly Thr Thr Ile Ser Gly Phe Pro Asn Phe Phe Met Ile Leu Gly  
 385 390 395 400

Pro Asn Ser Val Val Tyr Thr Ser Gln Val Val Thr Ile Glu Ala Gln  
 405 410 415

Val Glu Tyr Ile Val Ser Cys Ile Leu Gln Met Asp Glu Arg Gly Ile



420                      425                      430  
 Gly Ser Ile Asp Val Arg Ala Asp Val Gln Arg Glu Phe Val Arg Ala  
          435                      440                      445  
 Thr Asp Arg Arg Leu Ala Thr Ser Val Trp Asn Ala Gly Gly Cys Ser  
          450                      455                      460  
 Ser Tyr Tyr Leu Val Asp Gly Gly Arg Asn Tyr Thr Phe Tyr Pro Gly  
          465                      470                      475                      480  
 Phe Asn Arg Ser Phe Arg Ala Arg Thr Lys Arg Ala Asp Leu Ala His  
                          485                      490                      495  
 Tyr Ala Gln Val Gln Pro Val Ser Ser Ala Ala Leu Thr Thr Ala Arg  
                          500                      505                      510  
 Glu Thr Val Arg Ser Arg  
                          515

&lt;210&gt; 25

&lt;211&gt; 1626

&lt;212&gt; DNA

&lt;213&gt; Rhodococcus erythropolis AN12

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 gcgggctccg gagtgggcgg cacgtggtat ttcaaccgct accccggcgc acgctgcgac 180  
 gtcgagagtt tcgactactc ctactcgttc tccgaagagc tgcaacagga ttgggactgg 240  
 agcgagaagt acgccgcgca accggagatc ctctcgtacc tcgatcacgt ggctgatcgc 300  
 ttcgacctac gacttggett caccttcgac acacgcgttc tgagcgcaca gttcgacgag 360  
 ggtactgcca cgtggcgagt acagaccgac ggcgggtcacg acgtcacctc acgcttcgtc 420  
 gtgtgcgcca cgggcagcct ctcgaccgca aacgttccga acattgcggg ccgtgagacc 480  
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 aaacgcgtcg gcgtgatcgg caccggatcc tcgggcatcc agtccattcc gctgatcgcc 600  
 gagcaggccg atcatctcta cgtgttccag cgggtccgca attacagtgt gccggcagga 660  
 aacacgcctc tcgatgacaa gcgccgcgcc gagatcaagg ccggctacgc agagcgctga 720

gcgctgtcca agcgagtggt cgggtggatcg ccgttcggtt cggatcctcg cagcgccctc 780  
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 gcgtttgccg aacgcaagat tcgctcggaa gtccaggatc aggcgatcgc cgacctgctc 960  
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 gagacgttgc gcgagaactg gcatgcgggt ccaaggacgt atctaggcct cggagtacac 1260  
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 gcgggcatcg acaccatcga accgagtgcc gaagccgtgg acaactggct cgacgaatgc 1440  
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 attccgggaa agccgaggat attcatgcca ttcacggag gattcgggtg ctactccgac 1560  
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 gcatga 1626

<210> 26

<211> 541

<212> PRT

<213> Rhodococcus erythropolis AN12

<400> 26

Met Thr Asp Pro Asp Phe Ser Thr Ala Pro Leu Asp Val Val Val Ile  
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Gly Ala Gly Val Ala Gly Met Tyr Ala Met His Arg Leu Arg Glu Gln  
 20 25 30

Gly Leu Arg Val His Gly Phe Glu Ala Gly Ser Gly Val Gly Gly Thr  
 35 40 45

Trp Tyr Phe Asn Arg Tyr Pro Gly Ala Arg Cys Asp Val Glu Ser Phe  
 50 55 60

Asp Tyr Ser Tyr Ser Phe Ser Glu Glu Leu Gln Gln Asp Trp Asp Trp  
 65 70 75 80  
 Ser Glu Lys Tyr Ala Ala Gln Pro Glu Ile Leu Ser Tyr Leu Asp His  
 85 90 95  
 Val Ala Asp Arg Phe Asp Leu Arg Thr Gly Phe Thr Phe Asp Thr Arg  
 100 105 110  
 Val Leu Ser Ala Gln Phe Asp Glu Gly Thr Ala Thr Trp Arg Val Gln  
 115 120 125  
 Thr Asp Gly Gly His Asp Val Thr Ser Arg Phe Val Val Cys Ala Thr  
 130 135 140  
 Gly Ser Leu Ser Thr Ala Asn Val Pro Asn Ile Ala Gly Arg Glu Thr  
 145 150 155 160  
 Phe Gly Gly Asp Val Phe His Thr Gly Phe Trp Pro His Glu Gly Val  
 165 170 175  
 Asp Phe Thr Gly Lys Arg Val Gly Val Ile Gly Thr Gly Ser Ser Gly  
 180 185 190  
 Ile Gln Ser Ile Pro Leu Ile Ala Glu Gln Ala Asp His Leu Tyr Val  
 195 200 205  
 Phe Gln Arg Ser Ala Asn Tyr Ser Val Pro Ala Gly Asn Thr Pro Leu  
 210 215 220  
 Asp Asp Lys Arg Arg Ala Glu Ile Lys Ala Gly Tyr Ala Glu Arg Arg  
 225 230 235 240  
 Ala Leu Ser Lys Arg Ser Gly Gly Gly Ser Pro Phe Val Ser Asp Pro  
 245 250 255  
 Arg Ser Ala Leu Glu Val Ser Glu Ala Glu Arg Asn Ala Ala Tyr Glu  
 260 265 270  
 Glu Arg Trp Lys Leu Gly Gly Val Leu Phe Ala Lys Thr Phe Ala Asp  
 275 280 285  
 Gln Thr Ser Asn Ile Glu Ala Asn Gly Thr Ala Ala Ala Phe Ala Glu  
 290 295 300

Arg Lys Ile Arg Ser Glu Val Gln Asp Gln Ala Ile Ala Asp Leu Leu  
 305 310 315 320

Ile Pro Asn Asp His Pro Ile Gly Thr Lys Arg Ile Val Thr Asp Thr  
 325 330 335

Asn Tyr Tyr Gln Ser Tyr Asn Arg Asp Asn Val Ser Leu Val Asp Leu  
 340 345 350

Lys Ser Ala Pro Ile Glu Ala Ile Asp Glu Ala Gly Ile Lys Thr Ala  
 355 360 365

Asp Ala His Tyr Glu Leu Asp Ala Leu Val Phe Ala Thr Gly Phe Asp  
 370 375 380

Ala Met Thr Gly Ala Leu Asp Arg Ile Glu Ile Arg Gly Arg Asn Gly  
 385 390 395 400

Glu Thr Leu Arg Glu Asn Trp His Ala Gly Pro Arg Thr Tyr Leu Gly  
 405 410 415

Leu Gly Val His Gly Phe Pro Asn Leu Phe Ile Val Thr Gly Pro Gly  
 420 425 430

Ser Pro Ser Val Leu Ser Asn Met Ile Leu Ala Ala Glu Gln His Val  
 435 440 445

Asp Trp Ile Ala Gly Ala Ile Asn His Leu Asp Ser Ala Gly Ile Asp  
 450 455 460

Thr Ile Glu Pro Ser Ala Glu Ala Val Asp Asn Trp Leu Asp Glu Cys  
 465 470 475 480

Ser Arg Arg Ala Ser Ala Thr Leu Phe Pro Ser Ala Asn Ser Trp Tyr  
 485 490 495

Met Gly Ala Asn Ile Pro Gly Lys Pro Arg Ile Phe Met Pro Phe Ile  
 500 505 510

Gly Gly Phe Gly Val Tyr Ser Asp Ile Cys Ala Asp Val Ala Ala Ala  
 515 520 525

Gly Tyr Arg Gly Phe Glu Leu Asn Ser Ala Val His Ala  
 530 535 540

&lt;210&gt; 27

&lt;211&gt; 1389

&lt;212&gt; DNA

<213> *Rhodococcus erythropolis* AN12

&lt;400&gt; 27

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gtcggcggaa actggtacta caagaacccc aacggaatgt cggcctgcta ccagagcctg     180
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ctcatgcaca gccacagcta caacgacccg ttcgatccga tcgacatgcg cggcaagaaa     540
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&lt;210&gt; 28

&lt;211&gt; 462

&lt;212&gt; PRT

&lt;213&gt; Rhodococcus erythropolis AN12

&lt;400&gt; 28

Met Ser Pro Ser Pro Leu Pro Ser Val Cys Ile Ile Gly Ala Gly Pro  
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Thr Gly Ile Thr Thr Ala Lys Arg Met Lys Glu Phe Gly Ile Pro Phe  
 20 25 30

Asp Cys Tyr Glu Ala Ser Asp Glu Val Gly Gly Asn Trp Tyr Tyr Lys  
 35 40 45

Asn Pro Asn Gly Met Ser Ala Cys Tyr Gln Ser Leu His Ile Asp Thr  
 50 55 60

Ser Lys Trp Arg Leu Ala Phe Glu Asp Phe Pro Val Ser Ala Asp Leu  
 65 70 75 80

Pro Asp Phe Pro His His Ser Glu Leu Phe Gln Tyr Phe Lys Asp Tyr  
 85 90 95

Val Glu His Phe Gly Leu Arg Glu Ser Ile Ile Phe Asn Thr Ser Val  
 100 105 110

Val Ala Ala Glu Arg Asp Ala Asn Gly Leu Trp Thr Val Thr Arg Ser  
 115 120 125

Asp Gly Glu Val Arg Thr Tyr Asp Val Leu Met Val Cys Asn Gly His  
 130 135 140

His Trp Asp Pro Asn Ile Pro Asp Tyr Pro Gly Glu Phe Asp Gly Val  
 145 150 155 160

Leu Met His Ser His Ser Tyr Asn Asp Pro Phe Asp Pro Ile Asp Met  
 165 170 175

Arg Gly Lys Lys Val Val Val Val Gly Met Gly Asn Ser Gly Leu Asp  
 180 185 190

Ile Ala Ser Glu Leu Gly Gln Arg Tyr Leu Ala Asp Lys Leu Ile Val

195	200	205
Ser Ala Arg Arg Gly Val Trp Val Leu Pro Lys Tyr Leu Gly Gly Val		
210	215	220
Pro Gly Asp Lys Leu Ile Thr Pro Pro Trp Met Pro Arg Gly Leu Arg		
225	230	235 240
Leu Phe Leu Ser Arg Arg Phe Leu Gly Lys Asn Leu Gly Thr Met Glu		
	245	250 255
Gly Tyr Gly Leu Pro Lys Pro Asp His Arg Pro Phe Glu Ala His Pro		
	260	265 270
Ser Ala Ser Gly Glu Phe Leu Gly Arg Ala Gly Ser Gly Asp Ile Thr		
	275	280 285
Phe Lys Pro Ala Ile Thr Lys Leu Asp Gly Lys Gln Val His Phe Ala		
	290	295 300
Asp Gly Thr Ala Glu Asp Val Asp Val Val Val Cys Ala Thr Gly Tyr		
305	310	315 320
Asn Ile Ser Phe Pro Phe Phe Asp Asp Pro Asn Leu Leu Pro Asp Lys		
	325	330 335
Asp Asn Arg Phe Pro Leu Phe Lys Arg Met Met Lys Pro Gly Ile Asp		
	340	345 350
Asn Leu Phe Phe Met Gly Leu Ala Gln Pro Met Pro Thr Leu Val Asn		
	355	360 365
Phe Ala Glu Gln Gln Ser Lys Leu Val Ala Ala Tyr Leu Thr Gly Lys		
	370	375 380
Tyr Gln Leu Pro Ser Ala Asn Glu Met Gln Glu Ile Thr Lys Ala Asp		
385	390	395 400
Glu Ala Tyr Phe Leu Ala Pro Tyr Tyr Lys Ser Pro Arg His Thr Ile		
	405	410 415
Gln Leu Glu Phe Asp Pro Tyr Val Arg Asn Met Asn Lys Glu Ile Ala		
	420	425 430
Lys Gly Thr Lys Arg Ala Ala Ala Ser Gly Asn Lys Leu Pro Val Ala		
	435	440 445

Ala Arg Ala Ala Ala His Glu Leu Glu Lys Ala Asp Arg Ala  
 450 455 460

<210> 29

<211> 1572

<212> DNA

<213> *Rhodococcus erythropolis* AN12

<400> 29

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gactccctcg gtggaacctg gcgcgccaac acctatcccg ggtgcgcctg cgacgttcca      180
tccgggtctgt actcgtaactc ctttgccgcc aatccggatt ggacgcgctt gttcgcggag      240
caaccggaga tccgcgaata catcgagaac acggcgggca cgcacggagt cgacaaacac      300
gttcgcttcg gggtcgaaat gctctccgcg cgatgggatg cgtcgcaatc actgtggaag      360
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ggagcgtcgg cagtccagtt cgttcgcgc atcgtctccc aggtctccgc ccttcacctc      600
taccagcgaa ccgctcaatg ggttctcccc aaaccgatac actacgtacc gcggatcgaa      660
aggtccgtca tgcgattcgt gccgggagca cagaaagcct tgcgcagcat cgaatacggg      720
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gccaactga actatgtggc gcaggcaatc ggccaagccc gtcgtcacgg ctggcagacc     1260
atcgacgtgc gagaggaagt tcaggcagcc ttcaattctc aggttcagga ggcattgggg     1320

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accacggtct acaacgccgg tgggtgcgaa agctatttct tcgacgtcaa cggccgcaac 1380  
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 tatgcctaca accacacgtc gaacctgag tcagacaaca cggccctga acccagcca 1500  
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<210> 30

<211> 523

<212> PRT

<213> Rhodococcus erythropolis AN12

<400> 30

Val Asn Asn Glu Ser Asp His Phe Glu Val Val Ile Ile Gly Gly Gly  
 1 5 10 15

Ile Ser Gly Ile Gly Ala Ala Ile His Leu Gln Arg Leu Gly Ile Asp  
 20 25 30

Asn Phe Ala Leu Leu Glu Lys Ala Asp Ser Leu Gly Gly Thr Trp Arg  
 35 40 45

Ala Asn Thr Tyr Pro Gly Cys Ala Cys Asp Val Pro Ser Gly Leu Tyr  
 50 55 60

Ser Tyr Ser Phe Ala Ala Asn Pro Asp Trp Thr Arg Leu Phe Ala Glu  
 65 70 75 80

Gln Pro Glu Ile Arg Glu Tyr Ile Glu Asn Thr Ala Gly Thr His Gly  
 85 90 95

Val Asp Lys His Val Arg Phe Gly Val Glu Met Leu Ser Ala Arg Trp  
 100 105 110

Asp Ala Ser Gln Ser Leu Trp Lys Ile Thr Thr Ser Ser Gly Glu Leu  
 115 120 125

Thr Ala Arg Phe Val Ile Ala Ala Ala Gly Pro Trp Asn Glu Pro Leu  
 130 135 140

Thr Pro Ala Ile Pro Gly Leu Glu Ala Phe Glu Gly Glu Val Phe His  
 145 150 155 160

Ser Ser Gln Trp Asn His Asp Tyr Asp Leu Thr Gly Lys Leu Val Ala  
 165 170 175

Val Val Gly Thr Gly Ala Ser Ala Val Gln Phe Val Pro Arg Ile Val  
 180 185 190

Ser Gln Val Ser Ala Leu His Leu Tyr Gln Arg Thr Ala Gln Trp Val  
 195 200 205

Leu Pro Lys Pro Asp His Tyr Val Pro Arg Ile Glu Arg Ser Val Met  
 210 215 220

Arg Phe Val Pro Gly Ala Gln Lys Ala Leu Arg Ser Ile Glu Tyr Gly  
 225 230 235 240

Ile Met Glu Ala Leu Gly Leu Gly Phe Arg Asn Pro Trp Ile Leu Arg  
 245 250 255

Ile Val Gln Lys Leu Gly Ser Ala Gln Leu Arg Leu Gln Val Arg Asp  
 260 265 270

Pro Lys Leu Arg Lys Ala Leu Thr Pro Asp Tyr Thr Leu Gly Cys Lys  
 275 280 285

Arg Leu Leu Met Ser Asn Ser Tyr Tyr Pro Ala Leu Gly Lys Pro Asn  
 290 295 300

Val Ser Val His Ala Asn Ala Val Glu Gln Ile Arg Gly Asn Thr Val  
 305 310 315 320

Ile Gly Ala Asp Gly Val Glu Ala Glu Val Asp Ala Ile Ile Phe Gly  
 325 330 335

Thr Gly Phe His Ile Leu Asp Met Pro Ile Ala Ser Lys Val Phe Asp  
 340 345 350

Gly Glu Gly Arg Ser Leu Asp Asp His Trp Gln Gly Ser Pro Gln Ala  
 355 360 365

Tyr Phe Gly Ser Ala Val Ser Gly Phe Pro Asn Ala Phe Ile Leu Leu  
 370 375 380

Gly Pro Ser Leu Gly Thr Gly His Thr Ser Ala Phe Met Ile Leu Glu  
 385 390 395 400

Ala Gln Leu Asn Tyr Val Ala Gln Ala Ile Gly His Ala Arg Arg His  
405 410 415

Gly Trp Gln Thr Ile Asp Val Arg Glu Glu Val Gln Ala Ala Phe Asn  
420 425 430

Ser Gln Val Gln Glu Ala Leu Gly Thr Thr Val Tyr Asn Ala Gly Gly  
435 440 445

Cys Glu Ser Tyr Phe Phe Asp Val Asn Gly Arg Asn Ser Phe Asn Trp  
450 455 460

Pro Trp Ser Ser Gly Ala Met Arg Arg Arg Leu Arg Asp Phe Asp Pro  
465 470 475 480

Tyr Ala Tyr Asn His Thr Ser Asn Pro Glu Ser Asp Asn Thr Pro Pro  
485 490 495

Glu Pro Thr Pro Ser Glu Pro Thr Pro Ser Glu Pro Thr Pro Ser Glu  
500 505 510

Pro Thr Thr Ser Pro Glu Pro Glu Tyr Thr Ala  
515 520

<210> 31

<211> 1482

<212> DNA

<213> Rhodococcus erythropolis AN12

<400> 31

atgagcaccg aacacctcga tgcctcgatc gtcggcgccg gcttggtccgg catcggtgct	60
gcttatcgac tccagaccga gctcccagga aagtcgtacg caatcctcga ggcccagacg	120
aacagcggcg gaacctggga cctcttcaag tatcccggca tccgatcgga ttccgacatg	180
ttcacgctcg gctacccgtt tcgcccgtgg acagatgcc aagcaatcgc cgacggtgat	240
tcgatcctgc ggtacgtgcg cgacaccgcg cgagagaacg ggatcgacaa gaagattcgg	300
tacaaccgga aggtgacggc cgcacatcatg tcgtcagcga cctcgacctg gacagtcacg	360
gtcacgaccg ggcacgaaga cgaaacattg acctgtaact tcctctatct ctgcagcggg	420
tactacagct acgacggcgg atacaccccc gacttccccg gacgtgaatc gtttgccggt	480
gaggtagtgc acccccagtt ctggcccga gaactcgatt actccgacaa gaaggtcggt	540

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gtgatcggaa gcggcgccac cgcagtcact ttggtcccca cgatgtcacg ggacgcaagc   600
cacgtcacga tgctccagcg atcaccgacg tacattctgg cgcttccgtc cagcgacaaa   660
ttatcggaca ccattcgcg cgtactgccg aatcaactcg cgcacagcat cgctcgatgg   720
aagagcgtcg tagtgaacct gagtttctac caactgtgcc gacgcagtcc ggcgcggtgca   780
aagaggatgc tgaacctcgc gatcagtcgt caactcccga aagacatccc cctcgatcct   840
cacttcacac cctcctacga tccctgggac cagcgcttgt gcgtcgtacc cgacggcgat   900
ttgttcaaag ccctccgac cggcaaggcc tcgatcgaga ccgatcacat cgacaccttc   960
accgagaccg ggatccttct cgcgtcaggt cgcgaactcg aagctgacat catcgtcact  1020
gcaacaggat tgaagatgga ggcggtcggc gggatgtcca tcgaagtgga cggcgaactc  1080
gtcaccctcg gtgatcggtta cgcctacaag ggcgatgatga tcagcgacgt accgaacttc  1140
gcgatgtgcg tcggctacac caacgcctcg tggactctgc gagcagatct cacgtcgatg  1200
tacgtgtgcc gactgctgac ggagatggac aagcgcgact attcgaagtg cgtgccgcac  1260
gcgaccgaag aaatggacca gcggccgac ctggatctgg cgtcggggta cgtcatgcgt  1320
gccgtggaac agttcccga gacgggatcg aagtcaccgt ggaacatgcg tcagaactac  1380
atccttgacc gtcttcactc cacgttcggg agcatcaacg accacatgac gttctcgaag  1440
gcaccagctc gacattcgac gccggtaccg agcaagagtt ga                      1482

```

<210> 32

<211> 493

<212> PRT

<213> Rhodococcus erythropolis AN12

<400> 32

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Met Ser Thr Glu His Leu Asp Val Leu Ile Val Gly Ala Gly Leu Ser
1           5           10          15

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```

Gly Ile Gly Ala Ala Tyr Arg Leu Gln Thr Glu Leu Pro Gly Lys Ser
          20          25          30

```

```

Tyr Ala Ile Leu Glu Ala Arg Ala Asn Ser Gly Gly Thr Trp Asp Leu
35          40          45

```

```

Phe Lys Tyr Pro Gly Ile Arg Ser Asp Ser Asp Met Phe Thr Leu Gly
50          55          60

```

Tyr Pro Phe Arg Pro Trp Thr Asp Ala Lys Ala Ile Ala Asp Gly Asp  
 65 70 75 80

Ser Ile Leu Arg Tyr Val Arg Asp Thr Ala Arg Glu Asn Gly Ile Asp  
 85 90 95

Lys Lys Ile Arg Tyr Asn Arg Lys Val Thr Ala Ala Ser Trp Ser Ser  
 100 105 110

Ala Thr Ser Thr Trp Thr Val Thr Val Thr Thr Gly Asp Glu Asp Glu  
 115 120 125

Thr Leu Thr Cys Asn Phe Leu Tyr Leu Cys Ser Gly Tyr Tyr Ser Tyr  
 130 135 140

Asp Gly Gly Tyr Thr Pro Asp Phe Pro Gly Arg Glu Ser Phe Ala Gly  
 145 150 155 160

Glu Val Val His Pro Gln Phe Trp Pro Glu Glu Leu Asp Tyr Ser Asp  
 165 170 175

Lys Lys Val Val Val Ile Gly Ser Gly Ala Thr Ala Val Thr Leu Val  
 180 185 190

Pro Thr Met Ser Arg Asp Ala Ser His Val Thr Met Leu Gln Arg Ser  
 195 200 205

Pro Thr Tyr Ile Leu Ala Leu Pro Ser Ser Asp Lys Leu Ser Asp Thr  
 210 215 220

Ile Arg Ala Val Leu Pro Asn Gln Leu Ala His Ser Ile Ala Arg Trp  
 225 230 235 240

Lys Ser Val Val Val Asn Leu Ser Phe Tyr Gln Leu Cys Arg Arg Ser  
 245 250 255

Pro Ala Arg Ala Lys Arg Met Leu Asn Leu Ala Ile Ser Arg Gln Leu  
 260 265 270

Pro Lys Asp Ile Pro Leu Asp Pro His Phe Thr Pro Ser Tyr Asp Pro  
 275 280 285

Trp Asp Gln Arg Leu Cys Val Val Pro Asp Gly Asp Leu Phe Lys Ala  
 290 295 300

Leu Arg Ser Gly Lys Ala Ser Ile Glu Thr Asp His Ile Asp Thr Phe  
 305 310 315 320

Thr Glu Thr Gly Ile Leu Leu Ala Ser Gly Arg Glu Leu Glu Ala Asp  
 325 330 335

Ile Ile Val Thr Ala Thr Gly Leu Lys Met Glu Ala Cys Gly Gly Met  
 340 345 350

Ser Ile Glu Val Asp Gly Glu Leu Val Thr Leu Gly Asp Arg Tyr Ala  
 355 360 365

Tyr Lys Gly Met Met Ile Ser Asp Val Pro Asn Phe Ala Met Cys Val  
 370 375 380

Gly Tyr Thr Asn Ala Ser Trp Thr Leu Arg Ala Asp Leu Thr Ser Met  
 385 390 395 400

Tyr Val Cys Arg Leu Leu Thr Glu Met Asp Lys Arg Asp Tyr Ser Lys  
 405 410 415

Cys Val Pro His Ala Thr Glu Glu Met Asp Gln Arg Pro Ile Leu Asp  
 420 425 430

Leu Ala Ser Gly Tyr Val Met Arg Ala Val Glu Gln Phe Pro Lys Gln  
 435 440 445

Gly Ser Lys Ser Pro Trp Asn Met Arg Gln Asn Tyr Ile Leu Asp Arg  
 450 455 460

Leu His Ser Thr Phe Gly Ser Ile Asn Asp His Met Thr Phe Ser Lys  
 465 470 475 480

Ala Pro Ala Arg His Ser Thr Pro Val Pro Ser Lys Ser  
 485 490

<210> 33

<211> 1620

<212> DNA

<213> Rhodococcus erythropolis AN12

<400> 33

atgacagacg aattcgacgt agtgcacgtg ggtgcaggtc tcgcaggtat gcagatgctg 60

```

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ggcacctggg attggaaccg ctacccgggt gctcgggtgtg acgtggagag tttggagtac    180
tcctatcagt tctccgaggt gctccaacag gaatgggaat ggacccgccg gtacgcagat    240
caggccgaga tcatgcgcta catcagccac gtcgtcgaaa ccttcgacct ggcccgcgac    300
atcaggtttc atacccgggt cgaggcgatg acctacgagg agaccaccgc caggtggacg    360
gttcagacgg acagtgccgg cgaggttgtg gccaaattcg tgattatggc caccgggtgt    420
ctgtcggagc cgaacgtgcc gtacataccg ggtgtggaga cattcgcggg cgacgtgctg    480
cacaccgggc gctggccgca ggatcccgtc gacttcacag gcaagcgggt cggcgtgatc    540
ggaaccggat catctggcgt gcaagccatc ccactcatcg cgcggaagc ggccgagctc    600
gtagtctttc agcgactcc tgcatacacg ttgcccgctg tcgacgagcc gctcgacccg    660
gaattgcagg cggcgatcaa ggccgattac aggggggttc gtgcgcgaaa caacgaagtg    720
cccaccgcgg gactctcccg atttccgacg aatccgaact cggttttcct gttctcaacg    780
aaggagcggg atgccatcct cgaacacaat tggaaaccgag gcggggccgtt gatgctgcgc    840
gccttcggcg atctgctggt ggactcagcc gctaacgagg tggtagccga gttcgtccgc    900
aacaagatcc gccagatcgt taccgacccc gaggtcgctg cgaagctcac accgacacac    960
gtgatcggat gaaaacgaat ctgtctcagc gacggctatt acgagacctc caaccgggtc   1020
aacgtgcgct tagtcgacat caaacgccac ccaatcgagg agatcacgcc tactacagcc   1080
cggaccggcg aggactcgca tgacctggac atgctcgtgt tcgccactgg ctacgatgcc   1140
atcactggcg cactctcacg catcgacatc cgcgcccgcg cagggttgtc attgcaggaa   1200
gcatggtcgg acggaccgcg cacctatctc gggtcggggg tctccggctt cccaaatctg   1260
ttcatcatga ccggccccgg aagcccatcg gtattgacca atgttcttgt cgccatacac   1320
caacatgcga catggatcgg cgaatgcctg aagcatatga ccgacaacga tattcggaca   1380
atggaagcca cgcccgaagc cgagcagaac tggggggacc acgtgcgcga cctcgccgag   1440
cagaccctgc tctcatcgtg cgggtcctgg tacctcggag caaacatccc cggtaagaga   1500
caagtattca tgccgctggt cgggtttccg gactacgcca agaaatgcgc ggaaatcgca   1560
tccgcgggct acccgggctt cgccttcag tacgaccccg tccctgtgaa ccagagctga   1620

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&lt;210&gt; 34

&lt;211&gt; 539

&lt;212&gt; PRT

&lt;213&gt; Rhodococcus erythropolis AN12

&lt;400&gt; 34

Met Thr Asp Glu Phe Asp Val Val Ile Val Gly Ala Gly Leu Ala Gly  
 1 5 10 15

Met Gln Met Leu His Glu Val Arg Met Val Gly Leu Thr Ala Lys Val  
 20 25 30

Phe Glu Ala Gly Gly Gly Ala Gly Gly Thr Trp Tyr Trp Asn Arg Tyr  
 35 40 45

Pro Gly Ala Arg Cys Asp Val Glu Ser Leu Glu Tyr Ser Tyr Gln Phe  
 50 55 60

Ser Glu Val Leu Gln Gln Glu Trp Glu Trp Thr Arg Arg Tyr Ala Asp  
 65 70 75 80

Gln Ala Glu Ile Met Arg Tyr Ile Ser His Val Val Glu Thr Phe Asp  
 85 90 95

Leu Ala Arg Asp Ile Arg Phe His Thr Arg Val Glu Ala Met Thr Tyr  
 100 105 110

Glu Glu Thr Thr Ala Arg Trp Thr Val Gln Thr Asp Ser Ala Gly Glu  
 115 120 125

Val Val Ala Lys Phe Val Ile Met Ala Thr Gly Cys Leu Ser Glu Pro  
 130 135 140

Asn Val Pro Tyr Ile Pro Gly Val Glu Thr Phe Ala Gly Asp Val Leu  
 145 150 155 160

His Thr Gly Arg Trp Pro Gln Asp Pro Val Asp Phe Thr Gly Lys Arg  
 165 170 175

Val Gly Val Ile Gly Thr Gly Ser Ser Gly Val Gln Ala Ile Pro Leu  
 180 185 190

Ile Ala Arg Gln Ala Ala Glu Leu Val Val Phe Gln Arg Thr Pro Ala  
 195 200 205

Tyr Thr Leu Pro Ala Val Asp Glu Pro Leu Asp Pro Glu Leu Gln Ala  
 210 215 220

Ala Ile Lys Ala Asp Tyr Arg Gly Phe Arg Ala Arg Asn Asn Glu Val



225		230		235		240
Pro Thr Ala Gly	Leu Ser Arg Phe	Pro Thr Asn	Pro Asn Ser	Val Phe		
	245	250		255		
Leu Phe Ser Thr	Lys Glu Arg Asp	Ala Ile Leu	Glu His Asn	Trp Asn		
	260	265		270		
Arg Gly Gly Pro	Leu Met Leu Arg	Ala Phe Gly	Asp Leu Leu	Val Asp		
	275	280		285		
Ser Ala Ala Asn	Glu Val Val Ala	Glu Phe Val	Arg Asn Lys	Ile Arg		
	290	295		300		
Gln Ile Val Thr	Asp Pro Glu Val	Ala Ala Lys	Leu Thr Pro	Thr His		
	305	310		315		320
Val Ile Gly Cys	Lys Arg Ile Cys	Leu Ser Asp	Gly Tyr Tyr	Glu Thr		
	325	330		335		
Tyr Asn Arg Val	Asn Val Arg Leu	Val Asp Ile	Lys Arg His	Pro Ile		
	340	345		350		
Glu Glu Ile Thr	Pro Thr Thr Ala	Arg Thr Gly	Glu Asp Ser	His Asp		
	355	360		365		
Leu Asp Met Leu	Val Phe Ala Thr	Gly Tyr Asp	Ala Ile Thr	Gly Ala		
	370	375		380		
Leu Ser Arg Ile	Asp Ile Arg Gly	Arg Ala Gly	Leu Ser Leu	Gln Glu		
	385	390		395		400
Ala Trp Ser Asp	Gly Pro Arg Thr	Tyr Leu Gly	Leu Gly Val	Ser Gly		
	405	410		415		
Phe Pro Asn Leu	Phe Ile Met Thr	Gly Pro Gly	Ser Pro Ser	Val Leu		
	420	425		430		
Thr Asn Val Leu	Val Ala Ile His	Gln His Ala	Thr Trp Ile	Gly Glu		
	435	440		445		
Cys Leu Lys His	Met Thr Asp Asn	Asp Ile Arg	Thr Met Glu	Ala Thr		
	450	455		460		
Pro Glu Ala Glu	Gln Asn Trp Gly	Asp His Val	Arg Asp Leu	Ala Glu		
	465	470		475		480



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cgggccatcg cggatcgtgt cggatcgtg acgatcttcc agcgctcacc acagtggatc 1020
gcaccgaacg acgactactt caccgaccatc gacgacggcg tccactggct gatggacaac 1080
atccccggct atcgcgagtg gtaccgggcg cgtctgtcgt ggatcttcaa cgacaaggtg 1140
tactcgtccc tccaggtcga ccccgactgg ccagagccga ggcctcgtat caatgcgacc 1200
aaccatgggtc atcgcaagtt ctacgaacgc tatctccgcg atcagctggg tgatcgaaca 1260
gatctgatcg aggcattctt tccggactat ccgccctttg gtaagcgaat gctgctggac 1320
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tccggccggt ccaccggaga aatctggggc gagcacgacg cgcgcgccta cctggggatc 1560
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tcggtcccga ctccgaccgc gcgaggggtga 1950

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&lt;210&gt; 36

&lt;211&gt; 649

&lt;212&gt; PRT

&lt;213&gt; Rhodococcus erythropolis AN12

&lt;400&gt; 36

```

Met Thr Ile Val Thr Asp Leu Asp Arg Asp His Leu Arg Ser Ala Val
1           5           10           15

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```

Leu Arg Gly Asn Val Pro Thr Met Leu Ala Val Leu Leu Glu Leu Thr
20           25           30

```

```

Ala Asp Glu Arg Trp Val Ala Pro Arg Tyr Gln Pro Thr Arg Ser Arg
35           40           45

```

```

Gly Met Asp Asp Asn Ser Thr Gly Gly Leu Pro Glu Glu Val Gln Ser
50           55           60

```

Glu Ile Arg Ser Ala Leu Ile Asp Ala Val Glu Arg Trp Trp Thr Leu  
 65 70 75 80

Asp Glu Pro Ser Arg Arg Thr Leu Asp Ser Ser Glu Val Glu Arg Ile  
 85 90 95

Leu Asn Phe Thr Cys Ser Glu Thr Val Pro Pro Asp Phe Ala Pro Met  
 100 105 110

Met Ala Glu Ile Val Asn Gly Pro Gln Ile Lys Pro Ala Thr Ala Lys  
 115 120 125

Cys Asp Glu Arg Leu His Ala Ile Val Ile Gly Ala Gly Ile Ala Gly  
 130 135 140

Met Leu Ala Ser Val Glu Leu Ser Arg Ala Gly Ile Pro His Val Ile  
 145 150 155 160

Leu Glu Lys Asn Asp Asp Val Gly Gly Ser Trp Trp Glu Asn Arg Tyr  
 165 170 175

Pro Gly Ala Gly Val Asp Thr Pro Ser His Leu Tyr Ser Ile Ser Ser  
 180 185 190

Phe Pro Arg Asn Trp Ser Thr His Phe Gly Lys Arg Asp Glu Val Gln  
 195 200 205

Gly Tyr Leu Glu Asp Phe Ala Glu Ala Asn Asp Ile Arg Arg Asn Val  
 210 215 220

Arg Phe Arg His Glu Val Thr Arg Ala Glu Phe Glu Glu Ser Lys Gln  
 225 230 235 240

Ser Trp Arg Val Ser Val Gln Arg Pro Gly Glu Ala Ser Glu Thr Leu  
 245 250 255

Glu Ala Pro Ile Leu Ile Ser Ala Val Gly Leu Leu Asn Arg Pro Lys  
 260 265 270

Ile Pro His Leu Pro Gly Ile Glu Thr Phe Arg Gly Arg Leu Phe His  
 275 280 285

Ser Ala Glu Trp Pro Ser Glu Leu Asp Asp Pro Glu Ser Leu Arg Gly  
 290 295 300

Lys Arg Val Gly Ile Val Gly Thr Gly Ala Ser Ala Met Gln Ile Gly  
 305 310 315 320

Pro Ala Ile Ala Asp Arg Val Gly Ser Leu Thr Ile Phe Gln Arg Ser  
 325 330 335

Pro Gln Trp Ile Ala Pro Asn Asp Asp Tyr Phe Thr Thr Ile Asp Asp  
 340 345 350

Gly Val His Trp Leu Met Asp Asn Ile Pro Gly Tyr Arg Glu Trp Tyr  
 355 360 365

Arg Ala Arg Leu Ser Trp Ile Phe Asn Asp Lys Val Tyr Ser Ser Leu  
 370 375 380

Gln Val Asp Pro Asp Trp Pro Glu Pro Ser Ala Ser Ile Asn Ala Thr  
 385 390 395 400

Asn His Gly His Arg Lys Phe Tyr Glu Arg Tyr Leu Arg Asp Gln Leu  
 405 410 415

Gly Asp Arg Thr Asp Leu Ile Glu Ala Ser Leu Pro Asp Tyr Pro Pro  
 420 425 430

Phe Gly Lys Arg Met Leu Leu Asp Asn Gly Trp Phe Thr Met Leu Arg  
 435 440 445

Lys Pro Asp Val Thr Leu Val Pro His Gly Val Asp Ala Leu Thr Pro  
 450 455 460

Ser Gly Leu Val Asp Thr Asn Gly Val Glu His Gln Leu Asp Val Ile  
 465 470 475 480

Val Met Ala Thr Gly Phe His Ser Val Arg Val Leu Tyr Pro Met Asp  
 485 490 495

Ile Val Gly Arg Ser Gly Arg Ser Thr Gly Glu Ile Trp Gly Glu His  
 500 505 510

Asp Ala Arg Ala Tyr Leu Gly Ile Thr Val Pro Asp Phe Pro Asn Phe  
 515 520 525

Phe Val Met Thr Gly Pro Asn Thr Gly Leu Gly His Gly Gly Ser Phe  
 530 535 540

Ile Thr Ile Leu Glu Cys Gln Val Arg Tyr Ile Met Asp Ala Leu Lys  
 545 550 555 560

Leu Met Gln Ser Glu Asn Leu Gly Ala Met Glu Cys Arg Ala Glu Val  
 565 570 575

Asn Asp Arg Tyr Asn Glu Ala Val Asp Arg Gln His Ala Gln Met Val  
 580 585 590

Trp Thr His Pro Ala Met Glu Asn Trp Tyr Arg Asn Pro Asp Gly Arg  
 595 600 605

Val Val Ser Val Leu Pro Trp Arg Ile Asn Asp Tyr Trp Ala Met Thr  
 610 615 620

Tyr Arg Val Asp Pro Ser Asp Phe Arg Thr Glu Pro Ala Arg Ser Glu  
 625 630 635 640

Ser Val Pro Thr Pro Thr Ala Arg Gly  
 645

<210> 37

<211> 1485

<212> DNA

<213> Rhodococcus erythropolis AN12

<400> 37  
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 gctgatatcg gtggcacctg gcgagacaac acctaccag gttgtgcctg tgacgtgccg 180  
 acggcgctgt actcgattc ttttgcgccg agcgctgatt ggagtcatac ctttgctcgt 240  
 cagcccgaga tctacgacta tctgaagaaa gtggccgcag acaccggcat cggggatcgc 300  
 gtaatcctga actgcgaact cgaagccgct gtgtgggacg aggatgcggc gctgtggcgg 360  
 gtccggacat ccctggggtc gttgacagtc aaagcgctgg tcgctgcgac cggggcgttg 420  
 tcgacacca agatcccga ttttcccgt ctcgaccaat tctccggtac cactttccat 480  
 tcggcgacgt ggaaccacga acacgaactg cgtgggtgagc gcgtagccgt gatcggaacg 540  
 ggagcgctcg cggttcagtt cgttcccga attgccgacc ctgctgcca tgtcaccgtg 600  
 ttccagagaa ctccggcctg ggtgattccg cgaatggatc gcaccctgcc tgcggcgag 660

```

aaggccgtct actcgcggat tcccgtacg cagaaagttg ttcgcggagc ggtttacggt      720
tttcgcgagt tgctcgggtc cgcgatgtca catgcgacgt gggtcctgcc ggccctcgag      780
gcggccgcgc gcctccatct gcgcagacag gtgaaagatc cggagttgcg ccggaaactg      840
actcccgatt tcacgatcgg ttgcaagcgc atgcttctgt ccaacgactg gttgcgcacc      900
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gtggtcgacg ggcacggagt cgagcacaag gtcgacacca tcattcttcgc cacgggggttc    1020
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aactcggtag agtggccgac gttcacattc aaattccggt cgctgctgga gcatttcgat    1440
cgtgagaact actccgctcg caagatcgaa agcgtccagg catga                        1485

```

<210> 38

<211> 494

<212> PRT

<213> Rhodococcus erythropolis AN12

<400> 38

```

Val Lys Leu Pro Glu His Val Glu Thr Leu Ile Val Gly Ala Gly Phe
1              5              10              15

```

```

Ala Gly Met Gly Leu Ala Ala Arg Met Leu Arg Asp Asn Arg Thr Ala
20              25              30

```

```

Asp Val Val Leu Ile Glu Arg Gly Ala Asp Ile Gly Gly Thr Trp Arg
35              40              45

```

```

Asp Asn Thr Tyr Pro Gly Cys Ala Cys Asp Val Pro Thr Ala Leu Tyr
50              55              60

```

```

Ser Tyr Ser Phe Ala Pro Ser Ala Asp Trp Ser His Thr Phe Ala Arg
65              70              75              80

```

Gln Pro Glu Ile Tyr Asp Tyr Leu Lys Lys Val Ala Ala Asp Thr Gly  
                             85                            90                            95

Ile Gly Asp Arg Val Ile Leu Asn Cys Glu Leu Glu Ala Ala Val Trp  
                             100                            105                            110

Asp Glu Asp Ala Ala Leu Trp Arg Val Arg Thr Ser Leu Gly Ser Leu  
                             115                            120                            125

Thr Val Lys Ala Leu Val Ala Ala Thr Gly Ala Leu Ser Thr Pro Lys  
                             130                            135                            140

Ile Pro Asp Phe Pro Gly Leu Asp Gln Phe Ser Gly Thr Thr Phe His  
 145                            150                            155                            160

Ser Ala Thr Trp Asn His Glu His Glu Leu Arg Gly Glu Arg Val Ala  
                             165                            170                            175

Val Ile Gly Thr Gly Ala Ser Ala Val Gln Phe Val Pro Glu Ile Ala  
                             180                            185                            190

Asp Pro Ala Ala His Val Thr Val Phe Gln Arg Thr Pro Ala Trp Val  
                             195                            200                            205

Ile Pro Arg Met Asp Arg Thr Leu Pro Ala Ala Gln Lys Ala Val Tyr  
                             210                            215                            220

Ser Arg Ile Pro Ala Thr Gln Lys Val Val Arg Gly Ala Val Tyr Gly  
 225                            230                            235                            240

Phe Arg Glu Leu Leu Gly Ala Ala Met Ser His Ala Thr Trp Val Leu  
                             245                            250                            255

Pro Ala Phe Glu Ala Ala Ala Arg Leu His Leu Arg Arg Gln Val Lys  
                             260                            265                            270

Asp Pro Glu Leu Arg Arg Lys Leu Thr Pro Asp Phe Thr Ile Gly Cys  
                             275                            280                            285

Lys Arg Met Leu Leu Ser Asn Asp Trp Leu Arg Thr Leu Asp Arg Ala  
                             290                            295                            300

Asp Val Ser Leu Val Asp Ser Gly Leu Val Ser Val Thr Glu Gly Gly  
 305                            310                            315                            320

Val Val Asp Gly His Gly Val Glu His Lys Val Asp Thr Ile Ile Phe



					325					330						335
Ala	Thr	Gly	Phe	Thr	Pro	Thr	Glu	Pro	Pro	Val	Ala	His	Leu	Ile	Thr	
			340					345					350			
Gly	Lys	Arg	Gly	Glu	Thr	Leu	Ala	Ala	His	Trp	Asn	Gly	Ser	Pro	Asn	
		355					360					365				
Ala	Tyr	Lys	Gly	Thr	Ala	Val	Ser	Gly	Phe	Pro	Asn	Leu	Phe	Leu	Met	
	370					375					380					
Tyr	Gly	Pro	Asn	Thr	Asn	Leu	Gly	His	Ser	Ser	Ile	Val	Tyr	Met	Leu	
385					390					395						400
Glu	Ser	Gln	Ala	Glu	Tyr	Val	Asn	Asp	Ala	Leu	Asn	Thr	Met	Lys	Arg	
				405					410					415		
Glu	Arg	Leu	Asp	Ala	Leu	Asp	Val	Asn	Glu	Ser	Val	Gln	Val	His	Tyr	
			420					425					430			
Asn	Lys	Gly	Ile	Gln	His	Glu	Leu	Gln	His	Thr	Val	Trp	Asn	Lys	Gly	
		435					440					445				
Gly	Cys	Ser	Ser	Trp	Tyr	Ile	Asp	Pro	Glu	Gly	Arg	Asn	Ser	Val	Gln	
	450					455					460					
Trp	Pro	Thr	Phe	Thr	Phe	Lys	Phe	Arg	Ser	Leu	Leu	Glu	His	Phe	Asp	
465					470					475					480	
Arg	Glu	Asn	Tyr	Ser	Ala	Arg	Lys	Ile	Glu	Ser	Val	Gln	Ala			
				485					490							

<210> 39

<211> 1500

**<212> DNA**

<213> Rhodococcus erythropolis AN12

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<400> 39
atgacacagc atgtcgacgt actgatcatc ggcgctggct tgtccggaat cggcgcggtc 60
tgccacctca ttcgtgagca gaccggaagc acttacgcga tcctcgagcg ccgcgagaac 120
atcgggtggca cctgggacct gttcaagta cccggcatcc gttcggactc cgacatgctc 180
accttcggat tcggtttccg tccttggaac ggcaccaaag tgctcgcaga cggcgccagt 240
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atccgtgact acgtcgagga aaccgccaag gaatacggcg tcaccgacca catcaacttc 300
ggccgcaagg tcgtggctat ggacttcgac cgtaccgccg cgcagtggtc cgtgaccgtc 360
ctggtcgagg cgacagggga gaccgagacg tggaccgca acgtcctcgt cggcgccctgt 420
ggttactaca actacgacaa gggttaccgc cccgccttcc ccggtgagga cgacttcgcg 480
ggtcagatcg tgcacccgca gcactggccg gaggatctcg attacaccgg aaagaaggta 540
gtggtcatcg gttccggcgc caccgcgatc acgctgatcc cgtcgatggc cccaccgcc 600
ggtcacgtca ccatgctgca gcgctcgccc acgtggatcc aggcgcttcc gtccgaggac 660
cctgttgcca aggtctctaa gctcgcacgc gttcccgacc agattgctta caagattggt 720
cgagcccgca atatcgcaact gcaacgcgcc agctttcagc tttctcgca caaccggaag 780
ctggccaaga agctgttcct cgcccagatc cgcctgcagc tcggcaagaa cgtggacctg 840
cgtcaattca ctcccagcta caaccgtgg gatcagcgcc tgtgctgggt tcccaacggg 900
gacctgttca aggtgctcaa gagcggcaag gccgacatcg tcaccgaccg tatcgccacg 960
ttcaccgaga agggcatcgt gaccgagtcg ggccgcgaaa tcgaggccga cgtcatcgtc 1020
acggcgaccg gcttgaacgt acagattctg ggccgcgcaa ccatgagcat cgacggcgag 1080
ccggtcaagc tcaacgagac tgtggcctac aagagcgtgc tctactccga catcccgaac 1140
ttcctgatga tcctcggtta caccaacgcg tcgtggacgc tcaaggetga cctggccgcg 1200
tcctatctgt gtcgctgct caagatcatg cgcgatcgca gctacacgac tttcgaggtt 1260
cacgccgaac ccgaggactt cgccgaagaa tctctcatgg gcggagccct gacctcgggc 1320
tacatccagc gcggcgacgg agaaatgccg cgtcagggtg cccgcggcgc gtggaaagtg 1380
gtcaacaatt actaccgca ccgcaagctg atgcacgac ccgagatcga agacgggtgtg 1440
ctgcagttca gcaaggctga tattgctgtc gtgcctgata gcaaggctgc cagcgcatag 1500

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<210> 40

<211> 499

<212> PRT

<213> Rhodococcus erythropolis AN12

<400> 40

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Met Thr Gln His Val Asp Val Leu Ile Ile Gly Ala Gly Leu Ser Gly
1           5           10          15

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Ile Gly Ala Ala Cys His Leu Ile Arg Glu Gln Thr Gly Ser Thr Tyr

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20	25	30
Ala Ile Leu Glu Arg Arg Glu Asn Ile Gly Gly Thr Trp Asp Leu Phe		
35	40	45
Lys Tyr Pro Gly Ile Arg Ser Asp Ser Asp Met Leu Thr Phe Gly Phe		
50	55	60
Gly Phe Arg Pro Trp Ile Gly Thr Lys Val Leu Ala Asp Gly Ala Ser		
65	70	75
Ile Arg Asp Tyr Val Glu Glu Thr Ala Lys Glu Tyr Gly Val Thr Asp		
85	90	95
His Ile Asn Phe Gly Arg Lys Val Val Ala Met Asp Phe Asp Arg Thr		
100	105	110
Ala Ala Gln Trp Ser Val Thr Val Leu Val Glu Ala Thr Gly Glu Thr		
115	120	125
Glu Thr Trp Thr Ala Asn Val Leu Val Gly Ala Cys Gly Tyr Tyr Asn		
130	135	140
Tyr Asp Lys Gly Tyr Arg Pro Ala Phe Pro Gly Glu Asp Asp Phe Arg		
145	150	155
Gly Gln Ile Val His Pro Gln His Trp Pro Glu Asp Leu Asp Tyr Thr		
165	170	175
Gly Lys Lys Val Val Val Ile Gly Ser Gly Ala Thr Ala Ile Thr Leu		
180	185	190
Ile Pro Ser Met Ala Pro Thr Ala Gly His Val Thr Met Leu Gln Arg		
195	200	205
Ser Pro Thr Trp Ile Gln Ala Leu Pro Ser Glu Asp Pro Val Ala Lys		
210	215	220
Gly Leu Lys Leu Ala Arg Val Pro Asp Gln Ile Ala Tyr Lys Ile Gly		
225	230	235
Arg Ala Arg Asn Ile Ala Leu Gln Arg Ala Ser Phe Gln Leu Ser Arg		
245	250	255
Thr Asn Pro Lys Leu Ala Lys Lys Leu Phe Leu Ala Gln Ile Arg Leu		
260	265	270

Gln Leu Gly Lys Asn Val Asp Leu Arg His Phe Thr Pro Ser Tyr Asn  
 275 280 285

Pro Trp Asp Gln Arg Leu Cys Val Val Pro Asn Gly Asp Leu Phe Lys  
 290 295 300

Val Leu Lys Ser Gly Lys Ala Asp Ile Val Thr Asp Arg Ile Ala Thr  
 305 310 315 320

Phe Thr Glu Lys Gly Ile Val Thr Glu Ser Gly Arg Glu Ile Glu Ala  
 325 330 335

Asp Val Ile Val Thr Ala Thr Gly Leu Asn Val Gln Ile Leu Gly Gly  
 340 345 350

Ala Thr Met Ser Ile Asp Gly Glu Pro Val Lys Leu Asn Glu Thr Val  
 355 360 365

Ala Tyr Lys Ser Val Leu Tyr Ser Asp Ile Pro Asn Phe Leu Met Ile  
 370 375 380

Leu Gly Tyr Thr Asn Ala Ser Trp Thr Leu Lys Ala Asp Leu Ala Ala  
 385 390 395 400

Ser Tyr Leu Cys Arg Val Leu Lys Ile Met Arg Asp Arg Ser Tyr Thr  
 405 410 415

Thr Phe Glu Val His Ala Glu Pro Glu Asp Phe Ala Glu Glu Ser Leu  
 420 425 430

Met Gly Gly Ala Leu Thr Ser Gly Tyr Ile Gln Arg Gly Asp Gly Glu  
 435 440 445

Met Pro Arg Gln Gly Ala Arg Gly Ala Trp Lys Val Val Asn Asn Tyr  
 450 455 460

Tyr Arg Asp Arg Lys Leu Met His Asp Ala Glu Ile Glu Asp Gly Val  
 465 470 475 480

Leu Gln Phe Ser Lys Val Asp Ile Ala Val Val Pro Asp Ser Lys Val  
 485 490 495

Ala Ser Ala

&lt;210&gt; 41

&lt;211&gt; 1482

&lt;212&gt; DNA

<213> *Rhodococcus erythropolis* AN12

&lt;400&gt; 41

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atgtcatcac ggggtcaacga cggccacatc gcgatcatcg gaaccgggtt ttccgggctg      60
tgcattggcga tcgaactgaa gaagaagggc atcgacgact tcgtcctgta cgaacgcgcc      120
gacgatgtcg gcggaacctg gcgcgacaac acatacccag gggcagcctg cgatgtgccc      180
agcgtgttgt attcctactc cttcgctcag aaccggaact ggaccggtat cttcccgccca      240
tggtcggaaac tgctcgacta tctcagatct gttgctgcgc agtatgattt gctgccgcac      300
atccgcttcg gtgtcgaggt ctccgaaatg cgggtcgacg aggaccggct ccggtggaac      360
atccagttcg catccggcga atcagtgcgc gcggccggtt tcgtcaacgg ctcagggggc      420
ttgagtaatc cgtacatccc gcagctaccc ggactggaat cattcgaggg tgccgcattc      480
cactccgccca agtggcgaca tgacctgcac atgtcgggaa ggcgtgtcgc ggtgataggt      540
tccggcgcca gtgcgatcca gttcgtcccc gaaatcgccc cgcacaccga gacccttcac      600
gtgtttcagc gatcacccaa ctgggtcatg ccacgtggtg atgccgcgct gtcgcccgcc      660
acccgcgaaa gattctcacg gcgtccttat cgtcaacggt ggctgcgatg gcggacctac      720
tgggcattcg aaaagctcgc cagcgccttc ctcggaaatc gaaaactcgt cgaacagtac      780
cgatcccagg cgctcgccaa tcttcaacag caagtgcggg attcggactt gaggcagaag      840
gtcaccaccag attacgatcc tggctgtaaa cgctcgcttg tatccgacga ctggtacccc      900
gcgctgcaac gggaaaatgt gcacttgaac acctcggggg tttccgagat ccgcccgcat      960
tcgatcattg actcagaggg agcggaaacac gaagtcgaca ccctgatctt cgcgaccgga     1020
ttccaggcaa ccagcttcct ggcaccgatg aaagtattcg gccgcgaagg agtcgaactc     1080
tccgacagtt ggcgcgaggg cgcgcgaaca aagctcgggc ttgcatccgc cgcgttcccc     1140
aacctgtggt tcctcaacgg cccgaatacc ggtctcggtc acaactcgat catcttcacg     1200
atcgaagcac aagccagata catcgcttcg gcagtgcagt acatgcgccg aaaaagtatc     1260
actgccctcg aactcgatcg caccgtccag acaggcagct acgccgccac ccaagaacgc     1320
atgcgccgaa ctgtatgggc atcgggtggc tgcgacagct ggtatcaatc cgctgacggg     1380
cgaatcgaca ccctgtggcc ggccagcaca atcgaatact ggttgcgcac caggctattc     1440
cgcaagtccg acttccatgc actgacgaca ggcaaaggat ga                        1482

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&lt;210&gt; 42

&lt;211&gt; 493

&lt;212&gt; PRT

&lt;213&gt; Rhodococcus erythropolis AN12

&lt;400&gt; 42

Met Ser Ser Arg Val Asn Asp Gly His Ile Ala Ile Ile Gly Thr Gly  
 1 5 10 15

Phe Ser Gly Leu Cys Met Ala Ile Glu Leu Lys Lys Lys Gly Ile Asp  
 20 25 30

Asp Phe Val Leu Tyr Glu Arg Ala Asp Asp Val Gly Gly Thr Trp Arg  
 35 40 45

Asp Asn Thr Tyr Pro Gly Ala Ala Cys Asp Val Pro Ser Val Leu Tyr  
 50 55 60

Ser Tyr Ser Phe Ala Gln Asn Pro Asn Trp Thr Arg Ile Phe Pro Pro  
 65 70 75 80

Trp Ser Glu Leu Leu Asp Tyr Leu Arg Ser Val Ala Ala Gln Tyr Asp  
 85 90 95

Leu Leu Pro His Ile Arg Phe Gly Val Glu Val Ser Glu Met Arg Phe  
 100 105 110

Asp Glu Asp Arg Leu Arg Trp Asn Ile Gln Phe Ala Ser Gly Glu Ser  
 115 120 125

Val Thr Ala Ala Val Val Val Asn Gly Ser Gly Gly Leu Ser Asn Pro  
 130 135 140

Tyr Ile Pro Gln Leu Pro Gly Leu Glu Ser Phe Glu Gly Ala Ala Phe  
 145 150 155 160

His Ser Ala Lys Trp Arg His Asp Leu Asp Met Ser Gly Arg Arg Val  
 165 170 175

Ala Val Ile Gly Ser Gly Ala Ser Ala Ile Gln Phe Val Pro Glu Ile  
 180 185 190

Ala Pro His Thr Glu Thr Leu His Val Phe Gln Arg Ser Pro Asn Trp  
 195 200 205

Val Met Pro Arg Gly Asp Ala Ala Leu Ser Pro Ala Thr Arg Glu Arg  
 210 215 220

Phe Ser Arg Arg Pro Tyr Arg Gln Arg Trp Leu Arg Trp Arg Thr Tyr  
 225 230 235 240

Trp Ala Phe Glu Lys Leu Ala Ser Ala Phe Leu Gly Asn Arg Lys Leu  
 245 250 255

Val Glu Gln Tyr Arg Ser Gln Ala Leu Ala Asn Leu Gln Gln Gln Val  
 260 265 270

Pro Asp Ser Asp Leu Arg Gln Lys Val Thr Pro Asp Tyr Asp Pro Gly  
 275 280 285

Cys Lys Arg Arg Leu Ile Ser Asp Asp Trp Tyr Pro Ala Leu Gln Arg  
 290 295 300

Glu Asn Val His Leu Asn Thr Ser Gly Val Ser Glu Ile Arg Pro His  
 305 310 315 320

Ser Ile Ile Asp Ser Glu Gly Ala Glu His Glu Val Asp Thr Leu Ile  
 325 330 335

Phe Ala Thr Gly Phe Gln Ala Thr Ser Phe Leu Ala Pro Met Lys Val  
 340 345 350

Phe Gly Arg Glu Gly Val Glu Leu Ser Asp Ser Trp Arg Glu Gly Ala  
 355 360 365

Ala Thr Lys Leu Gly Leu Ala Ser Ala Ala Phe Pro Asn Leu Trp Phe  
 370 375 380

Leu Asn Gly Pro Asn Thr Gly Leu Gly His Asn Ser Ile Ile Phe Met  
 385 390 395 400

Ile Glu Ala Gln Ala Arg Tyr Ile Ala Ser Ala Val Gln Tyr Met Arg  
 405 410 415

Arg Lys Ser Ile Thr Ala Leu Glu Leu Asp Arg Thr Val Gln Thr Gly  
 420 425 430

Ser Tyr Ala Ala Thr Gln Glu Arg Met Arg Arg Thr Val Trp Ala Ser  
 435 440 445

Gly Gly Cys Asp Ser Trp Tyr Gln Ser Ala Asp Gly Arg Ile Asp Thr  
 450 455 460

Leu Trp Pro Ala Ser Thr Ile Glu Tyr Trp Leu Arg Thr Arg Leu Phe  
 465 470 475 480

Arg Lys Ser Asp Phe His Ala Leu Thr Thr Gly Lys Gly  
 485 490

<210> 43

<211> 1626

<212> DNA

<213> Rhodococcus erythropolis AN12

<400> 43

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gggatctacg ccgtccacaa actggccaac gagctcggcc tcacgacggg cggcttcgac     120
aaggcagacg gcccgggcgg cacgtggtac tggaaccgct acccgggtgc actgtccgac     180
accgaaagcc acgtctaccg gttctcattc gaccgtgacc tgcttcagga cggtagctgg     240
aagcacacct acaccactca acccgagatt ctccaatacc ttgaggatgt cgtttcccg     300
ttcgacctac gccggcactt ccacttcggc actgccgtcg aatctgcggg gtatctcgaa     360
gacgaacaac tgtgggaagt caccaccgac acaggcgaga tctaccgcgc tacctacgtc     420
gtcaatgctg tcgggctcct ctccgccatc aatcgaccgg atctgcccgg tctcgagaca     480
ttcgaaggcg agaccatcca caccgcagcg tggcccaggg gcaaggatct caccggccgc     540
cgcgtcggcg tgatcggtag cggatctact gggcaacagg tcatcacggc cctggcgcca     600
acggtcgaac acctcactgt attcgtgcga actcccagc actcgggtgc ggtcggcaag     660
cgcgcggtga ccgacgagca gatcgacgca gtcaaagccg actacgagaa catctggact     720
cagggtcaaaa gatcctcggg ggcattcggc ttcgaggaat ctactgttcc ggccatgagc     780
gtgtccgcgg aagaacgcct cagggctctac gaagaggcat gggagcaggg cggcggtttc     840
cgattcatgt tcggaacctt cggtgacatc gctaccgacg aagaagccaa cgaaactgca     900
gcacgttca ttcgctcgaa gatcaccgcc atgatcgaag acccggagac tgcccgcaaa     960
ctgacgcca ccggactatt cgcgagacga ccgttgtgcg acgacgggta cttccaggtc    1020

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ttcaaccgcc cgaacgtcga ggcggtcgcc atcaaggaaa accccattcg tgagatcaca 1080
gccaagggcg tggtagaccga ggacggcgtc ctgcacaaat tggacgtcct ggtcctcgcc 1140
accggcttcg acgccgtcga cgggaactac cgccgcatga ccatttcggg tcgcggtggc 1200
ctgaacatca acgaccattg ggacggccaa cccaccagct acctggggat tgccaccgcg 1260
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ggaaagaagc ccagcgtgct gttctacctt ggcgggctcg gcaactaccg cgccgtcctg 1560
gcagacgtca ccgagggggg ctatcagggc tttgctctga agacggccga caccgtcgac 1620
gcctga 1626

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&lt;210&gt; 44

&lt;211&gt; 541

&lt;212&gt; PRT

&lt;213&gt; Rhodococcus erythropolis AN12

&lt;400&gt; 44

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Met Thr Thr Gln Lys Ala Leu Thr Thr Val Asp Ala Ile Val Ile Gly
1          5          10          15

Ala Gly Phe Gly Gly Ile Tyr Ala Val His Lys Leu Ala Asn Glu Leu
          20          25          30

Gly Leu Thr Thr Val Gly Phe Asp Lys Ala Asp Gly Pro Gly Gly Thr
          35          40          45

Trp Tyr Trp Asn Arg Tyr Pro Gly Ala Leu Ser Asp Thr Glu Ser His
          50          55          60

Val Tyr Arg Phe Ser Phe Asp Arg Asp Leu Leu Gln Asp Gly Thr Trp
65          70          75          80

Lys His Thr Tyr Thr Thr Gln Pro Glu Ile Leu Glu Tyr Leu Glu Asp
          85          90          95

Val Val Ser Arg Phe Asp Leu Arg Arg His Phe His Phe Gly Thr Ala
          100          105          110

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Val Glu Ser Ala Val Tyr Leu Glu Asp Glu Gln Leu Trp Glu Val Thr  
 115 120 125

Thr Asp Thr Gly Glu Ile Tyr Arg Ala Thr Tyr Val Val Asn Ala Val  
 130 135 140

Gly Leu Leu Ser Ala Ile Asn Arg Pro Asp Leu Pro Gly Leu Glu Thr  
 145 150 155 160

Phe Glu Gly Glu Thr Ile His Thr Ala Ala Trp Pro Glu Gly Lys Asp  
 165 170 175

Leu Thr Gly Arg Arg Val Gly Val Ile Gly Thr Gly Ser Thr Gly Gln  
 180 185 190

Gln Val Ile Thr Ala Leu Ala Pro Thr Val Glu His Leu Thr Val Phe  
 195 200 205

Val Arg Thr Pro Gln Tyr Ser Val Pro Val Gly Lys Arg Ala Val Thr  
 210 215 220

Asp Glu Gln Ile Asp Ala Val Lys Ala Asp Tyr Glu Asn Ile Trp Thr  
 225 230 235 240

Gln Val Lys Arg Ser Ser Val Ala Phe Gly Phe Glu Glu Ser Thr Val  
 245 250 255

Pro Ala Met Ser Val Ser Ala Glu Glu Arg Leu Arg Val Tyr Glu Glu  
 260 265 270

Ala Trp Glu Gln Gly Gly Gly Phe Arg Phe Met Phe Gly Thr Phe Gly  
 275 280 285

Asp Ile Ala Thr Asp Glu Glu Ala Asn Glu Thr Ala Ala Ser Phe Ile  
 290 295 300

Arg Ser Lys Ile Thr Ala Met Ile Glu Asp Pro Glu Thr Ala Arg Lys  
 305 310 315 320

Leu Thr Pro Thr Gly Leu Phe Ala Arg Arg Pro Leu Cys Asp Asp Gly  
 325 330 335

Tyr Phe Gln Val Phe Asn Arg Pro Asn Val Glu Ala Val Ala Ile Lys  
 340 345 350

Glu Asn Pro Ile Arg Glu Ile Thr Ala Lys Gly Val Val Thr Glu Asp  
 355 360 365

Gly Val Leu His Lys Leu Asp Val Leu Val Leu Ala Thr Gly Phe Asp  
 370 375 380

Ala Val Asp Gly Asn Tyr Arg Arg Met Thr Ile Ser Gly Arg Gly Gly  
 385 390 395 400

Leu Asn Ile Asn Asp His Trp Asp Gly Gln Pro Thr Ser Tyr Leu Gly  
 405 410 415

Ile Ala Thr Ala Asn Phe Pro Asn Trp Phe Met Val Leu Gly Pro Asn  
 420 425 430

Gly Pro Phe Thr Asn Leu Pro Pro Ser Ile Glu Thr Gln Val Glu Trp  
 435 440 445

Ile Ser Asp Thr Ile Gly Tyr Val Glu Arg Thr Gly Val Arg Ala Ile  
 450 455 460

Glu Pro Thr Pro Glu Ala Glu Ser Ala Trp Thr Ala Thr Cys Thr Asp  
 465 470 475 480

Ile Ala Asn Met Thr Val Phe Thr Lys Val Asp Ser Trp Ile Phe Gly  
 485 490 495

Ala Asn Val Pro Gly Lys Lys Pro Ser Val Leu Phe Tyr Leu Gly Gly  
 500 505 510

Leu Gly Asn Tyr Arg Ala Val Leu Ala Asp Val Thr Glu Gly Gly Tyr  
 515 520 525

Gln Gly Phe Ala Leu Lys Thr Ala Asp Thr Val Asp Ala  
 530 535 540

<210> 45

<211> 1638

<212> DNA

<213> Rhodococcus erythropolis AN12

<400> 45

atgacaacta ccgaatccag aactcagacc gacaaggctg gggccgtcac gctcgatgcg 60

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ctgaacgtcc ggccttacga cgctgcggaa gacgtcggcg gtacgtggta ctggaaccgt 180
taccaggcg cacgattcga ctccgaagcc tacatctacc agtacctgtt ctccgaggac 240
ctgtacaaga actggagctg gagtcaacgc ttcccgccc agcccgaaat tgagcgggtg 300
atgcgctacg tcgccgacac cctggacctg cgtcgcagca ttcagttttc cacaacaatc 360
accagcgccg agttcgacga ggtagctgag cgttggacca ttcgcaccga ccgcggcgag 420
gaaatcagca cccgattett catcacctgt tgcggaatgc tgtcggcgcc gatggaagat 480
ttgttccccg gacaacagga cttccggggg cagatcttcc acacctcgcg atggccgcac 540
ggagatgtag aactcaccgg taagcgtgtc ggtgtcgtcg gcgtcggcgc cactggcatt 600
caggtaatcc agaccatcgc cgacgaggtt gatcaactga aggtgttcgt gcggacaccc 660
cagtacgcct tgccgatgaa aaacctcag tacgacagcg acgacgtcgc ggcctacaag 720
gaccgattcg aggagcttcg aaccacactg ccgcacacct tcacaggctt cgaatacgat 780
ttcgaatacg tgtgggcccga cctagcccc gaacagcgcc gcgaggtgct cgagaacatc 840
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&lt;210&gt; 46

&lt;211&gt; 545

&lt;212&gt; PRT

&lt;213&gt; Rhodococcus erythropolis AN12

&lt;400&gt; 46

Met Thr Thr Thr Glu Ser Arg Thr Gln Thr Asp Lys Ala Gly Ala Val  
 1 5 10 15

Thr Leu Asp Ala Leu Ile Ile Gly Ala Gly Val Ala Gly Leu Tyr Gln  
 20 25 30

Leu His Met Leu Arg Glu Gln Gly Leu Asn Val Arg Ala Tyr Asp Ala  
 35 40 45

Ala Glu Asp Val Gly Gly Thr Trp Tyr Trp Asn Arg Tyr Pro Gly Ala  
 50 55 60

Arg Phe Asp Ser Glu Ala Tyr Ile Tyr Gln Tyr Leu Phe Ser Glu Asp  
 65 70 75 80

Leu Tyr Lys Asn Trp Ser Trp Ser Gln Arg Phe Pro Ala Gln Pro Glu  
 85 90 95

Ile Glu Arg Trp Met Arg Tyr Val Ala Asp Thr Leu Asp Leu Arg Arg  
 100 105 110

Ser Ile Gln Phe Ser Thr Thr Ile Thr Ser Ala Glu Phe Asp Glu Val  
 115 120 125

Ala Glu Arg Trp Thr Ile Arg Thr Asp Arg Gly Glu Glu Ile Ser Thr  
 130 135 140

Arg Phe Phe Ile Thr Cys Cys Gly Met Leu Ser Ala Pro Met Glu Asp  
 145 150 155 160

Leu Phe Pro Gly Gln Gln Asp Phe Arg Gly Gln Ile Phe His Thr Ser  
 165 170 175

Arg Trp Pro His Gly Asp Val Glu Leu Thr Gly Lys Arg Val Gly Val  
 180 185 190

Val Gly Val Gly Ala Thr Gly Ile Gln Val Ile Gln Thr Ile Ala Asp  
 195 200 205

Glu Val Asp Gln Leu Lys Val Phe Val Arg Thr Pro Gln Tyr Ala Leu

210	215	220
Pro Met Lys Asn Pro Gln Tyr Asp Ser Asp Asp Val Ala Ala Tyr Lys		
225	230	235 240
Asp Arg Phe Glu Glu Leu Arg Thr Thr Leu Pro His Thr Phe Thr Gly		
	245	250 255
Phe Glu Tyr Asp Phe Glu Tyr Val Trp Ala Asp Leu Ala Pro Glu Gln		
	260	265 270
Arg Arg Glu Val Leu Glu Asn Ile Tyr Glu Tyr Gly Ser Leu Lys Leu		
	275	280 285
Trp Leu Ser Ser Phe Ala Glu Met Phe Phe Asp Glu Gln Val Ser Asp		
	290	295 300
Glu Ile Ser Glu Phe Val Arg Glu Lys Met Arg Ala Arg Leu Ile Asp		
305	310	315 320
Pro Glu Leu Cys Asp Leu Leu Ile Pro Thr Asp Tyr Gly Phe Gly Thr		
	325	330 335
His Arg Val Pro Leu Glu Thr Asn Tyr Leu Glu Val Tyr His Arg Pro		
	340	345 350
Asn Val Thr Ala Ile Gly Val Lys Asn Asn Pro Ile Ala Arg Ile Val		
	355	360 365
Pro Gln Gly Ile Glu Leu Thr Asp Gly Thr Phe His Glu Leu Asp Val		
	370	375 380
Ile Ile Leu Ala Thr Gly Phe Asp Ala Gly Thr Gly Ala Leu Thr Arg		
385	390	395 400
Ile Asp Ile Arg Gly Arg Gly Gly Arg Ser Leu Lys Glu Asp Trp Gly		
	405	410 415
Arg Asp Ile Arg Thr Thr Met Gly Leu Met Val His Gly Tyr Pro Asn		
	420	425 430
Met Leu Thr Thr Ala Val Pro Leu Ala Pro Ser Ala Ala Leu Cys Asn		
	435	440 445
Met Thr Thr Cys Leu Gln Gln Gln Thr Glu Trp Ile Ser Glu Ala Ile		
450	455	460

Arg Tyr Met Gln Glu Arg Asp Leu Thr Val Ile Glu Pro Thr Lys Glu  
465 470 475 480

Ala Glu Asp Ala Trp Val Ala His His Asp Glu Thr Ala Ala Val Asn  
485 490 495

Leu Ile Ser Lys Thr Asp Ser Trp Tyr Val Gly Ser Asn Val Pro Gly  
500 505 510

Lys Pro Arg Arg Val Leu Ser Tyr Thr Gly Gly Val Gly Ala Tyr Arg  
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Glu Lys Ala Gln Glu Ile Ala Asp Ala Gly Tyr Lys Gly Phe Asn Leu  
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Arg  
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Leu Thr Val Val Gly Phe Asp Ala Ala Asp Gly Pro Gly Gly Thr Trp  
35 40 45

Tyr Trp Asn Arg Tyr Pro Gly Ala Leu Ser Asp Thr Glu Ser His Val  
50 55 60

Tyr Arg Phe Ser Phe Asp Glu Asp Leu Leu Gln Asp Trp Thr Trp Lys  
65 70 75 80

Glu	Thr	Tyr	Pro	Thr	Gln	Pro	Glu	Ile	Leu	Glu	Tyr	Leu	Glu	Asp	Val
				85					90					95	
Val	Asp	Arg	Phe	Asp	Leu	Arg	Arg	Asp	Phe	Arg	Phe	Gly	Thr	Glu	Val
			100					105					110		
Thr	Ser	Ala	Thr	Tyr	Leu	Glu	Asp	Glu	Asn	Leu	Trp	Glu	Val	Thr	Thr
		115					120					125			
Asp	Gly	Gly	Glu	Val	Tyr	Arg	Ala	Arg	Phe	Val	Val	Asn	Ala	Val	Gly
	130					135					140				
Leu	Leu	Ser	Ala	Ile	Asn	Phe	Pro	Asn	Ile	Pro	Gly	Leu	Asp	Thr	Phe
145					150					155					160
Glu	Gly	Glu	Thr	Ile	His	Thr	Ala	Ala	Trp	Pro	Glu	Gly	Val	Asp	Leu
				165					170					175	
Thr	Gly	Lys	Arg	Val	Gly	Val	Ile	Gly	Thr	Gly	Ser	Thr	Gly	Ile	Gln
			180					185					190		
Val	Ile	Thr	Ala	Leu	Ala	Pro	Glu	Val	Glu	His	Leu	Thr	Val	Phe	Val
		195					200					205			
Arg	Thr	Pro	Gln	Tyr	Ser	Val	Pro	Val	Gly	Asn	Arg	Pro	Val	Thr	Ala
	210					215					220				
Glu	Gln	Ile	Asp	Ala	Ile	Lys	Ala	Asp	Tyr	Asp	Glu	Ile	Trp	Ala	Gln
225					230					235					240
Val	Lys	Arg	Ser	Gly	Val	Ala	Phe	Gly	Phe	Glu	Glu	Ser	Thr	Val	Pro
				245					250					255	
Ala	Met	Ser	Val	Ser	Glu	Glu	Glu	Arg	Asn	Arg	Val	Phe	Glu	Glu	Ala
			260					265					270		
Trp	Glu	Glu	Gly	Gly	Gly	Phe	Arg	Phe	Met	Phe	Gly	Thr	Phe	Gly	Asp
		275					280					285			
Ile	Ala	Thr	Asp	Glu	Ala	Ala	Asn	Glu	Thr	Ala	Ala	Ser	Phe	Ile	Arg
						295					300				
Ser	Lys	Ile	Arg	Glu	Ile	Val	Lys	Asp	Pro	Glu	Thr	Ala	Arg	Lys	Leu
305					310					315					320



Thr Pro Thr Gly Leu Phe Ala Arg Arg Arg Leu Cys Asp Asp Gly Tyr  
                   325                                  330                                  335

Tyr Glu Val Tyr Asn Arg Pro Asn Val Glu Ala Val Asp Ile Lys Glu  
                   340                                  345                                  350

Asn Pro Ile Arg Glu Ile Thr Ala Lys Gly Val Val Thr Glu Asp Gly  
                   355                                  360                                  365

Val Leu His Glu Leu Asp Val Leu Val Phe Ala Thr Gly Phe Asp Ala  
                   370                                  375                                  380

Val Asp Gly Asn Tyr Arg Arg Ile Asp Ile Arg Gly Arg Gly Gly Leu  
                   385                                  390                                  395                                  400

Ser Leu Asn Asp His Trp Asp Gly Gln Pro Thr Ser Tyr Leu Gly Leu  
                                   405                                  410                                  415

Ser Thr Ala Gly Phe Pro Asn Trp Phe Met Val Leu Gly Pro Asn Gly  
                   420                                  425                                  430

Pro Phe Thr Asn Leu Pro Pro Ser Ile Glu Thr Gln Val Glu Trp Ile  
                   435                                  440                                  445

Ser Asp Thr Ile Ala Tyr Ala Glu Glu Asn Gly Ile Arg Ala Ile Glu  
                   450                                  455                                  460

Pro Thr Pro Glu Ala Glu Asp Glu Trp Thr Ala Thr Cys Thr Asp Ile  
                   465                                  470                                  475                                  480

Ala Asn Ala Thr Leu Phe Thr Lys Ala Asp Ser Trp Ile Phe Gly Ala  
                                   485                                  490                                  495

Asn Val Pro Gly Lys Lys Pro Ser Val Leu Phe Tyr Leu Gly Gly Leu  
                                   500                                  505                                  510

Gly Asn Tyr Arg Ala Val Leu Ala Asp Val Ala Ala Ala Gly Tyr Arg  
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Gly Phe Ala Leu Lys Ser Ala Asp Ala Val Thr Ala  
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Gly Phe Ala Gly Ile Gly Ala Ala Val Glu Leu Lys Arg Xaa Gly Ile  
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Asp Asp Phe Val Leu Leu Glu Arg Ala Asp Asp Val Gly Gly Thr Trp  
 35 40 45

Arg Asp Asn Thr Tyr Pro Gly Ala Ala Cys Asp Val Pro Ser Xaa Leu  
 50 55 60

Tyr Ser Tyr Ser Phe Ala Pro Asn Pro Asn Trp Thr Arg Leu Phe Ala  
 65 70 75 80

Xaa Gln Pro Glu Ile Tyr Asp Tyr Leu Glu Asp Val Ala Ala Xaa Xaa  
 85 90 95

Gly Leu Xaa Xaa His Val Arg Phe Gly Val Glu Val Thr Glu Ala Arg  
 100 105 110

Trp Asp Glu Ser Ala Gln Leu Trp Arg Val Xaa Thr Ala Ser Gly Glu  
 115 120 125

Leu Thr Ala Xaa Phe Leu Val Ala Ala Thr Gly Pro Leu Ser Xaa Pro  
 130 135 140

Lys Ile Pro Asp Leu Pro Gly Leu Glu Ser Phe Glu Gly Xaa Xaa Phe  
 145 150 155 160

His Ser Ala Xaa Trp Asn His Asp Leu Asp Leu Arg Gly Glu Arg Val  
 165 170 175

Ala Val Val Gly Thr Gly Ala Ser Ala Val Gln Phe Val Pro Glu Ile  
 180 185 190

Ala Asp Xaa Ala Xaa Thr Leu Thr Val Phe Gln Arg Thr Pro Gln Trp

195	200	205
Val Leu Pro Arg Pro Asp Xaa Thr Leu Pro Xaa Ala Xaa Arg Ala Val		
210	215	220
Phe Ser Arg Val Pro Gly Thr Gln Lys Trp Leu Arg Xaa Arg Leu Tyr		
225	230	235
Gly Ile Phe Glu Ala Leu Gly Ser Gly Phe Val Xaa Pro Xaa Trp Leu		
245	250	255
Leu Pro Xaa Xaa Xaa Ala Leu Ala Arg Ala His Leu Arg Arg Gln Val		
260	265	270
Arg Asp Pro Glu Leu Arg Xaa Lys Leu Thr Pro Asp Tyr Thr Pro Gly		
275	280	285
Cys Lys Arg Met Leu Leu Ser Asn Asp Trp Tyr Pro Ala Leu Xaa Lys		
290	295	300
Pro Asn Val Ser Leu Val Thr Ser Gly Val Val Glu Val Thr Glu Xaa		
305	310	315
Gly Val Val Asp Ala Asp Gly Val Glu His Glu Val Asp Thr Ile Ile		
325	330	335
Phe Ala Thr Gly Phe His Xaa Thr Asp Xaa Pro Xaa Ala Met Lys Ile		
340	345	350
Phe Gly Arg Glu Gly Arg Ser Leu Ala Asp His Trp Asn Gly Ser Ala		
355	360	365
Xaa Ala Tyr Leu Gly Thr Ala Val Ser Gly Phe Pro Asn Leu Phe Xaa		
370	375	380
Leu Leu Gly Pro Asn Thr Gly Leu Gly His Thr Ser Ile Val Xaa Ile		
385	390	395
Leu Glu Ala Gln Ala Glu Tyr Ile Ala Ser Ala Leu Xaa Xaa Met Arg		
405	410	415
Arg Glu Gly Leu Gly Ala Leu Asp Val Arg Ala Glu Val Gln Xaa Xaa		
420	425	430
Phe Asn Xaa Ala Val Gln Glu Arg Leu Ala Thr Thr Val Trp Asn Ala		
435	440	445

Gly Gly Cys Ser Ser Trp Tyr Xaa Asp Pro Asp Gly Arg Asn Ser Thr  
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Xaa Trp Pro Trp Ser Thr Xaa Xaa Phe Arg Ala Arg Thr Arg Arg Phe  
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Asp Pro Ser Asp Tyr Xaa Pro Ser Ser Pro Thr Pro Glu Thr Xaa Xaa  
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Gly

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Gly Ile Gly Ala Ala Xaa Arg Leu Xaa Arg Glu Xaa Gly Ile Xaa Phe  
 20 25 30

Ala Ile Leu Glu Ala Arg Asp Asn Val Gly Gly Thr Trp Asp Leu Phe  
 35 40 45

Asn Tyr Pro Gly Ile Arg Ser Asp Ser Asp His Leu Thr Xaa Gly Lys  
 50 55 60

Gly Ala Phe Arg Pro Phe Pro Xaa Ala Lys Xaa Leu Ala Asp Gly Pro

65	70	75	80
Ser His Glu Leu Xaa Xaa Tyr Val Arg Asp Thr Ala Xaa Glu Xaa Gly	85	90	95
Leu Arg Xaa His Ile Xaa Phe Gly Thr Lys Val Val Ala Ala Xaa Xaa	100	105	110
Xaa Ala Xaa Ser Leu Trp Thr Val Thr Val Xaa Xaa Xaa Gly Glu Thr	115	120	125
Glu Val Xaa Thr Tyr Asn Val Leu Xaa Xaa Ala Asn Gly Tyr Tyr Ser	130	135	140
Tyr Asp Lys Gly Asn Ile Pro Asp Phe Pro Gly Glu Phe Xaa Gly Xaa	145	150	155
Leu Val His Pro Gln Xaa Tyr Pro Glu Xaa Leu Asp Tyr Arg Gly Lys	165	170	175
Lys Val Val Val Ile Gly Ser Gly Ala Ser Gly Xaa Thr Leu Ala Pro	180	185	190
Xaa Met Xaa Xaa Xaa Ala Xaa His Val Thr Met Leu Gln Arg Ser Gly	195	200	205
Thr Tyr Ile Ala Leu Pro Ser Asp Ala Val Val Pro Xaa Gln Leu Ala	210	215	220
Gly Xaa Arg Xaa Xaa Xaa Xaa Xaa Leu Gln Xaa Xaa Gln Leu Arg Xaa	225	230	235
Pro Pro Trp Xaa Ala Lys Arg Leu Xaa Leu Leu Leu Ile Arg Arg Gln	245	250	255
Leu Gly Lys Asn Val Xaa Leu Xaa Gly Phe Pro Thr Pro Ser Tyr Xaa	260	265	270
Pro Trp Asp Gln His Leu Cys Val Val Pro Asn Gly Asp Leu Leu Lys	275	280	285
Xaa Leu Gly Ser Gly Asp Ala Xaa Ile Xaa Thr Asp Ile Asp Thr Phe	290	295	300
Thr Gly Lys Gly Val Xaa Phe Ala Ser Gly Arg Glu Xaa Asp Ala Asp	305	310	315
			320



Val Val Val Thr Ala Thr Gly Leu Asn Xaa Xaa Xaa Gly Gly Pro Phe  
                           325                          330                          335

Ile Xaa Xaa Asp Gly Leu Leu Val Asp Leu Xaa Xaa Arg Xaa Ala Leu  
                           340                          345                          350

Phe Tyr Lys Xaa Xaa Xaa Xaa Ser Asp Asn Leu Asn Phe Leu Gly Xaa  
                           355                          360                          365

Val Gly Tyr Thr Asn Ala Ser Trp Thr Leu Arg Ala Asp Leu Ala Xaa  
                           370                          375                          380

Leu Val Ala Cys Arg Leu Leu Xaa Xaa Met Xaa Xaa Arg Ser Ala Xaa  
                           385                          390                          395                          400

Xaa Xaa Xaa Xaa His Ala Xaa Ala Glu Xaa Xaa Xaa Xaa Leu Leu Ala  
                           405                          410                          415

Ser Gly Tyr Lys Xaa Arg Xaa Xaa Gly Xaa Met Pro Xaa Gln Gly Xaa  
                           420                          425                          430

Lys Xaa Xaa Trp Xaa Xaa Xaa Xaa Asn Tyr Xaa Xaa Asp Arg Xaa Leu  
                           435                          440                          445

Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Phe Ser Lys Xaa  
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